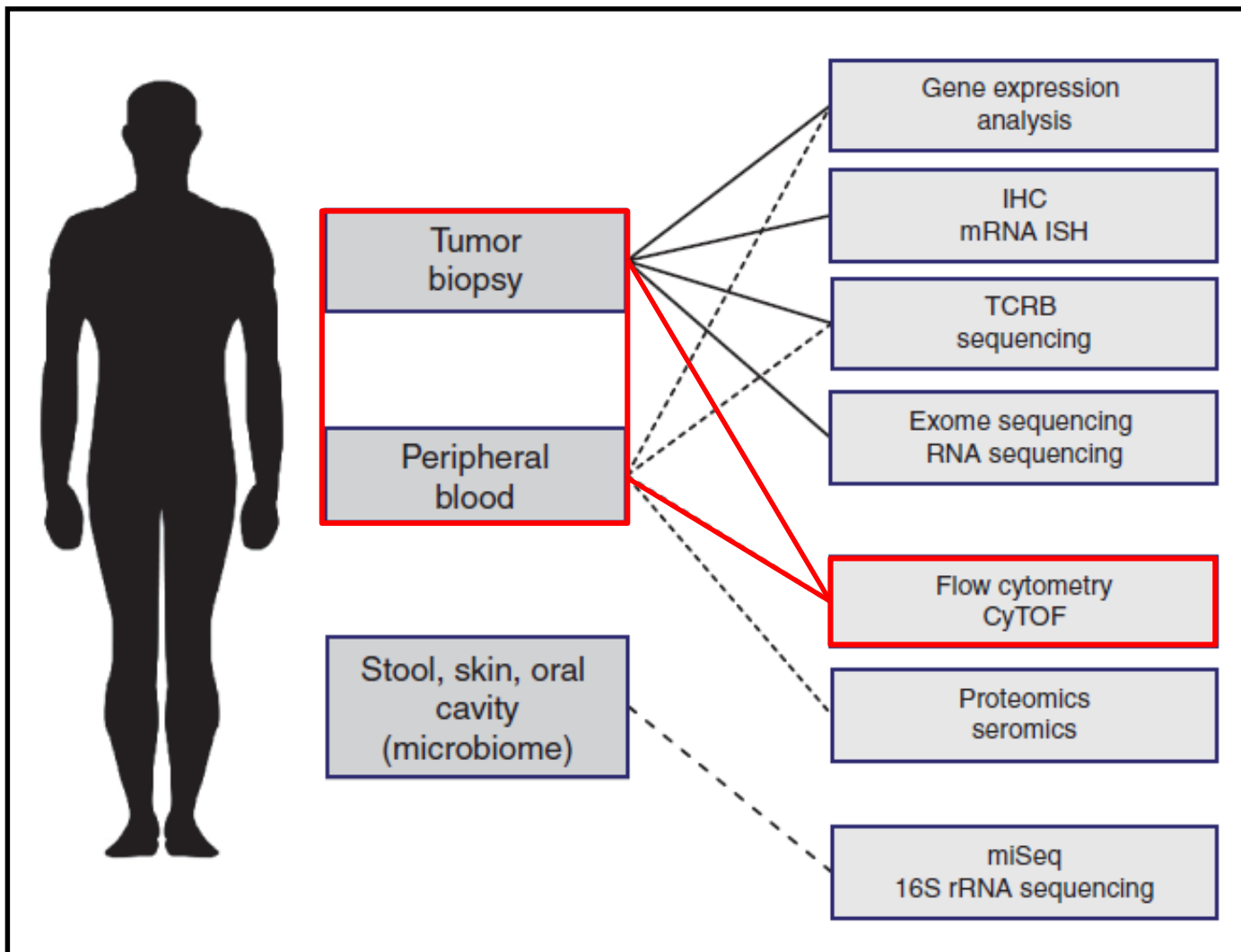


Flow Cytometry in Clinical Diagnosis

Ourania Tsitsilonis, MD, PhD

Flow Cytometry Unit, Department of Biology, NKUA

Platforms for translational research in immuno-oncology



Collaborators

FC Unit / Dept Biology

- Dr. Ioannis Kostopoulos
- Dr. Kostas Papadimitriou
- Pantelis Rousakis, MSc
- Crysanthi Panteli, MSc
- Nikos Angelis, MSc
- Georgia Dimitrakopoulou, MSc
- Panagiotis Bakouros, MSc
- Nikos Tsakirakis, MSc
- post and undergrads

University of Navarra

- Dr. Bruno Paiva

Therapeutic Clinic

- Prof. Meletios A. Dimopoulos
- Prof. Evangelos Terpos
- Prof. Efsthios Kastritis
- As. Prof. Maria Gavriatopoulou
- Faculty members
- MDs, interns, co-workers

University of Tuebingen

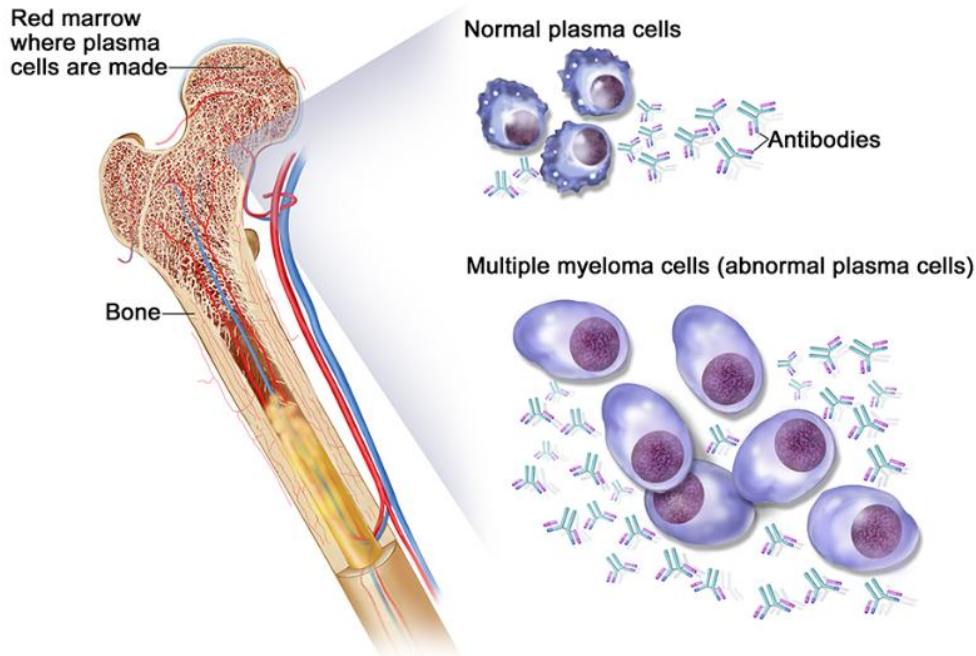
- Prof. Wolfgang Voelter 
- Prof. Hubert Kalbacher

Theagenion Cancer Hospital

- Dr. Eirini Katodritou

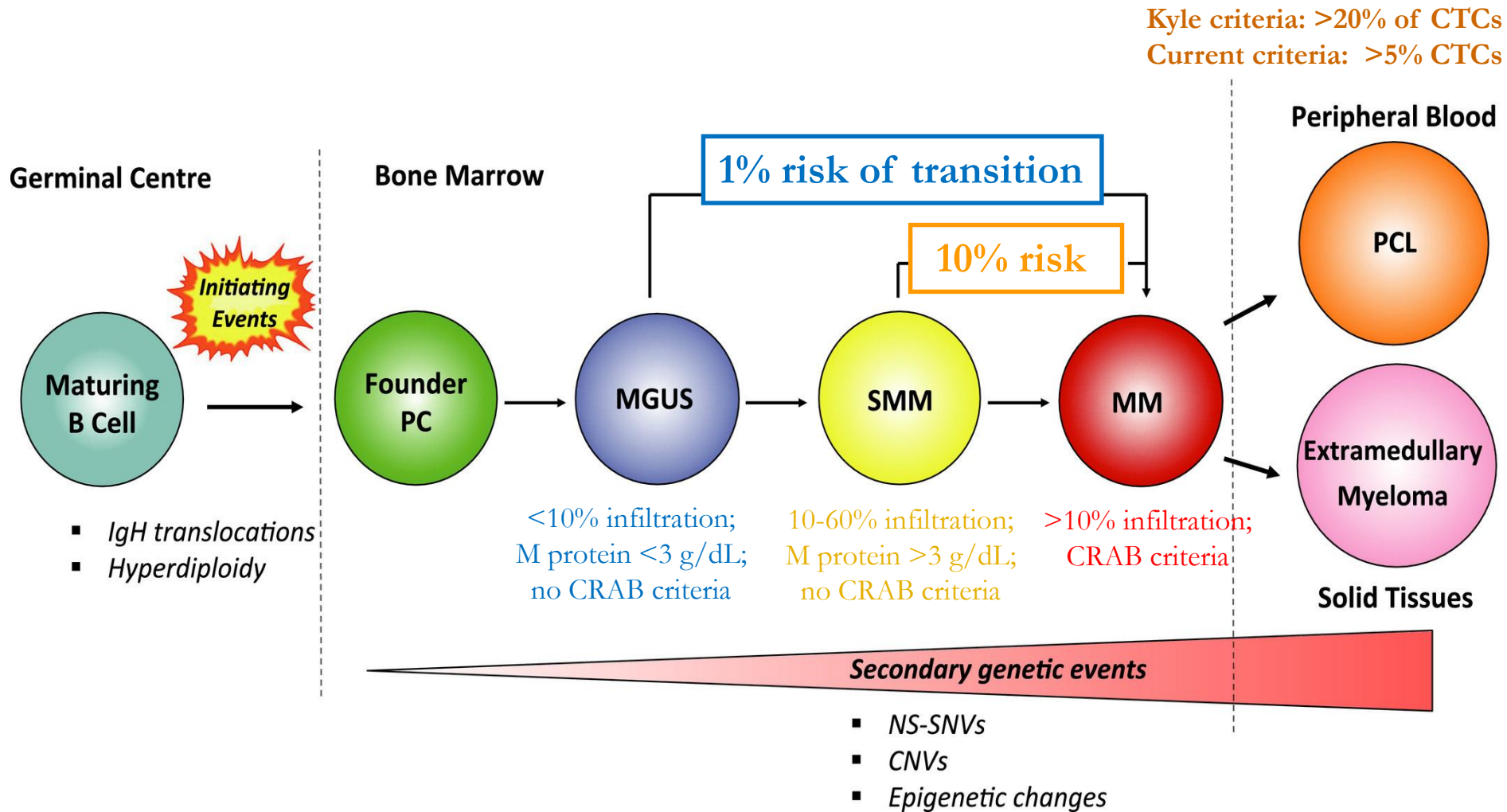
Multiple Myeloma

Multiple Myeloma

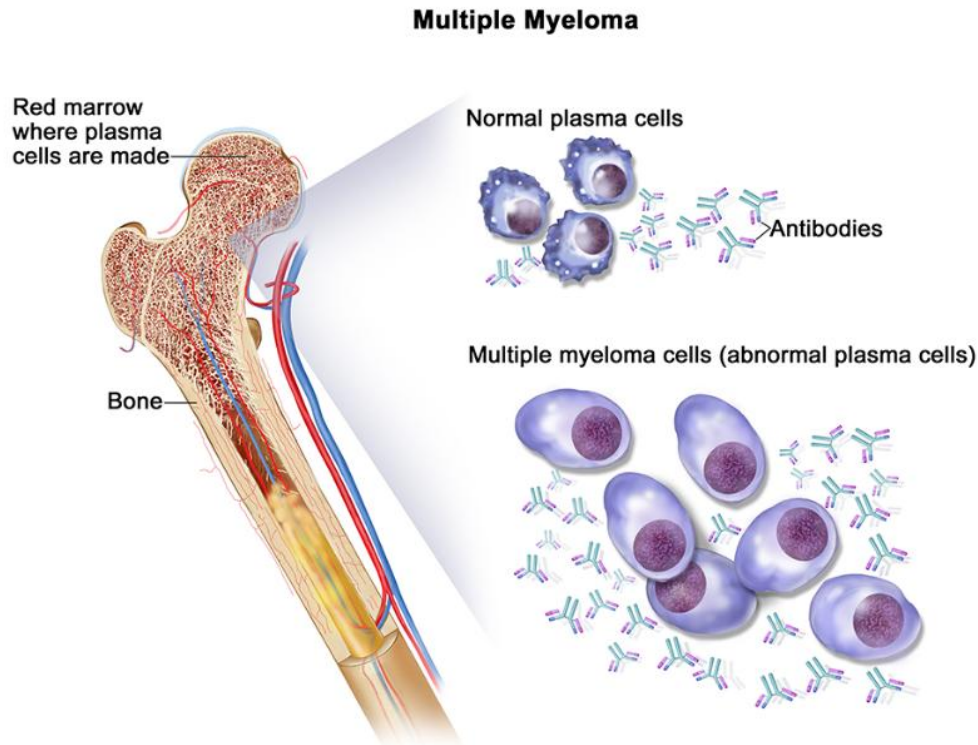


- **2nd most common hematologic malignancy** (0,7% probability in general population)
- Median age at diagnosis: 63 years
- 600-800 new cases per year in Greece
- Clonal plasma cells accumulate in the bone marrow (**>10% in BM biopsy**) where they interact with multiple cell types and massively proliferate
- Excess production of monoclonal immunoglobulin (M-protein)
- **Incurable disease:** Patients achieve complete remissions but finally relapse and succumb due to chemoresistance and end-stage disease

Evolution of MM



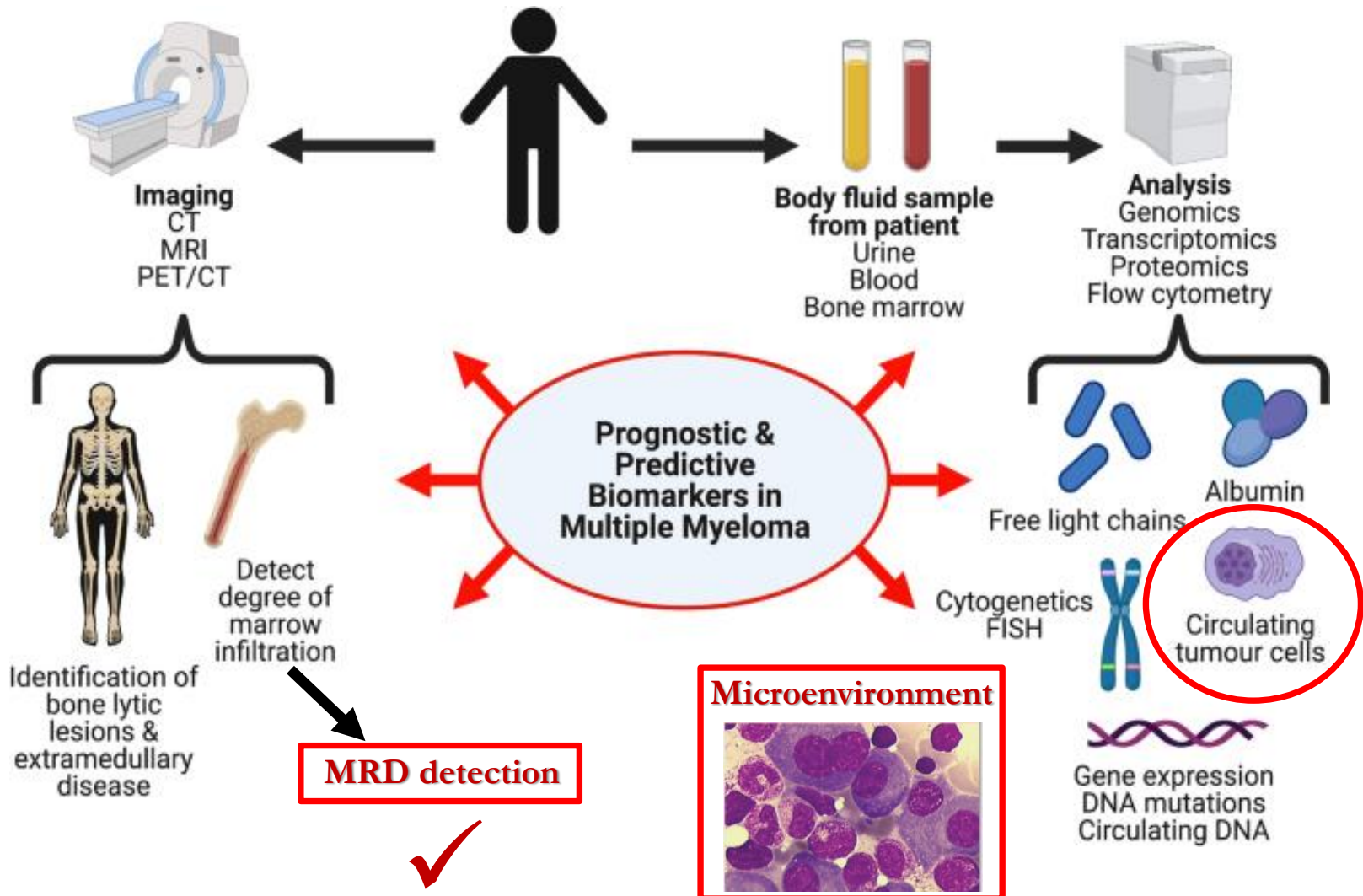
MM is a highly heterogeneous disease



Multi-level heterogeneity

- **Primary cytogenetic initiating events**
- **Secondary genetic events**
 - cytogenetic aberrations
 - gene mutations
- **Epigenetic changes**
 - chromatic modification
 - gene expression profiling
- **Phenotype** of clonal cells
- **Tumor burden**
- **Presence of multiple foci**

Need for identifying novel predictive/prognostic biomarkers

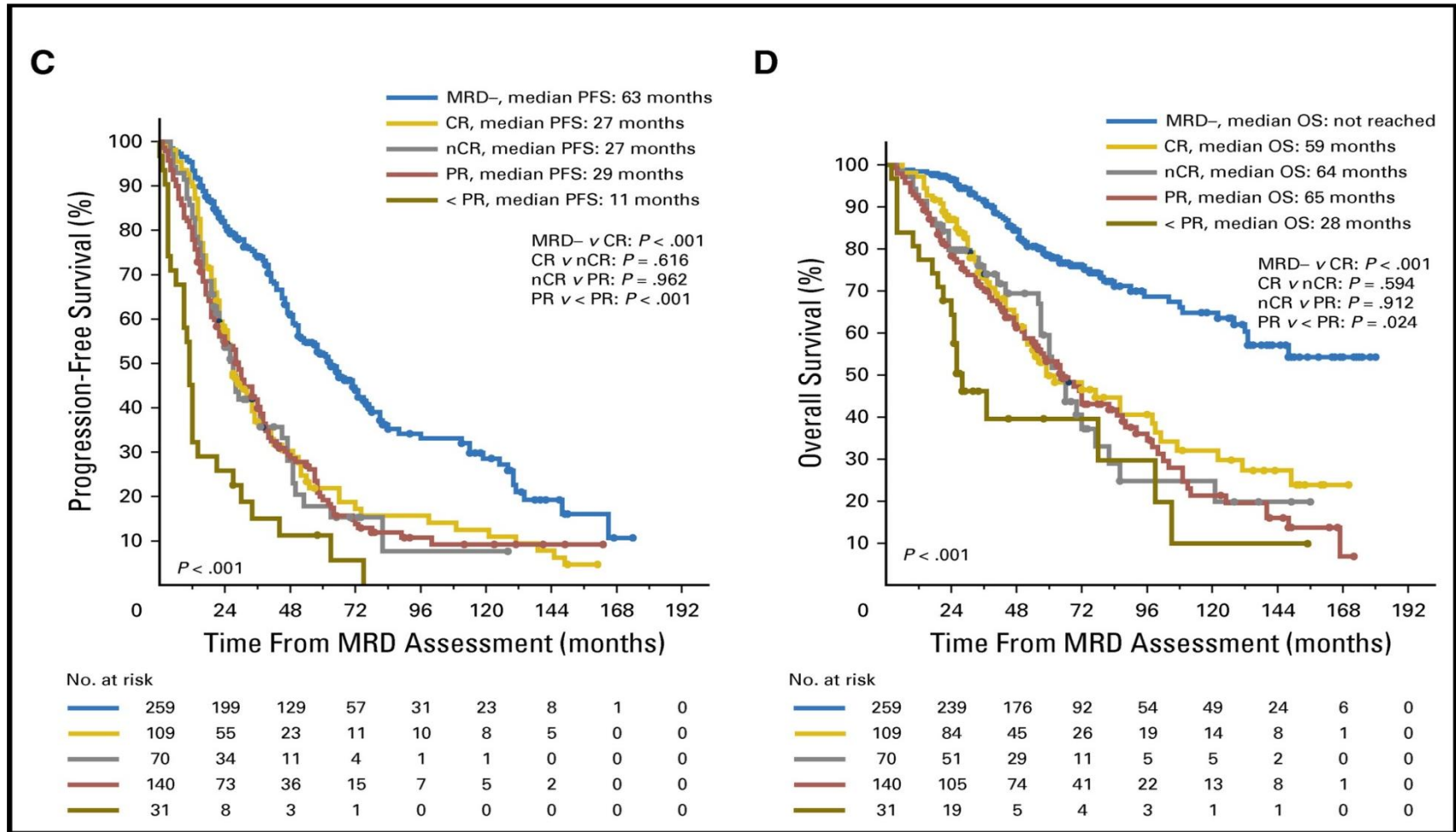


Definition of Minimal Residual Disease

+ **Disease** ~ pathology
+ **Residual** ~ after treatment
Minimal ~ low number of cells

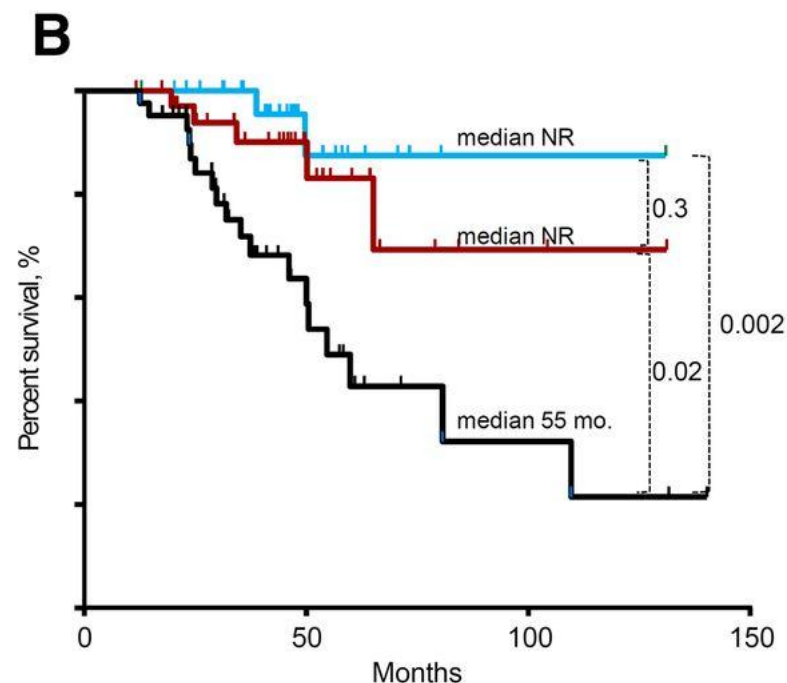
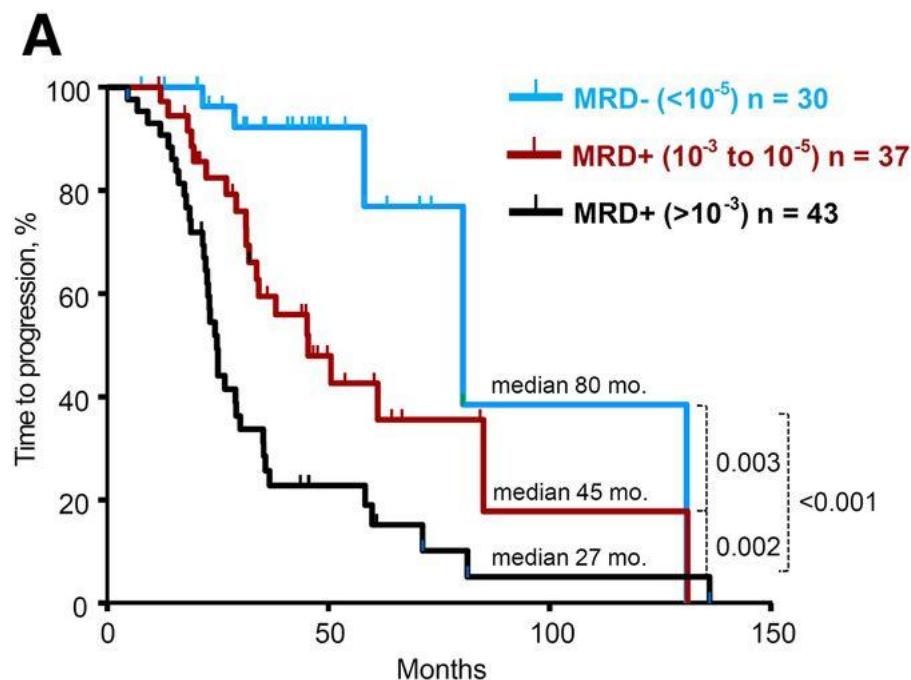
Wiki: MRD is the name given to small numbers of cancer cells that remain in the patient during or after treatment when the patient is in remission (no symptoms or signs of disease) and is the major cause of relapse in cancer.

PFS & OS of CR without MRD-negativity (MFC) are similar to PR



Sensitivity in MRD matters

Prognostic value of deep sequencing method for minimal residual disease detection in multiple myeloma



10^{-5} = target for definition of MRD negativity

2016: revised IMWG response criteria

THE LANCET
Oncology

Volume 17, Issue 8, August 2016, Pages e328-e346

International Myeloma Working Group consensus criteria for response and minimal residual disease assessment in multiple myeloma

Shaji Kumar, Bruno Paiva, Kenneth C Anderson, Brian Durie, Ola Landgren, Philippe Moreau, Nikhil Munshi, Sagar Lonial, Joan Bladé, Maria-Victoria Mateos, Meletios Dimopoulos, Efsthios Kastritis, Mario Boccadoro, Robert Orlowski, Hartmut Goldschmidt, Andrew Spencer, Jian Hou, Wee Joo Chng, Saad Z Usmani, Elena Zamagni, Kazuyuki Shimizu, Sundar Jagannath, Hans E Johnsen, Evangelos Terpos, Anthony Reiman, Robert A Kyle, Pieter Sonneveld, Paul G Richardson, Philip McCarthy, Heinz Ludwig, Wenming Chen, Michele Cavo, Jean-Luc Harousseau, Suzanne Lentzsch, Jens Hillengass, Antonio Palumbo, Alberto Orfao, S Vincent Rajkumar, Jesus San Miguel, Herve Avet-Loiseau

Response criteria*

IMWG MRD criteria (requires a complete response as defined below)

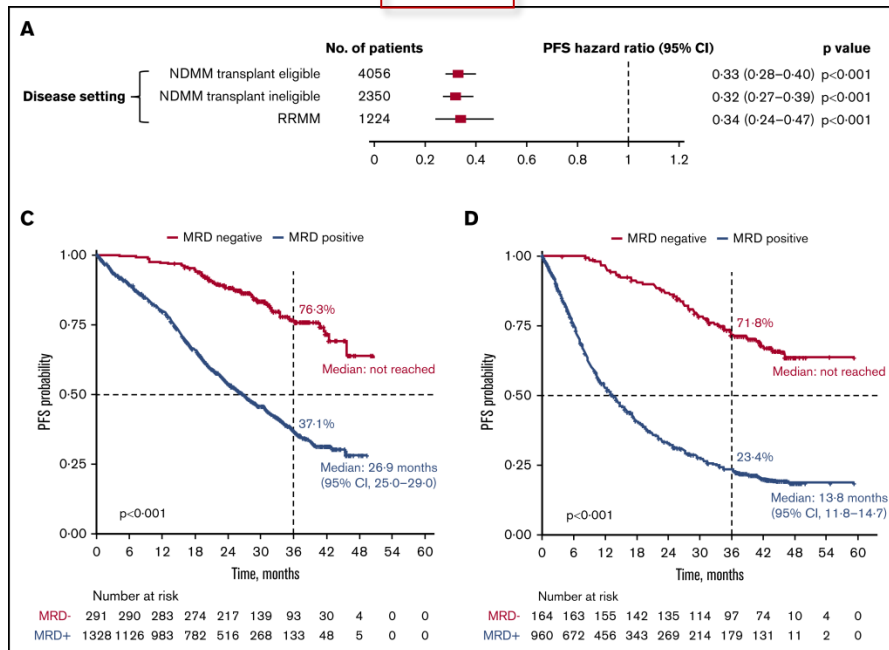
Sustained MRD-negative	MRD negativity in the marrow (NGF or NGS, or both) and by imaging as defined below, confirmed minimum of 1 year apart. Subsequent evaluations can be used to further specify the duration of negativity (eg, MRD-negative at 5 years)†
Flow MRD-negative	Absence of phenotypically aberrant clonal plasma cells by NGF‡ on bone marrow aspirates using the EuroFlow standard operation procedure for MRD detection in multiple myeloma (or validated equivalent method) with a minimum sensitivity of 1 in 10 ⁵ nucleated cells or higher
Sequencing MRD-negative	Absence of clonal plasma cells by NGS on bone marrow aspirate in which presence of a clone is defined as less than two identical sequencing reads obtained after DNA sequencing of bone marrow aspirates using the LymphoSIGHT platform (or validated equivalent method) with a minimum sensitivity of 1 in 10 ⁵ nucleated cells§ or higher
Imaging plus MRD-negative	MRD negativity as defined by NGF or NGS plus disappearance of every area of increased tracer uptake found at baseline or a preceding PET/CT or decrease to less than that of surrounding normal tissue¶

For MRD assessment, the first bone marrow aspirate should be sent to MRD (not for morphology) and this sample should be taken in one draw with a volume of minimally 2 mL (to obtain sufficient cells), but maximally 4–5 mL to avoid haemodilution.

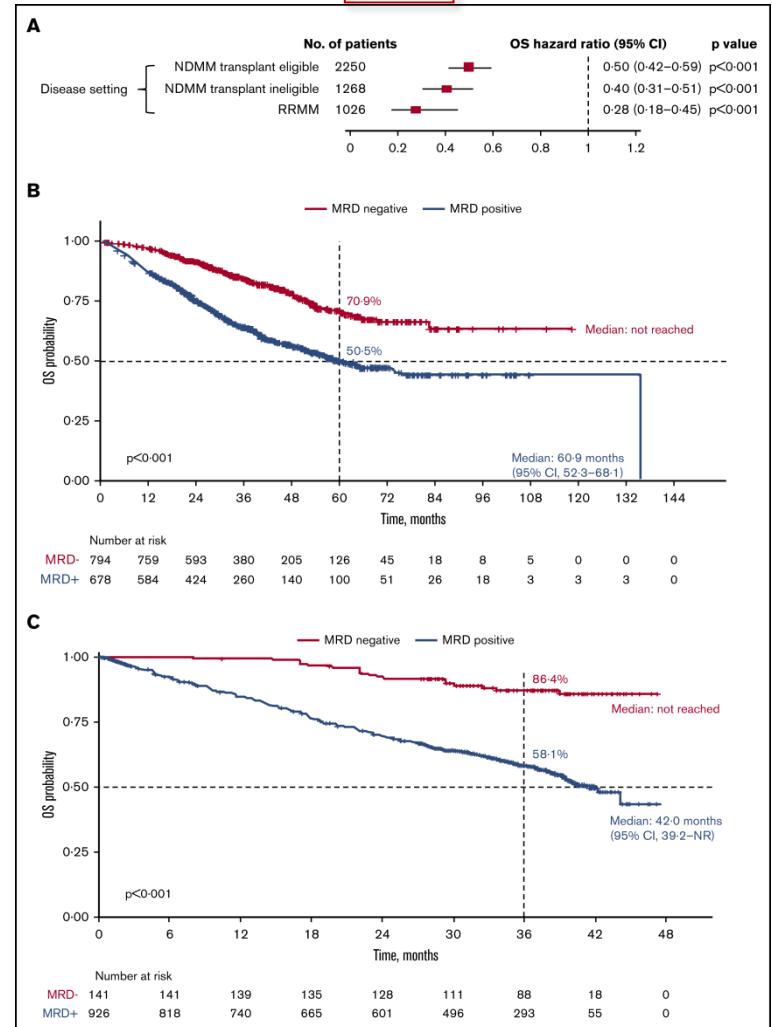
Role of MRD-negativity in MM: a clinically valid surrogate biomarker for PFS & OS

- Meta-analysis >8,000 pts
- MRD sensitivity 10^{-4} - 10^{-6}
- MRD-negative increased PFS & OS in NDMM & RRMM
- Confirmed robust association of MRD status and survival outcomes

PFS



OS



Looking for a needle in a haystack !

- There is a pressing need of highly sensitive methods for MRD monitoring



Techniques available to assess MRD in MM

	NGF (≥ 8 colors)	NGS	MRI/PET-CT
Diagnostic sample	Important but not mandatory	Mandatory	Important but not mandatory
Fresh sample/Viable cells	Needed (<36 h)	Not needed	n/a
Patchy sample	Impacts	Impacts	No impact
Hemodiluted aspirate	Requires sample quality control	Impacts ?	n/a
Time	2 hours	7 days	2 hours
Global standardization	Yes	No	No
Quantitative	Yes	Yes	Yes
Sensitivity	$10^{-5} - 10^{-6}$	10^{-6}	High ?
Applicability	~100%	~90%	~100%
Global cell characterization	Yes	No	No
Clonal heterogeneity	Can not be identified	Identified	n/a
Standardization	Ongoing (EuroFlow)	yes	?
Reproducibility among centers	High	Not reported	Moderate at MRD
Availability	High	Increasing	Intermediate
Cost	~250 \$	~700 \$	800-2000 \$

Modified from Paiva et al., 2015; Flores-Montero et al., 2017

The 5 Steps of Next Generation Flow Cytometry (NGF)

Cell/Sample Preparation



Staining with labeled Abs



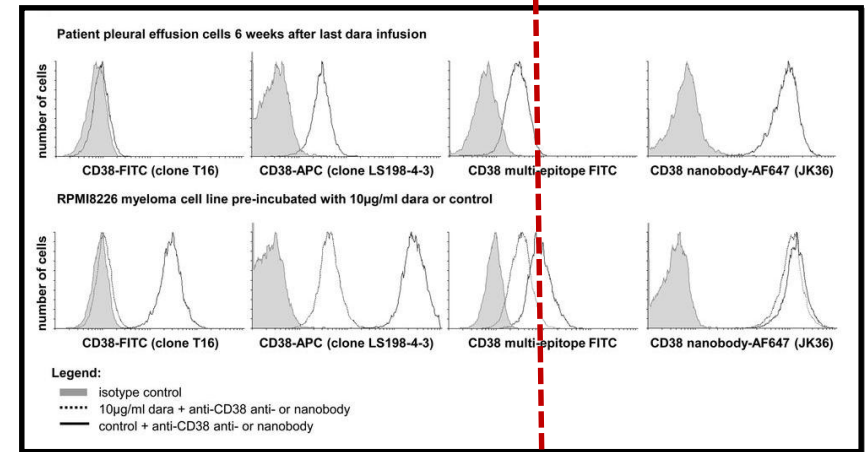
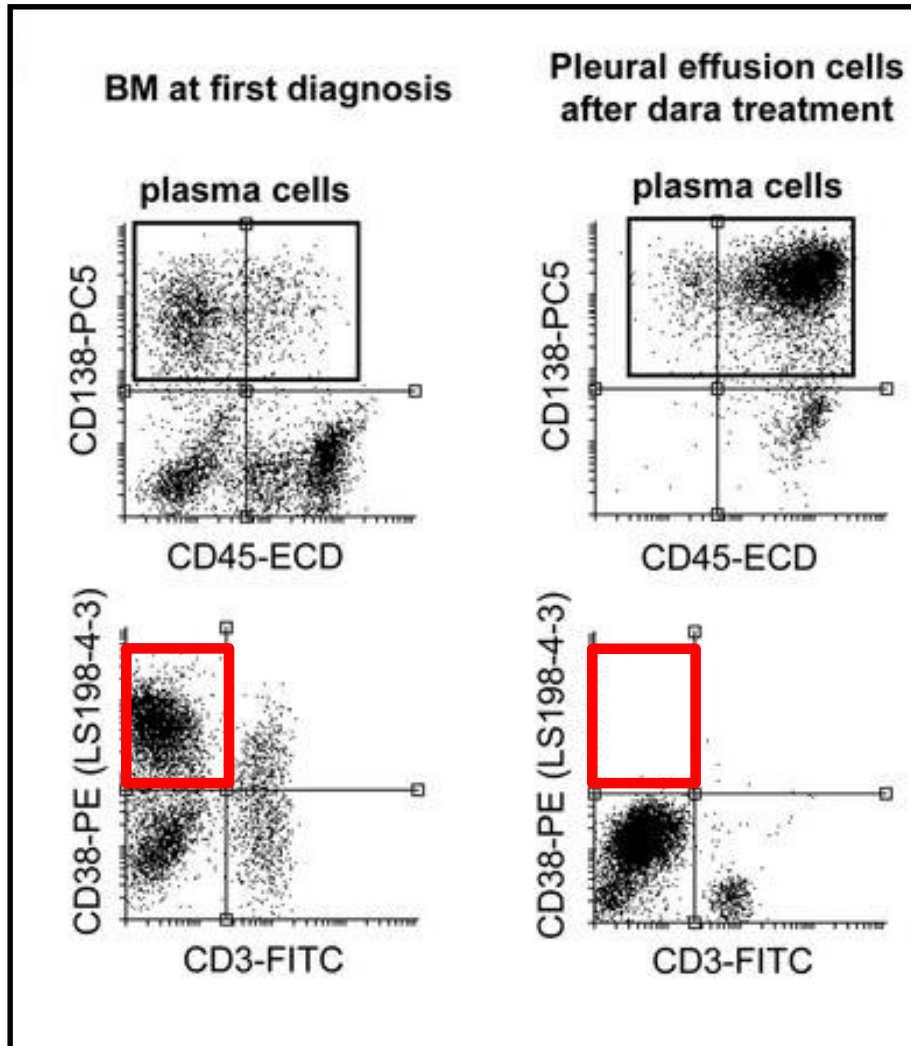
Cell Acquisition

Cell Analysis

Data Interpretation

Therapeutic anti-CD38 impact on MRD

MM patients treated with Daratumumab, Isatuximab, TAK079



The 5 Steps of Next Generation Flow Cytometry (NGF)

Cell/Sample Preparation

Staining with labeled Abs

Cell Acquisition

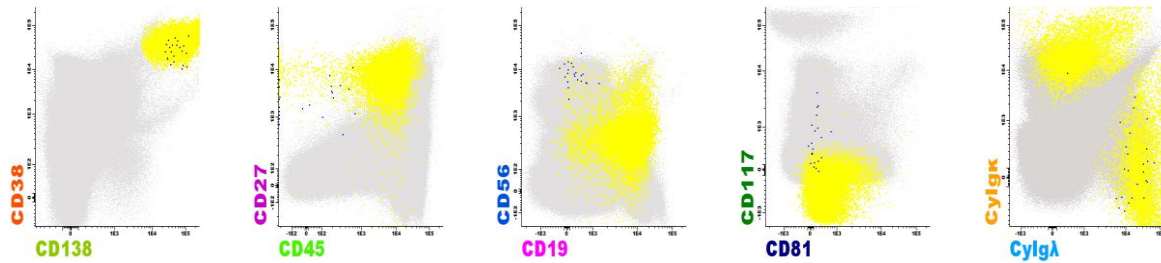


Cell Analysis

Data Interpretation

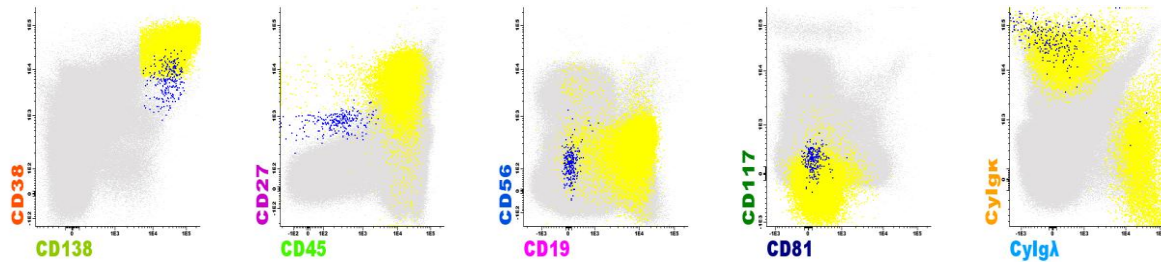
Sensitivity of MRD assessment by NGF

10^{-6}



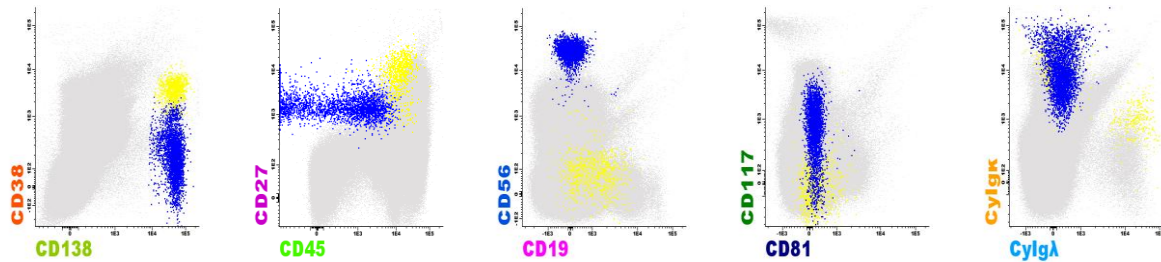
CD38^{dim/+}CD138⁺⁺CD45⁻
CD27^{dim}CD56⁺
CD19⁻CD117^{dim/-}CD81⁻ lambda

10^{-5}



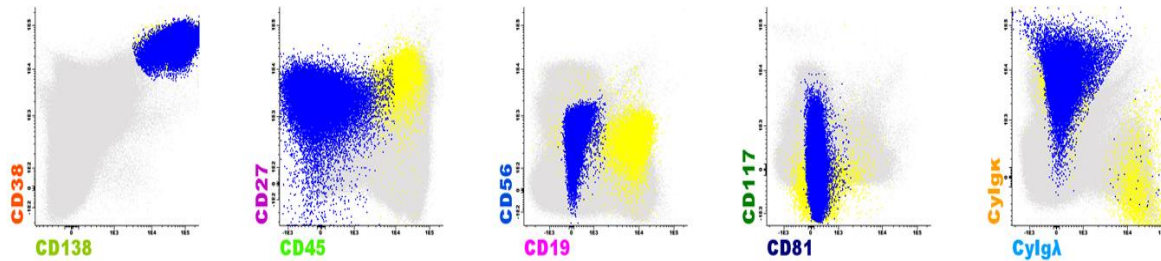
CD38^{dim}CD138⁺CD45⁻
CD27^{dim}CD56⁻
CD19⁻CD117⁻CD81⁻ kappa

10^{-4}



CD38^{dim/-}CD138⁺⁺CD45⁻
CD27^{dim}CD56⁺⁺
CD19⁻CD117^{dim/-}CD81⁻ kappa

10^{-3}



CD38⁺CD138⁺⁺CD45⁻
CD27^{dim/-}CD56⁻
CD19⁻CD117^{dim/-}CD81⁻ kappa

The 5 Steps of Next Generation Flow Cytometry (NGF)

Cell/Sample Preparation

Staining with labeled Abs

Cell Acquisition

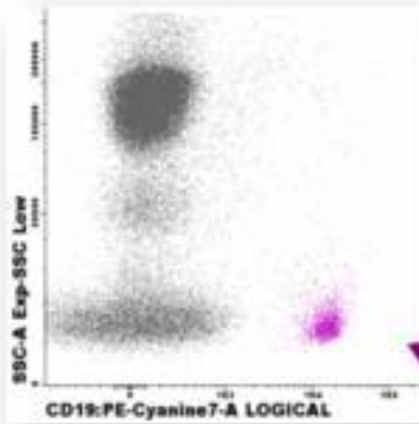
Cell Analysis

Data Interpretation

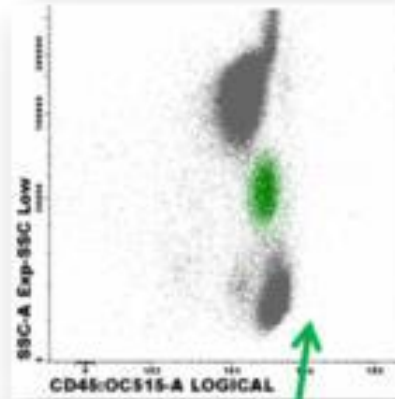


APS: automatic population separation

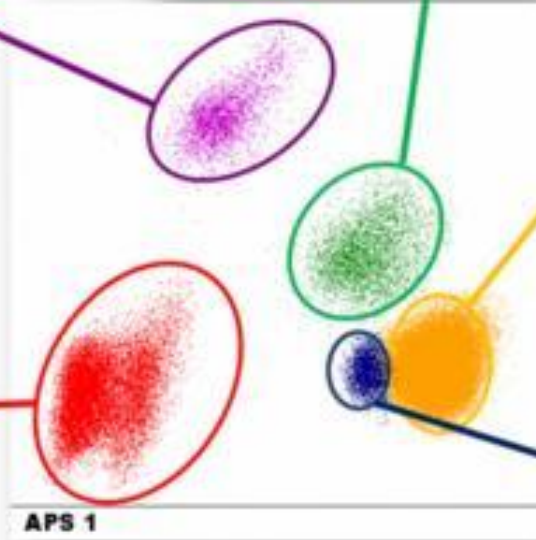
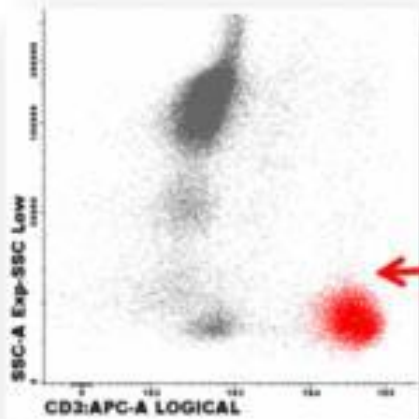
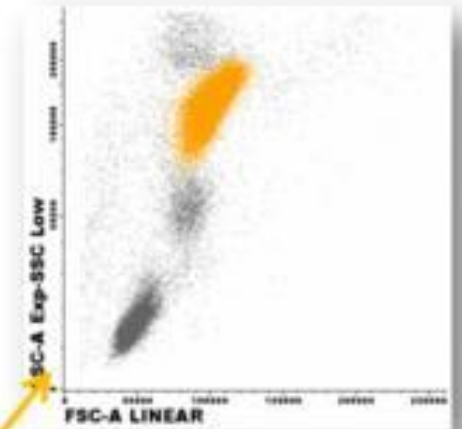
B-cells



Monocytes

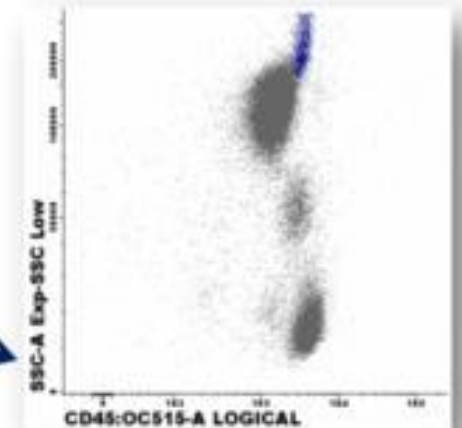


Neutrophils



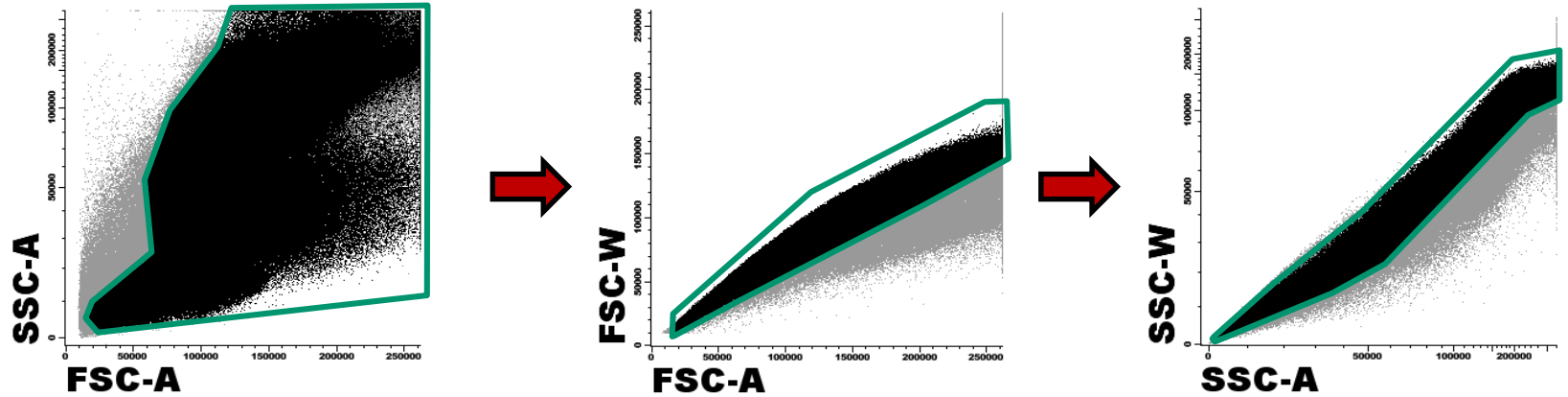
T-cells

Eosinophils

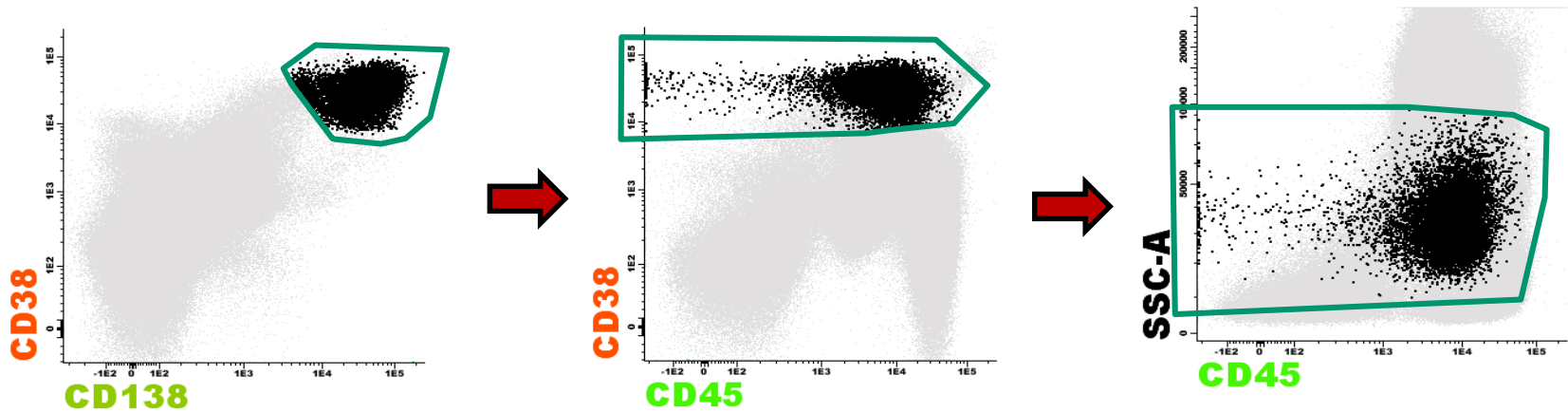


NGF analysis of a BM sample

Total events



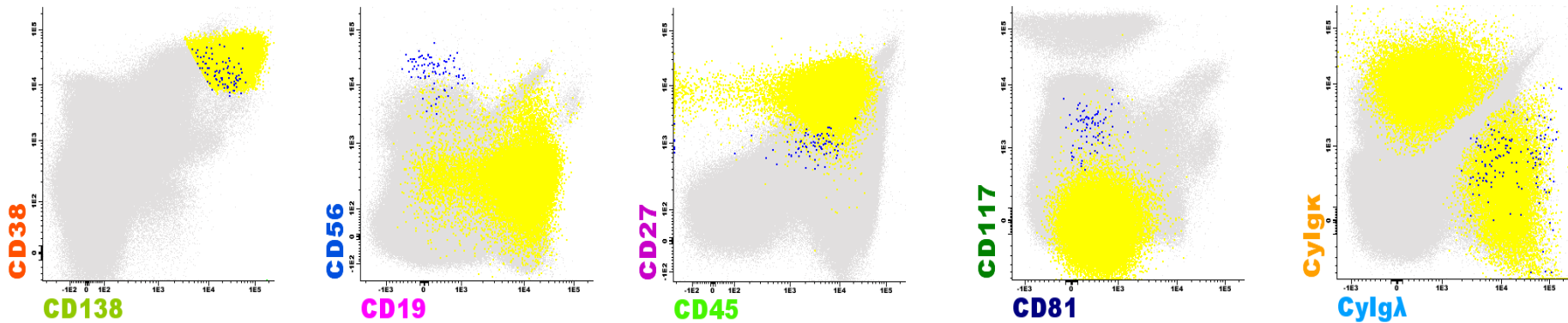
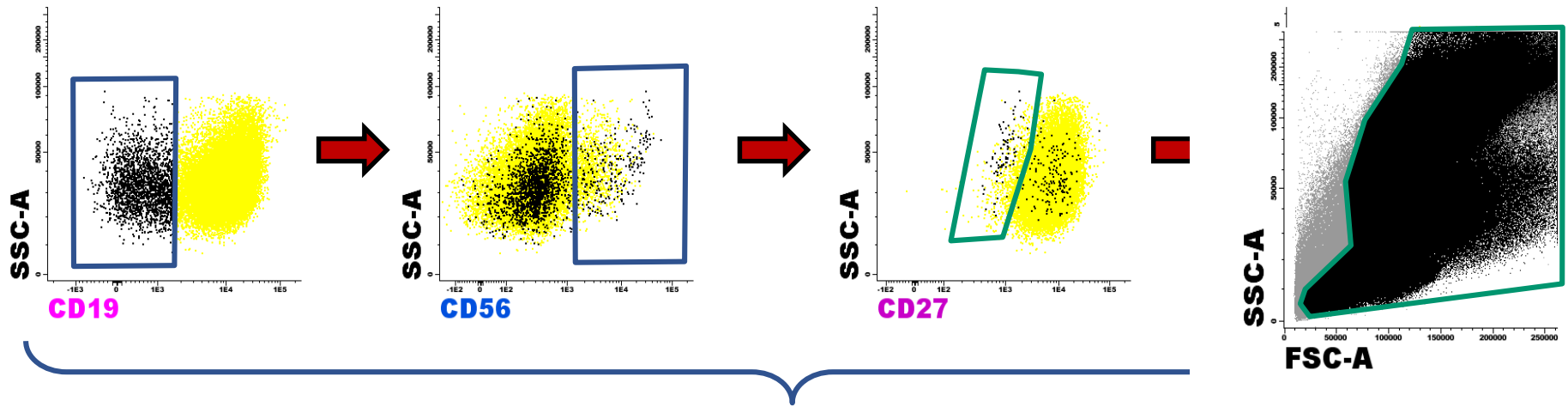
Nucleated cells



Total plasma cells

Total plasma
cells

Aberrant plasma
cells



CD38^{+/dim} CD138^{+/dim} CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27^{+/dim} CD117^{dim/+} CD81⁻ lambda

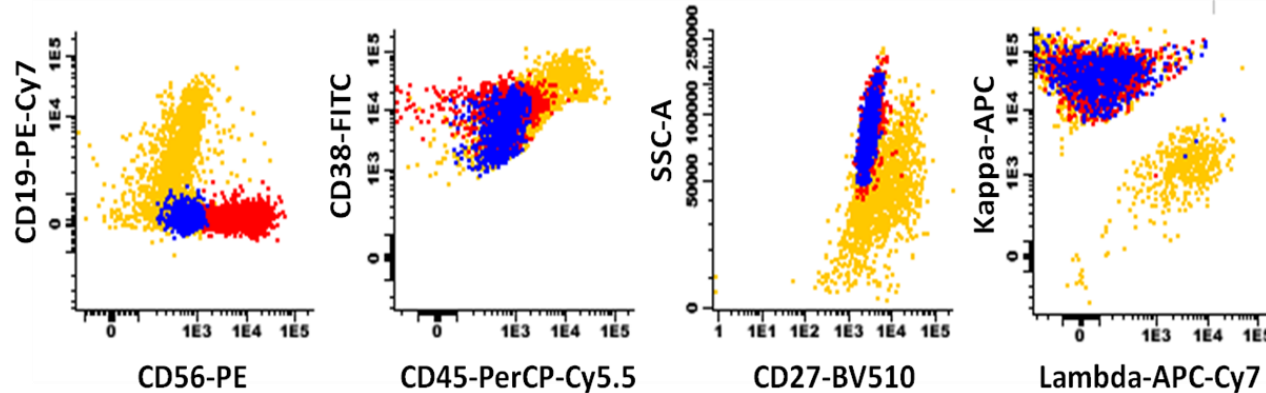
Phenotypic heterogeneity of clonal cells

MRD+ samples	CD19	CD27	CD38	CD45	CD56	CD81	CD117
1	-	dim	+	dim	-	-	+
2	-	dim	+	dim	+	-	+
3	-	dim	+	dim	-	-	-
4	-	dim	+	dim	+	-	+
5	-	+	+	dim	+	-	-
7	-	dim	dim	-	+	het	-
8	-	dim	+	dim	-	-	+
9	-	dim	+	dim	+	het	-
10	-	dim	+	dim	+	-	-
11	-	-	+	dim	-	-	-
12	-	-	dim	dim	+	-	-
13	-	dim	dim	dim	+	het	+
14	-	dim	+	dim	+	hi	dim
15	-	dim	dim	-	+	-	-
16	-	+	+	dim	+	-	-
18	-	dim	dim	dim	+	-	dim
19	-	dim	dim	dim	+	-	+
20	-	dim	+	dim	+	-	+
21	-	dim	+	dim	+	-	-
22	-	-	dim	dim	+	-	dim
29	-	dim	dim	dim	-	-	-
31	-	dim	+	dim	+	-	-
32	-	dim	dim	dim	+	-	-
33	-	dim	+	dim	+	-	-
34	-	+	+	dim	-	-	-
35	-	dim	dim	dim	+	-	+
37	-	-	dim	-	+	-	-
40	-	dim	+	dim	+	-	dim

MRD+ samples	CD19	CD27	CD38	CD45	CD56	CD81	CD117
6	-	-	+	dim	+	-	+
	-	-	dim	-	-	het	dim
23	-	dim	dim	-	dim	-	+
	-	-	dim	-	dim	-	-
25	+	dim	+	dim	+	-	dim
	-	dim	+	dim	+	het	-
30	-	dim	dim	dim	+	-	+
	-	dim	dim	dim	+	-	dim
38	-	dim	+	+	+	-	-
	-	dim	dim	dim	-	-	-

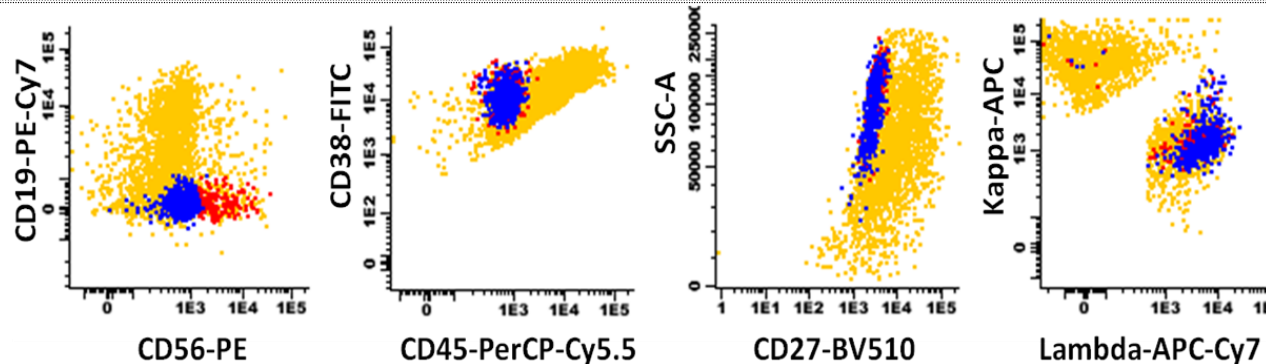
Examples of diphenotypic MRD+ cases

Normal PCs; Clone 1; Clone 2



CD19-CD56⁺CD45^{dim}CD27⁻
CD38⁺/^{dim}CD138^{hi}CD117⁺
CD81-kappa (63,5%)

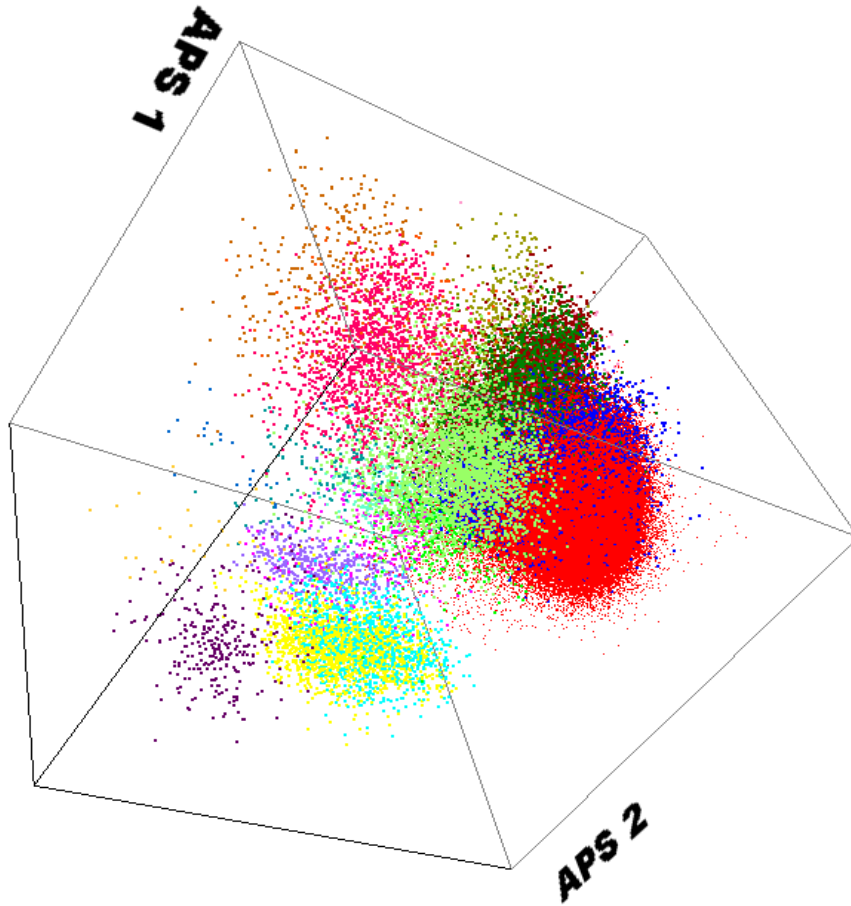
CD19-CD56⁻CD45^{dim}CD27⁻
CD38⁺/^{dim}CD138^{dim/+}CD117⁺
CD81-kappa (36,5%)



CD19-CD56⁺CD45^{dim}CD27⁻
CD38⁺CD138^{hi}CD117⁺
CD81-lambda (29,4%)

CD19-CD56⁻CD45^{dim}CD27⁻
CD38⁺CD138^{hi}CD117⁺
CD81-lambda (70,6 %)

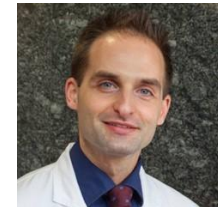
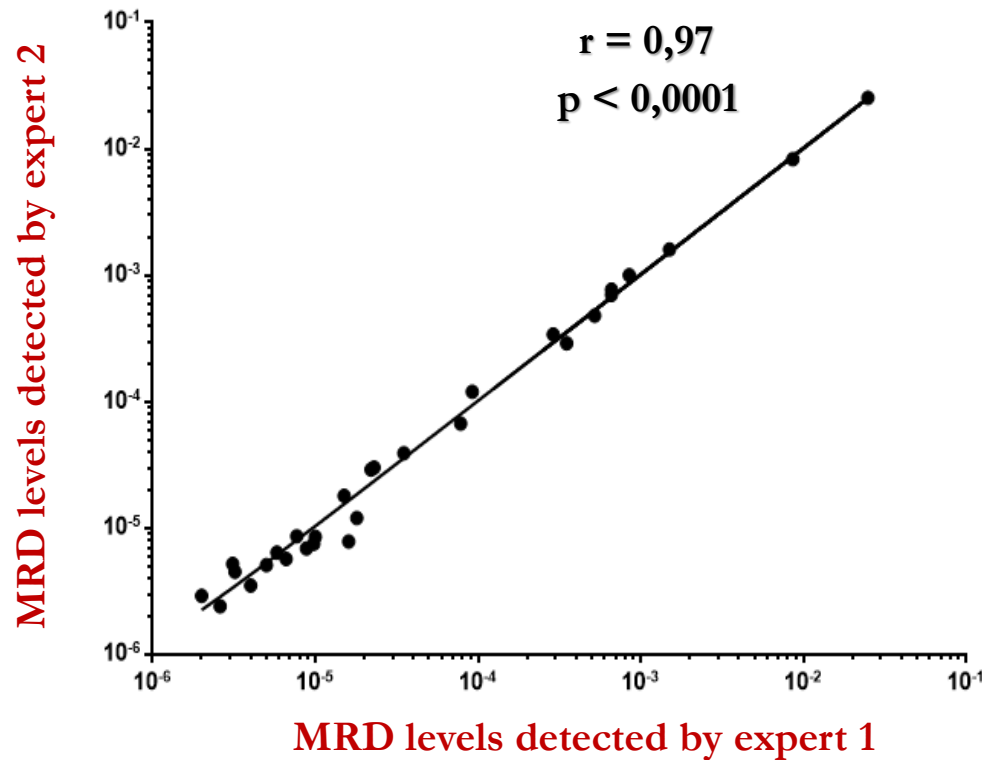
Examples of an extreme MRD+ case (with 20! phenotypes)



- 1) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81⁻ (84,6%)
- 2) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81^{dim/+} (10,8%)
- 3) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81⁻ (2,1%)
- 4) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81^{dim/+} (0,5%)
- 5) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45^{dim} CD81^{dim/+} (0,1%)
- 6) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45⁺ CD81⁻ (0,16%)
- 7) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁺ CD45⁻ CD81^{+/dim} (0,2%)
- 8) CD19⁻ CD56⁺ CD38^{dim} CD27^{dim} CD117⁺ CD45⁺ CD81^{dim} (0,04%)
- 9) CD19⁺ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81^{dim/+} (0,1%)
- 10) CD19⁺ CD56⁺ CD38^{dim} CD27^{dim} CD117⁺ CD45⁺ CD81^{dim/+} (0,16%)
- 11) CD19⁺ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45^{dim} CD81⁺ (0,01%)
- 12) CD19⁺ CD56⁺ CD38^{dim} CD27^{dim} CD117⁻ CD45⁺ CD81⁺ (0,1%)
- 13) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45^{dim} CD81^{dim/+} (0,12%)
- 14) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45⁺ CD81⁻ (0,25%)
- 15) CD19⁻ CD56⁻ CD38^{dim} CD27⁺ CD117⁻ CD45⁺ CD81⁻ (0,5%)
- 16) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁺ CD45⁻ CD81^{+/dim} (1,5%)
- 17) CD19⁻ CD56⁻ CD38^{dim} CD27^{dim} CD117⁺ CD45⁺ CD81^{dim} (0,04%)
- 18) CD19⁺ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45⁻ CD81^{dim/+} (1,3%)
- 19) CD19⁺ CD56⁻ CD38^{dim} CD27^{dim} CD117⁻ CD45⁺ CD81⁺ (0,07%)
- 20) CD19⁺ CD56⁻ CD38^{dim} CD27^{dim} CD117⁺ CD45⁻ CD81^{dim/+} (0,4%)

How reliable & reproducible are NGF data ?

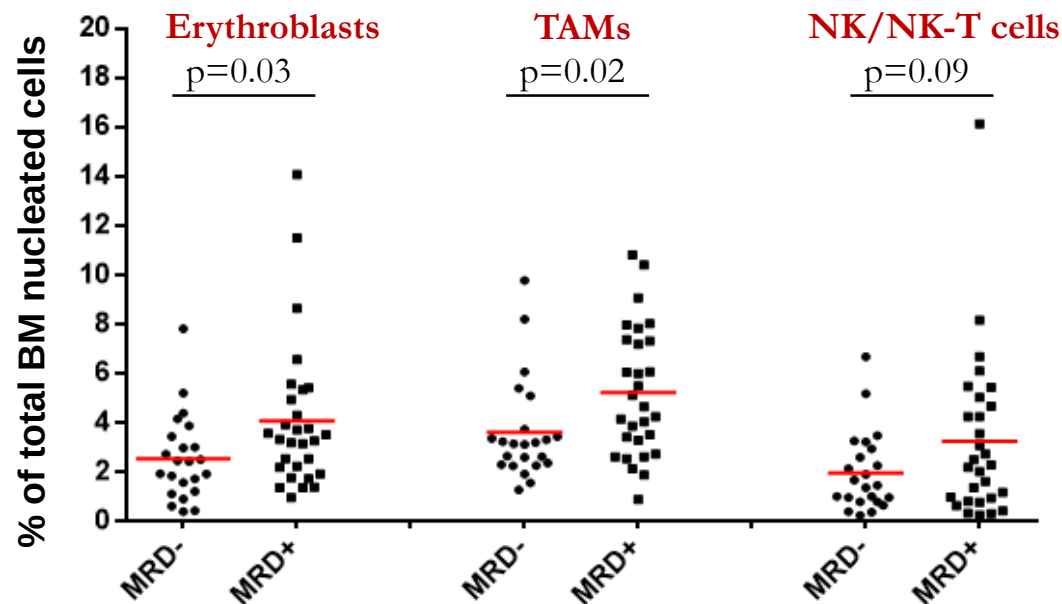
- Analysis of 74 samples
- Match in 73/74
- Reproducibility: 98.6 %



Study of all main BM populations (15)

BM subsets	CD19	CD27	CD38	CD45	CD56	CD81	CD117	CD138	SSC
Lymphoid lineage									
Plasma cells			bright					bright	
B cells	+			+ / dim					low
Naïve B cells	+	–	– / dim	+					
B-cells precursors	+	–	bright	dim					
Memory B cells	+	+	– / dim	+					
T cells	–			+	–				low
NK & NK-T cells	–			+	+				low
Myeloid lineage									
Basophils			+	dim		–			
Neutrophils				dim		–			high
Eosinophls				bright		bright			high
Myeloid progenitor cells			+	dim			+		high
Monocytes			+	+		+			int
Mast cells				dim			bright		
Erythroid lineage									
Erythroblasts			–	–					low
Erythroid progenitor cells			– / dim	– / dim			+		low

Differences in BM niche profile in MRD+ & MRD- pts



MM patients in CR ≥ 2 y
Median age: 64 y
Men/women: 1.25

Bone Marrow subset	Patient #8		Patient #32		Patient #46		Patient #47	
	MRD- 1 st evaluation	MRD+ Re-evaluation	MRD- 1 st evaluation	MRD+ Re-evaluation	MRD- 1 st evaluation	MRD+ Re-evaluation	MRD- 1 st evaluation	MRD+ Re-evaluation
Erythroblasts	0.97	2.21	1.24	3.34	3.14	4.97	2.87	2.94
Erythroid progenitors	0.04	0.04	0.08	0.24	0.16	0.15	0.05	0.05
T cells	1.73	2.06	3.61	0.94	11.87	8.92	3.80	5.42
NK cells	0.43	0.44	1.46	0.33	4.64	2.53	2.07	2.04
B cells (total)	3.80	2.13	0.15	0.21	2.52	2.24	0.81	0.65
Naïve B cells	2.79	1.74	0.04	0.04	1.90	1.47	0.38	0.29
Memory B cells	0.89	0.86	0.09	0.04	0.53	0.54	0.33	0.26
B-cell precursors	0.07	0.02	0.002	0.13	0.05	0.17	0.06	0.05
Myeloid precursors	0.19	0.06	0.15	0.11	0.27	0.62	0.85	0.23
Monocytes	3.64	7.34	6.02	1.90	4.08	7.85	5.84	7.98
Neutrophils	77.5	77.9	74.46	85.77	56.81	59.16	73.94	70.69
Basophils	0.02	0.02	0.16	0.05	0.07	0.22	0.34	0.14
Eosinophils	0.21	0.12	0.007	0.005	0.17	0.08	0.22	0.23
Mast cells	0.002	0.006	0.005	0.014	0.07	0.02	0.01	0.01

Immune signature of response to treatment in MM patients in CR ≥ 2 y

- Aberrant clonal plasma cells in the BM $\Rightarrow 10^{-3}$ to 10^{-6} does not make a difference
- Higher frequencies of erythroblasts $\Rightarrow$ support clonal cell propagation
- Higher frequencies of tumor-associated monocytes/macrophages $\Rightarrow$ secretion of IL-6 supporting myeloma cell growth
- Higher frequencies of NK/NK-T cells $\Rightarrow$ competent mechanism of immune surveillance
- No difference in B cells, T cells, neutrophils, basophils, eosinophils, mast cells, etc
- No impact of gender, sex, cytogenetic aberrations, type of frontline therapy, duration of CR, clinical and laboratory parameters

➤ ***Proposed immune signature:***

MRD+ with clonal cells CD19-CD45⁻/^{dim}CD56-CD27^{dim}

↑ CD45-CD38-SSC^{low}

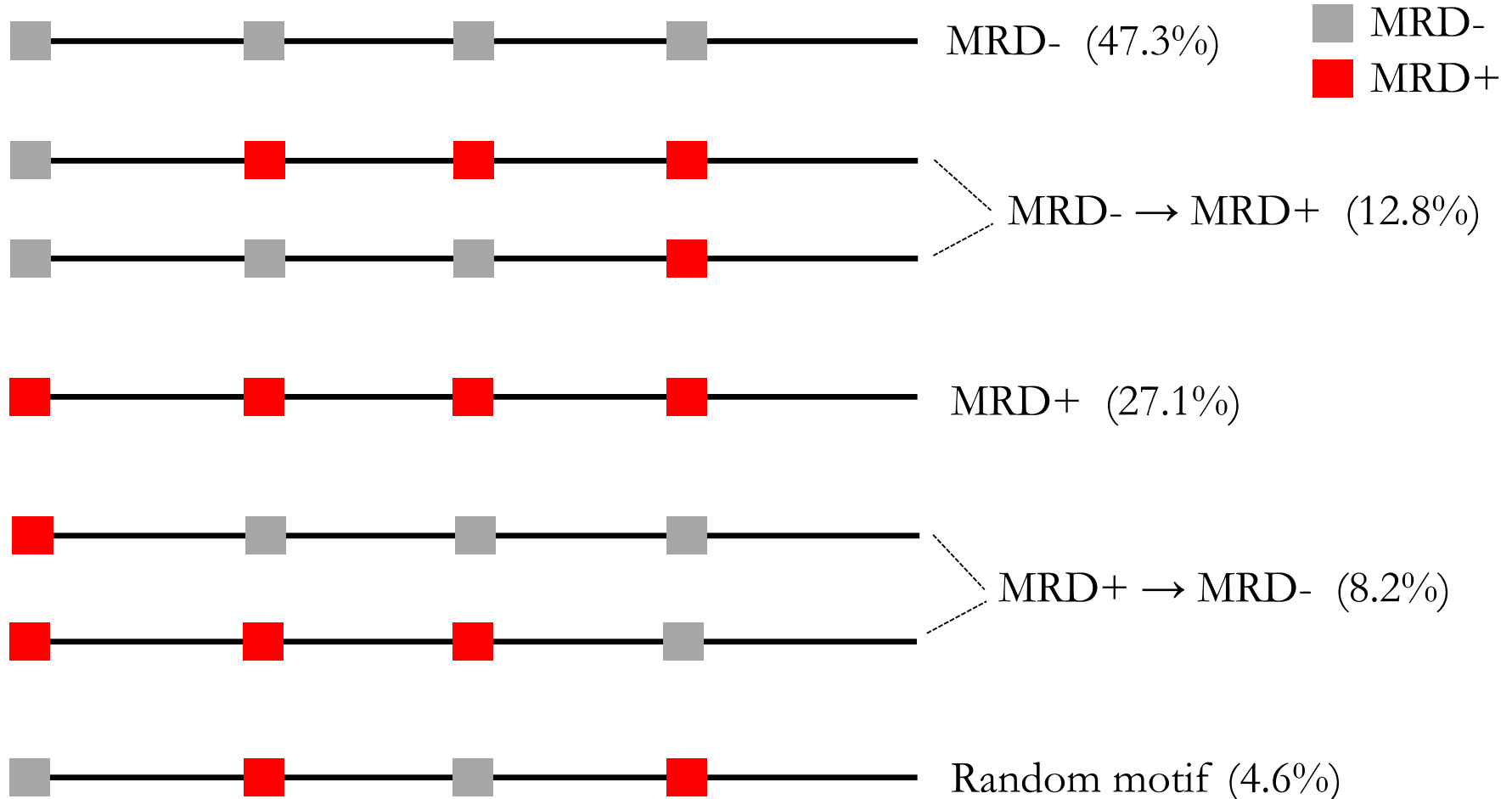
↑ CD45+CD38+CD81+SSC^{int}

↑ CD19-CD45+CD56+SSC^{low}

***Likely to
relapse***

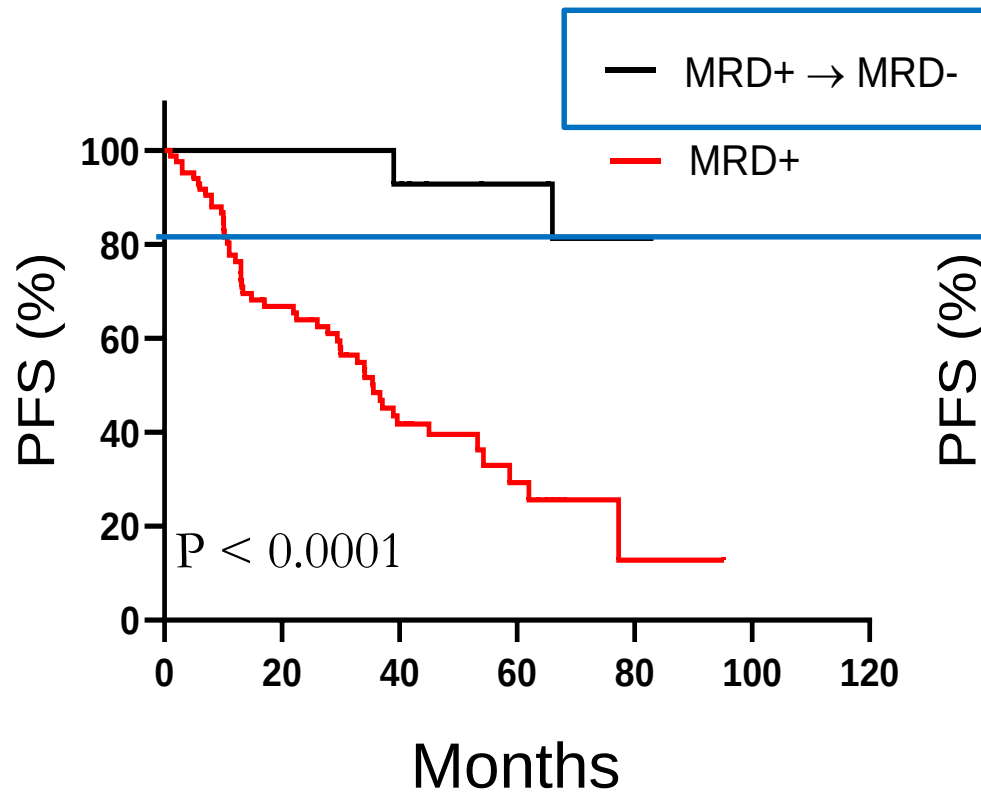
MRD patterns over subsequent evaluations

N=340 pts; N=1836 MRD evaluations; LOD=2.1x10⁻⁶; mean follow-up: 48 m; 14 subpopulations

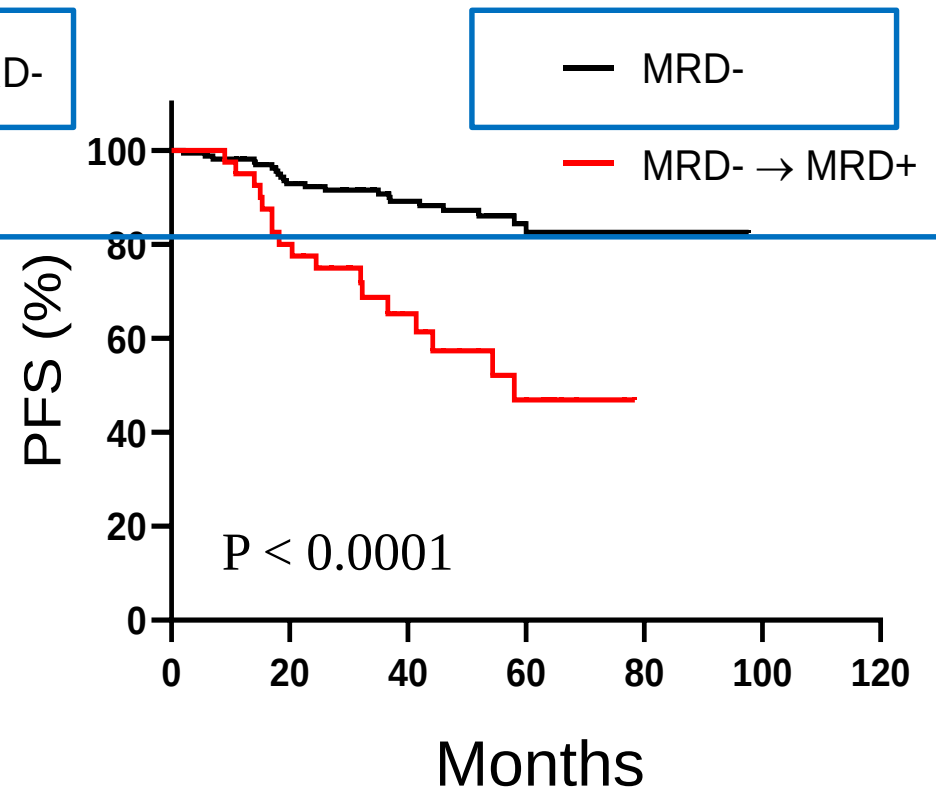


Prognostic value of MRD dynamics

HR 0.24, 95%CI: 0.13-0.43

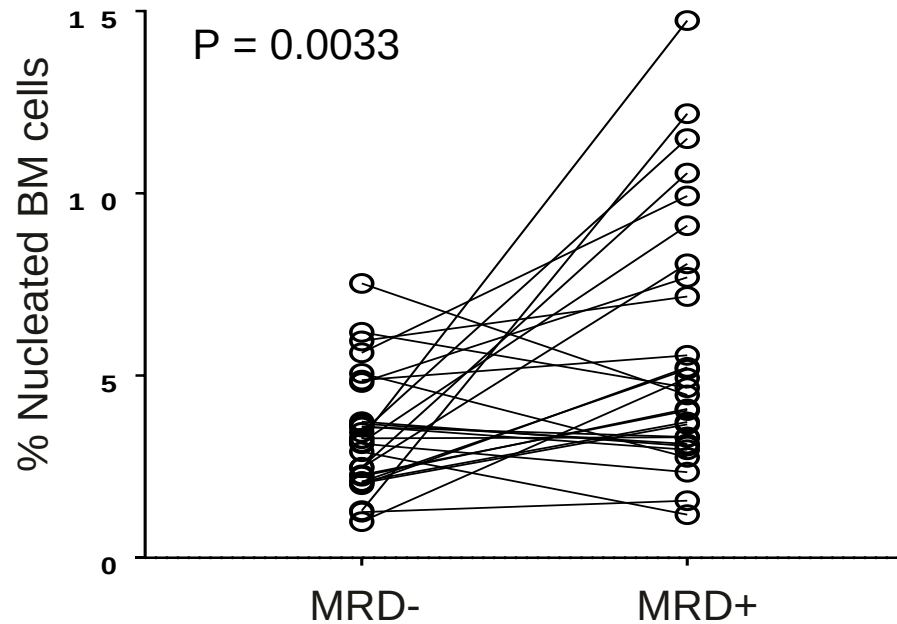


HR:0.26, 95%CI: 0.11-0.60

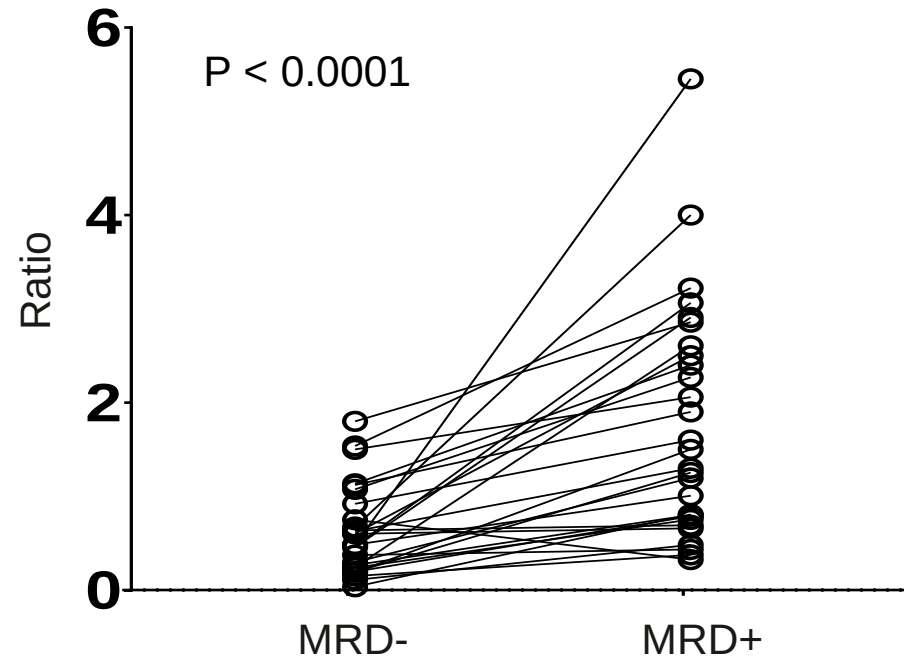


Changes in the TME at response-to-treatment

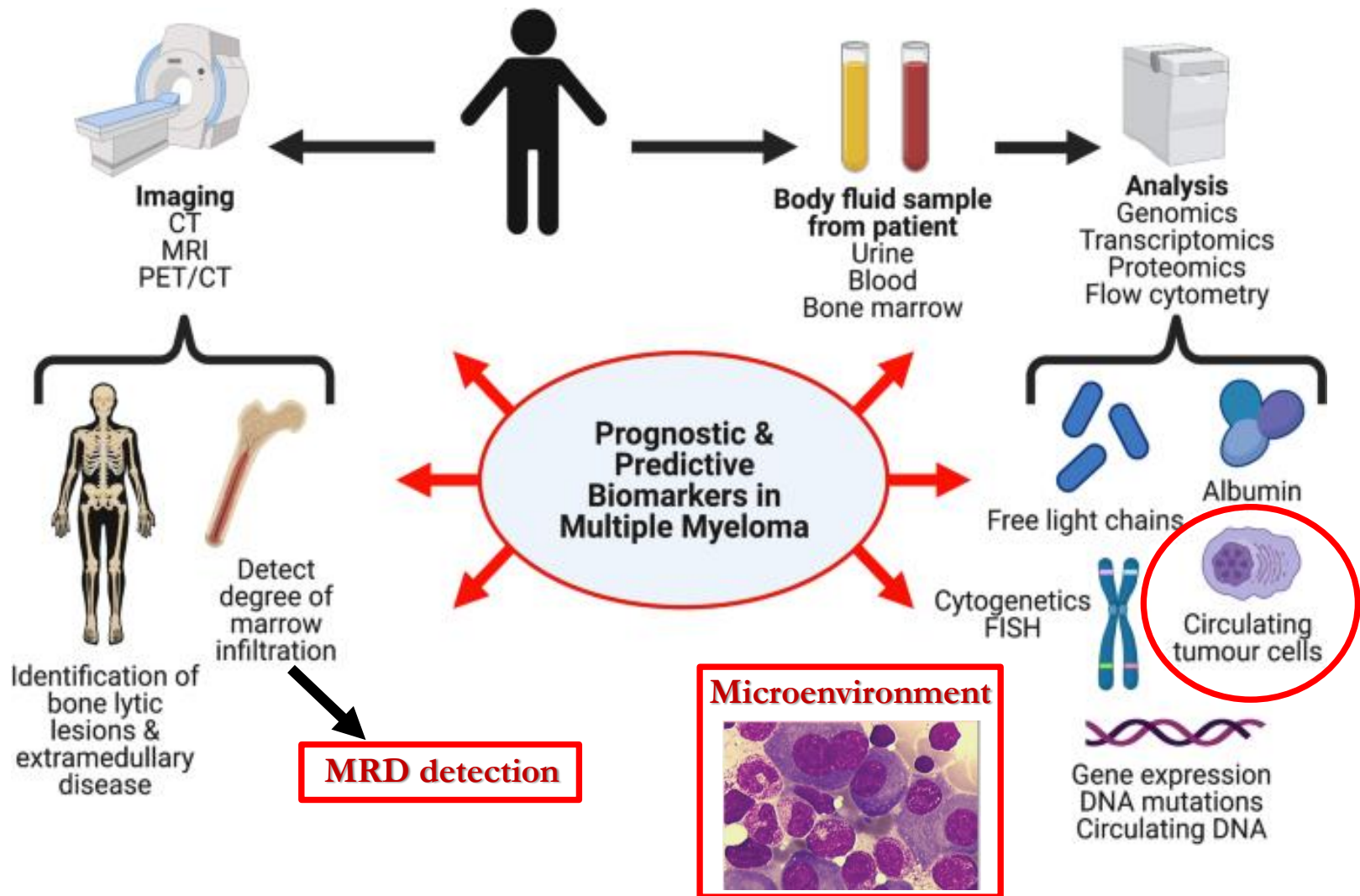
NK cells



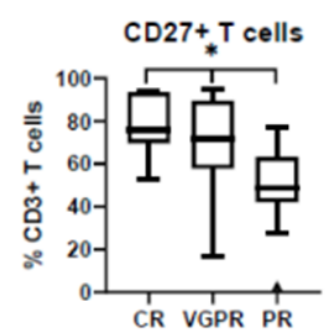
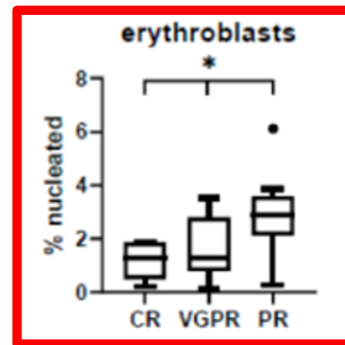
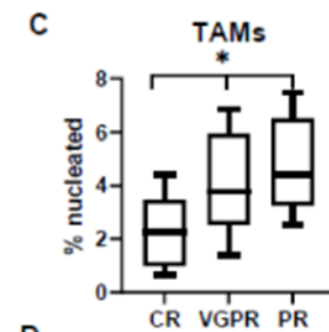
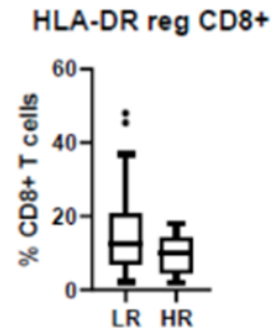
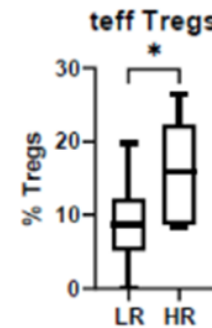
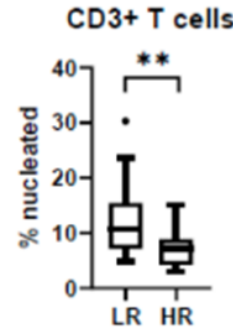
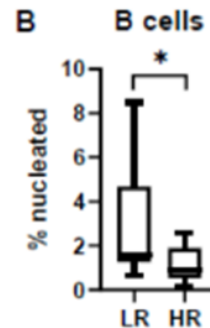
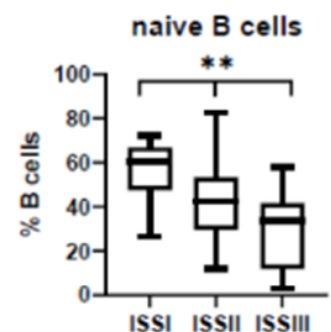
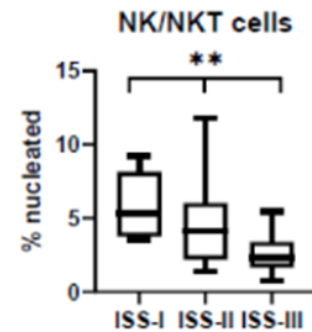
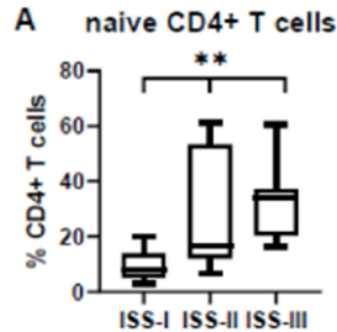
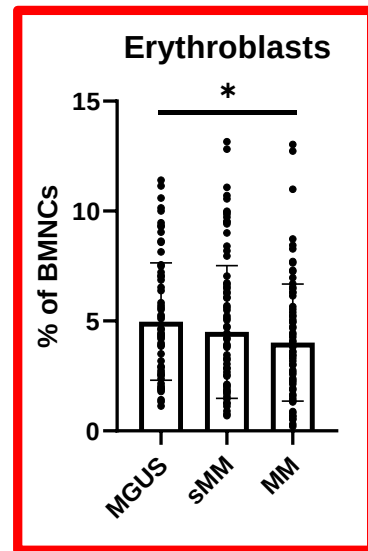
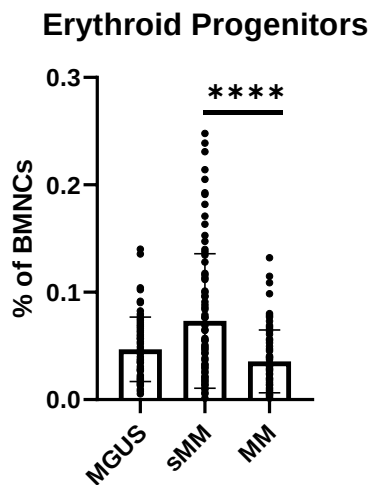
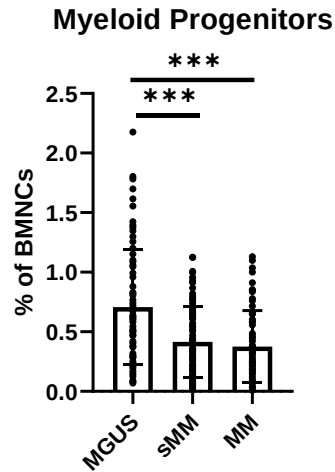
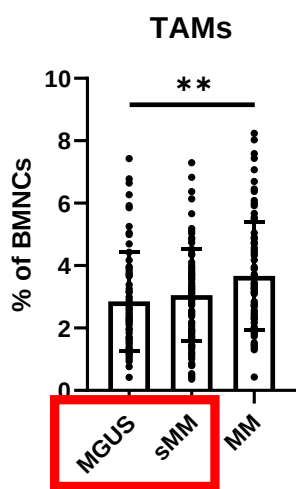
Memory/naïve B cells



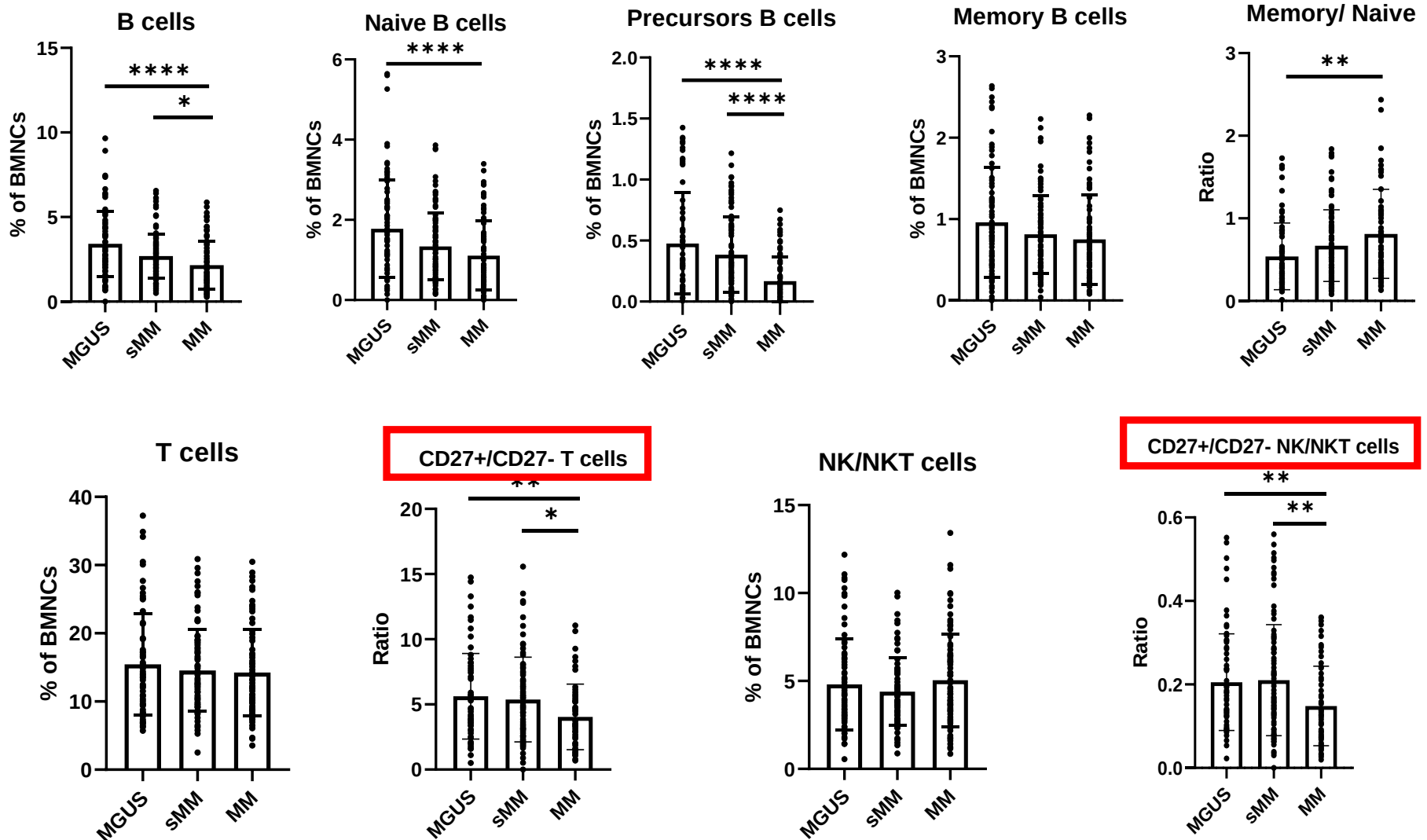
Need for identifying novel predictive/prognostic biomarkers



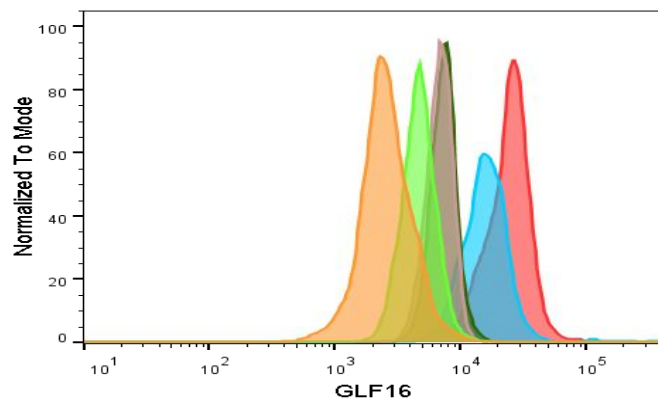
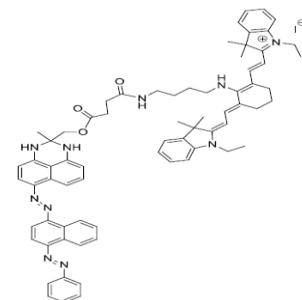
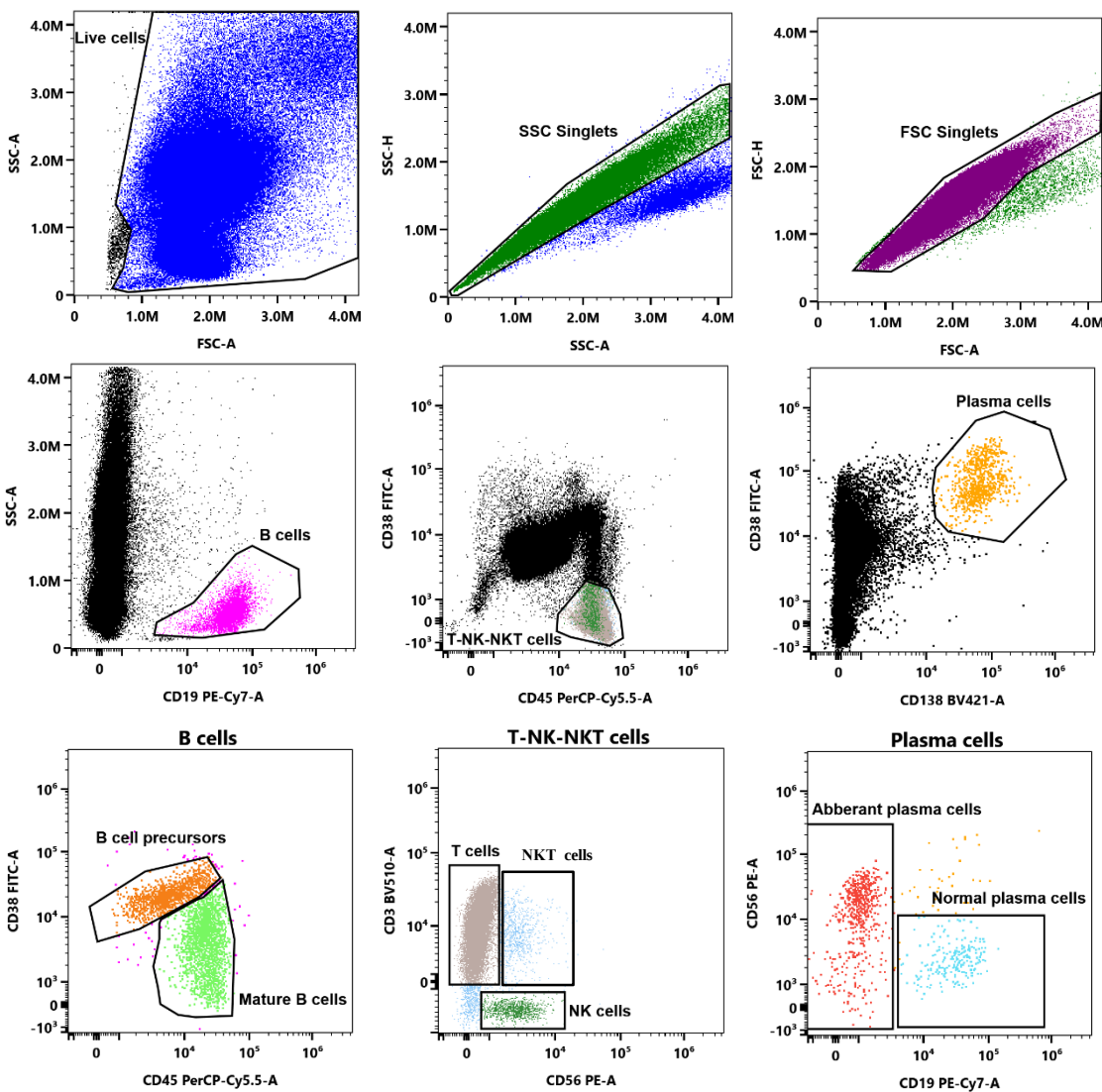
Immune subpopulations during disease evolution



Immune subpopulations during disease evolution



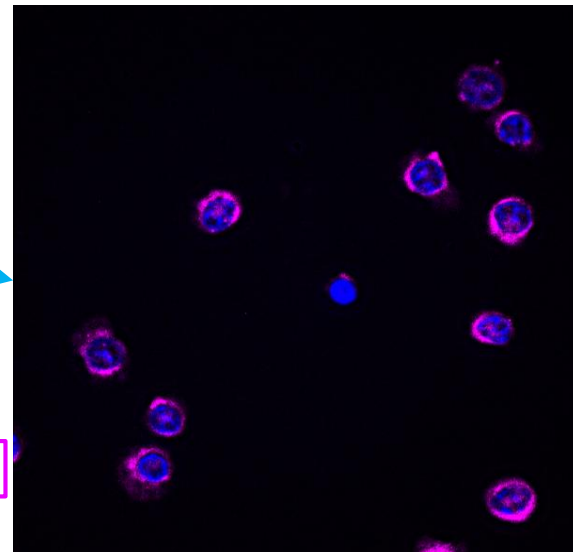
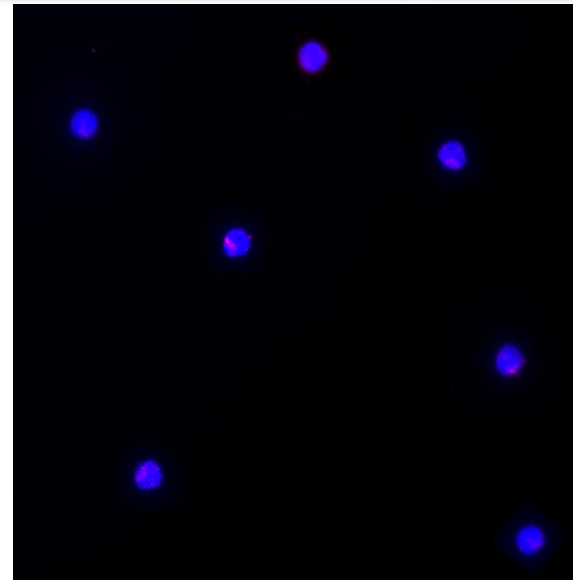
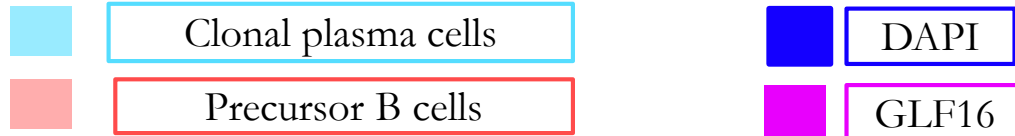
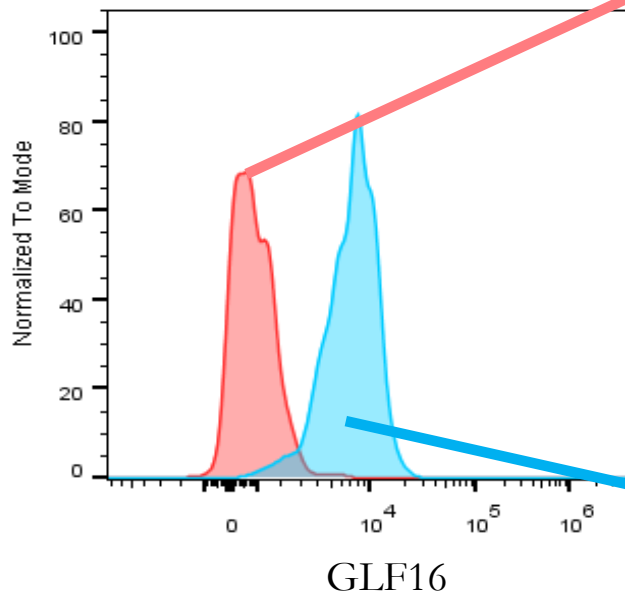
GLF16: fluorescence analogue of Sudan Black-B



		MedFl ± SE		
		MGUS	sMM	MM
B cell precursors		2870 ± 160	2614 ± 142	2799 ± 182
Mature B cells		4999 ± 234	4721 ± 198	4500 ± 256
T cells		6889 ± 355	6742 ± 296	6596 ± 329
NK cells		7365 ± 368	7020 ± 317	6171 ± 248
Normal Plasma cells		15962 ± 945	12973 ± 912	10716 ± 845
Aberrant Plasma cells		22934 ± 962	19567 ± 937	12926 ± 976

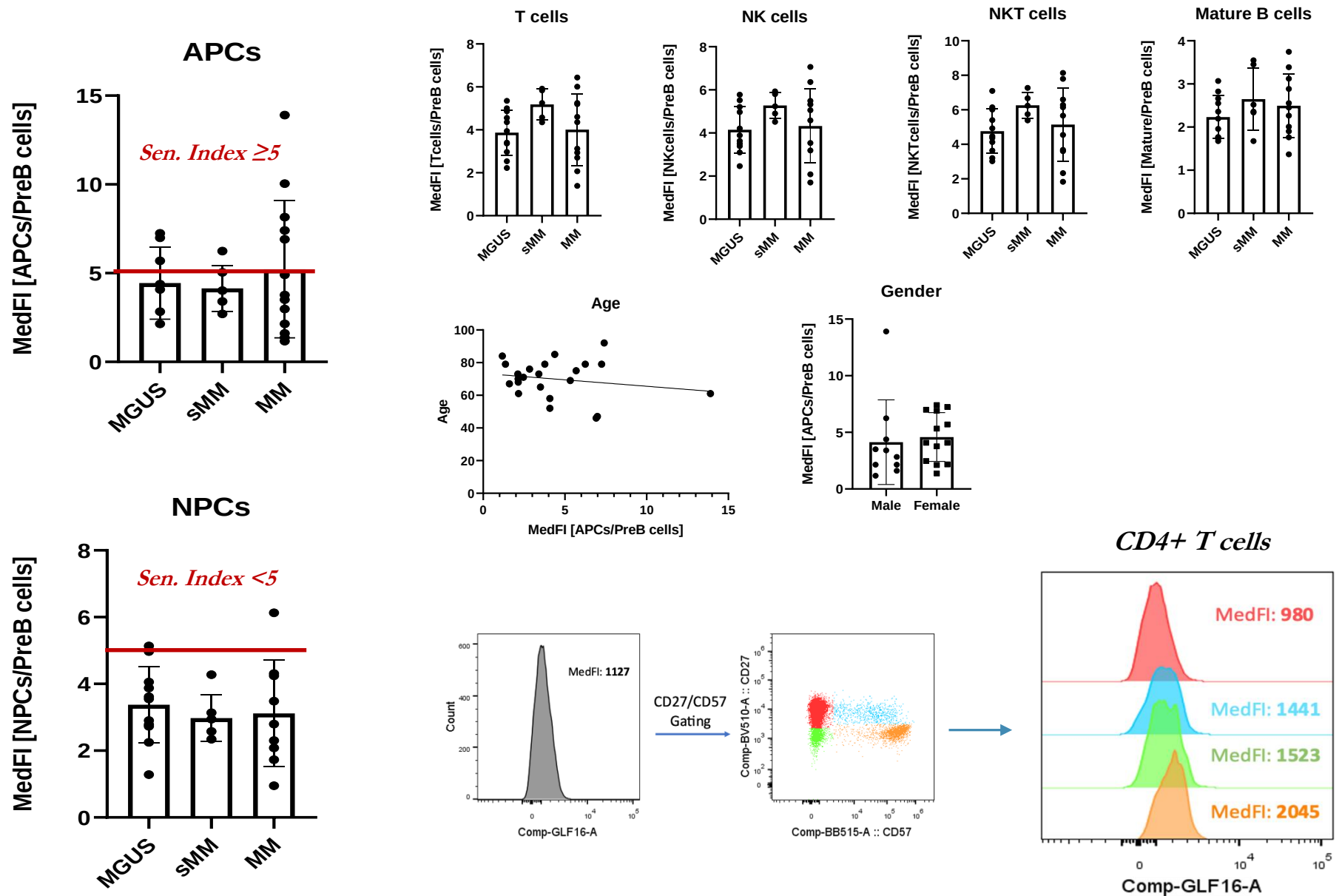
Unpublished data, pls do not copy

Clonal plasma cells stain +/-ve with GLF16

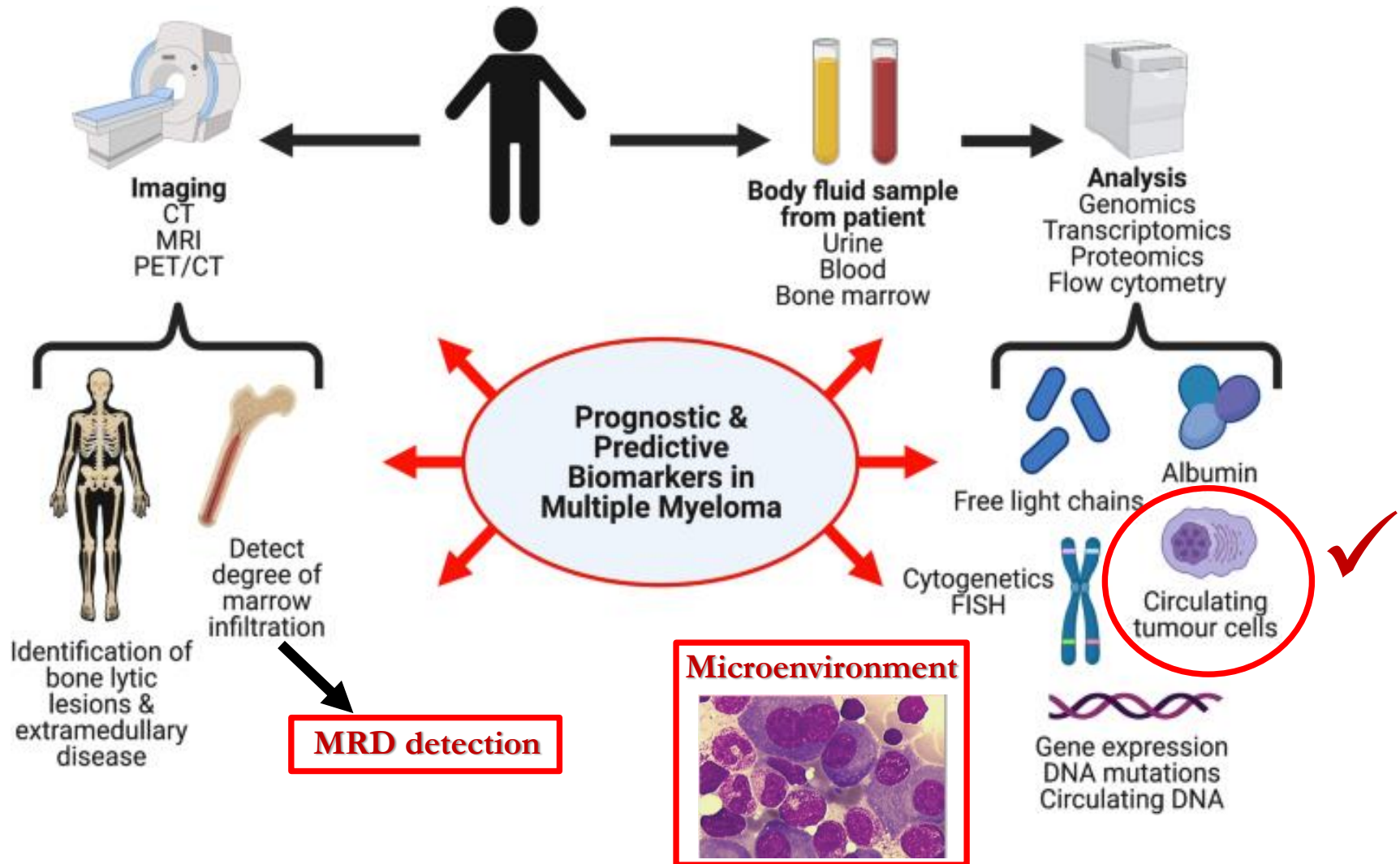


Senescence Index: MedFI [each subset/precursor B cells]

GLF16 expression in the BM during disease evolution



Need for identifying novel predictive/prognostic biomarkers



Bypassing BM aspiration...

- Functional imaging

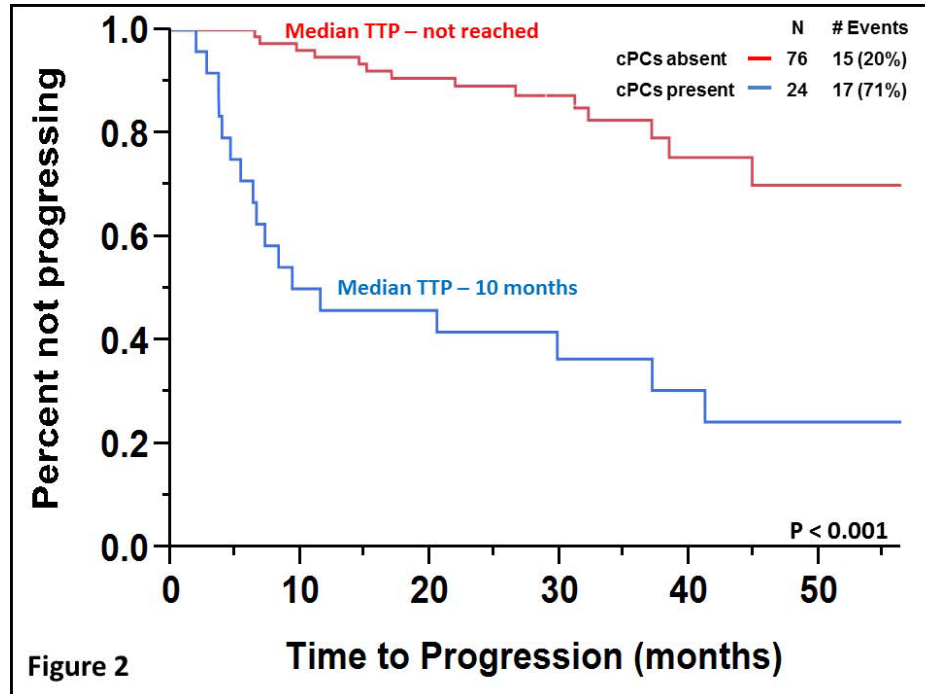
(FDG-PET, WBLDCT, MRI, DW-MRI, immunePET...)

- Liquid biopsy

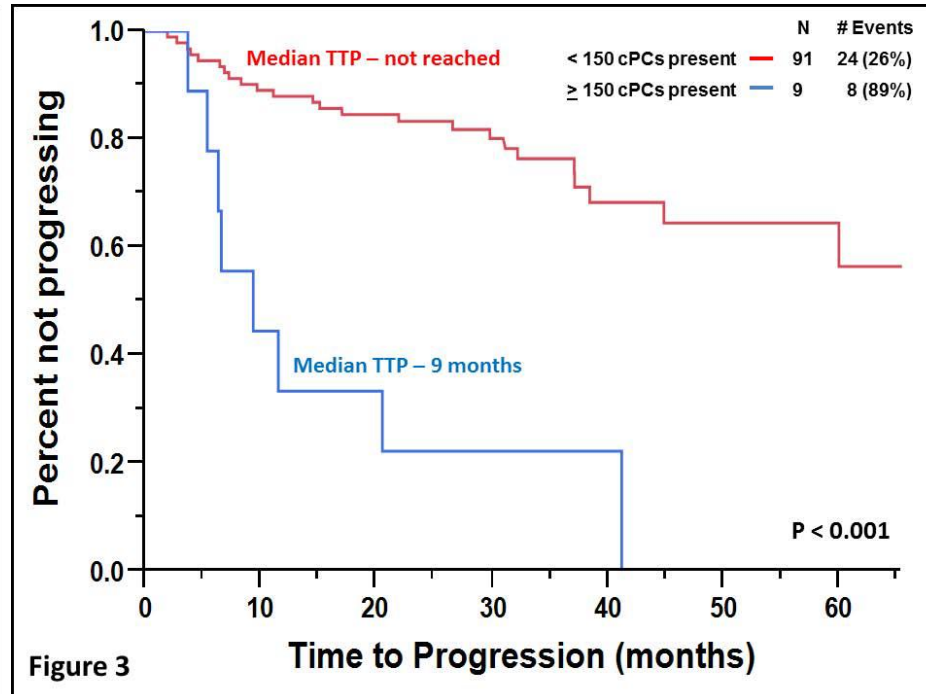
(MM circulating cells/CCPCs/CTCs, nucleic acids/cfDNA)

Higher number of CTCs in sMM patients at high risk of progression

CTC +/- versus -/+

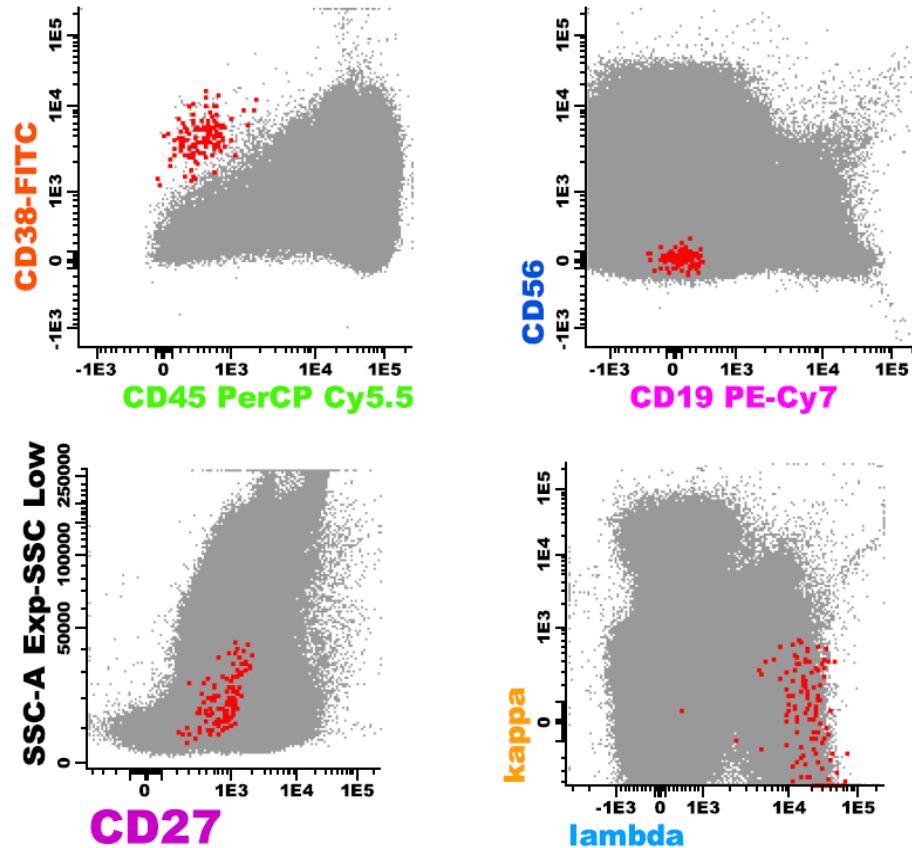


CTCs < 150 versus ≥ 150

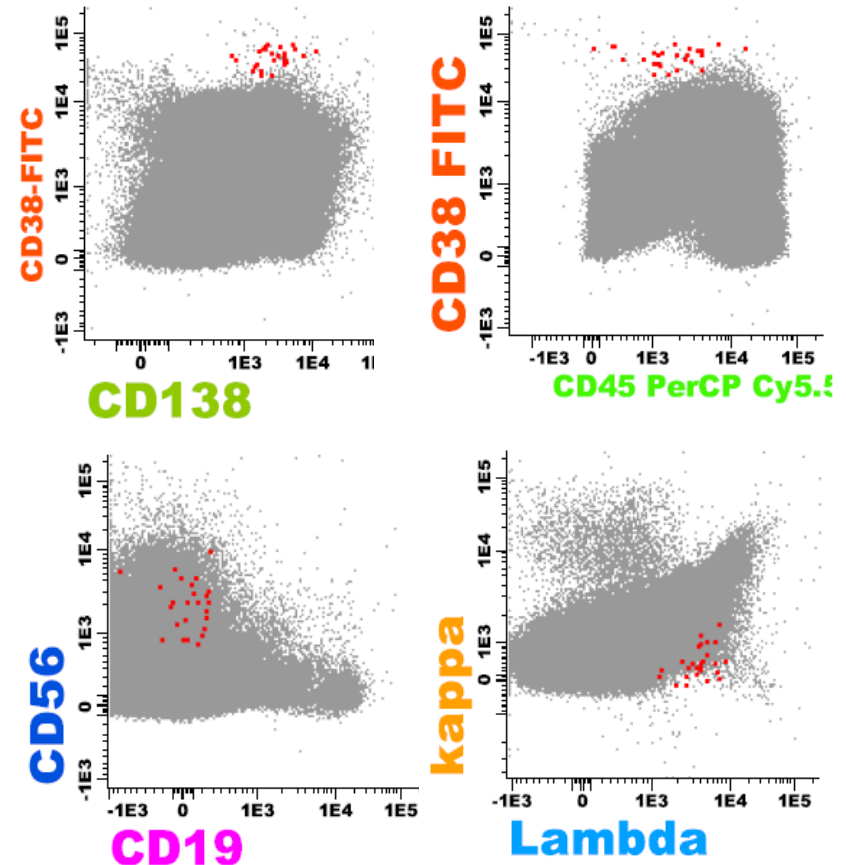


Circulating clonal plasma cells at diagnosis

CTCs: 0.003%



CTCs: 0.00048%



88% diagnostic MM samples & 53% sMM samples
with a minimum cut-off of 25 APCs for LOD

Comparison between BM and PB at diagnosis (I)

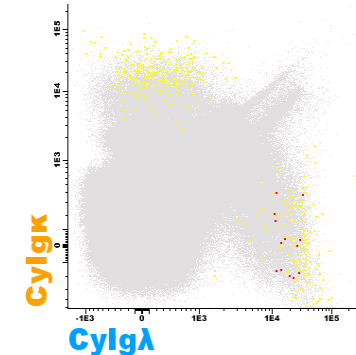
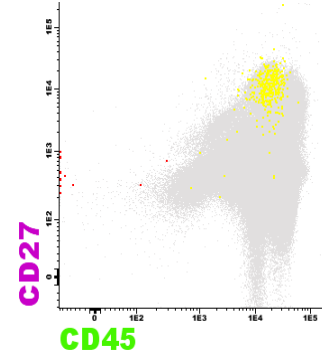
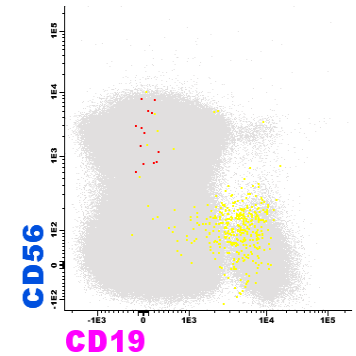
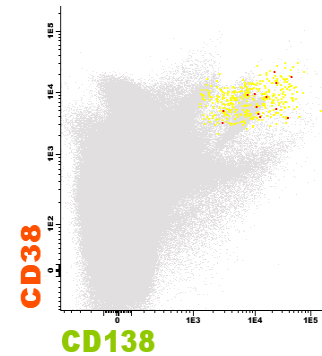
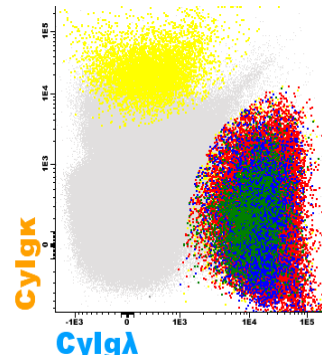
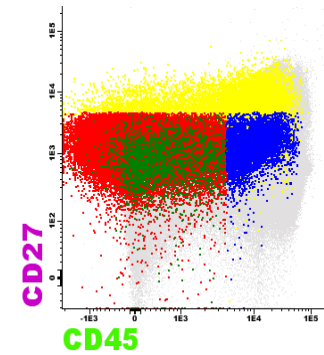
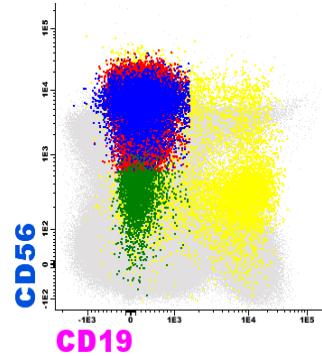
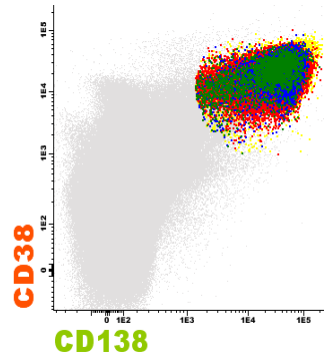
Bone marrow

Peripheral blood

Level of APCs

10^{-2}

10^{-6}



1. $CD38^{+}CD138^{++}CD19^{-}CD56^{+}CD45^{-}CD27^{dim}\lambda$ (80%)
2. $CD38^{+}CD138^{++}CD19^{-}CD56^{+}CD45^{+}CD27^{dim}\lambda$ (17%)
3. $CD38^{+}CD138^{++}CD19^{-}CD56^{-}CD45^{-}CD27^{dim}\lambda$ (3%)

$CD38^{+}CD138^{++}CD19^{-}CD56^{+}CD45^{-}CD27^{dim}\lambda$

Comparison between BM and PB at diagnosis (II)

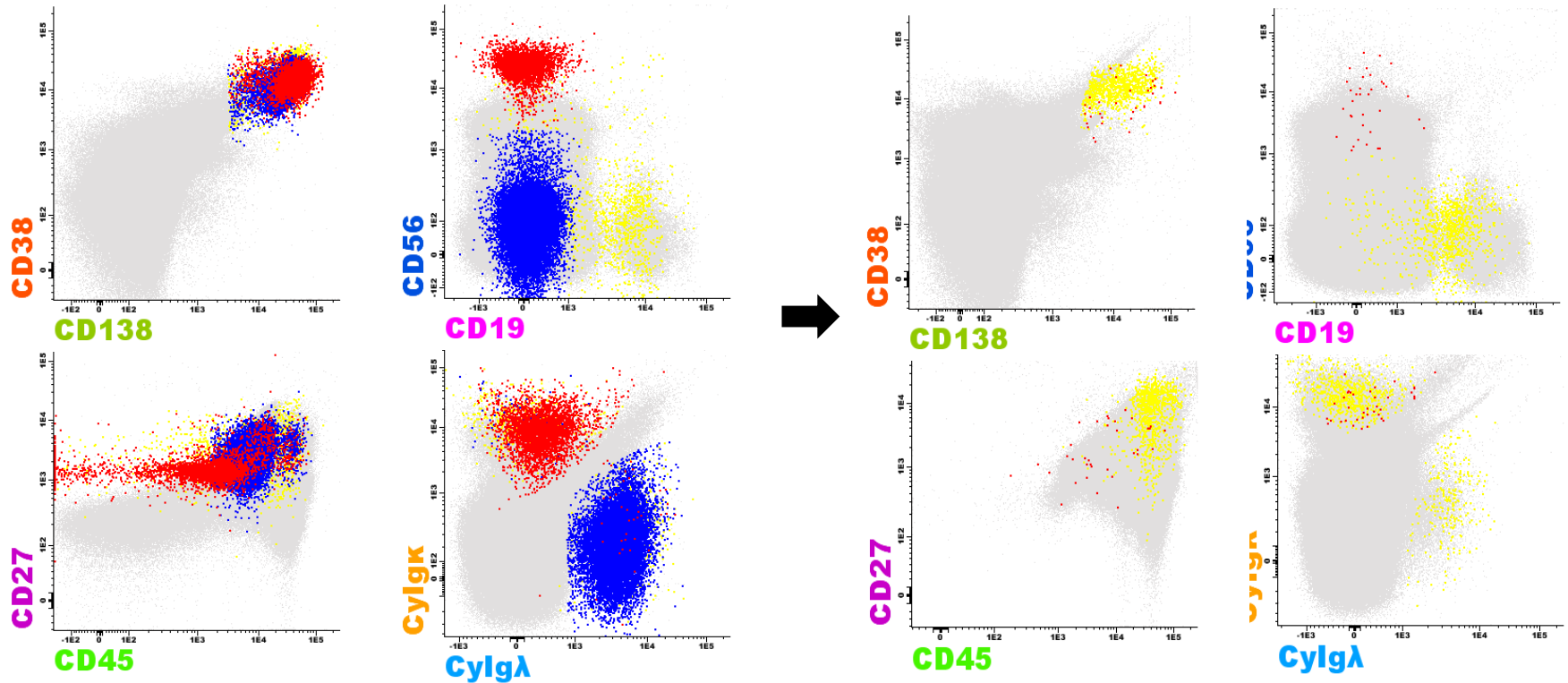
Bone marrow

Peripheral blood

Level of APCs

10^{-2}

10^{-6}



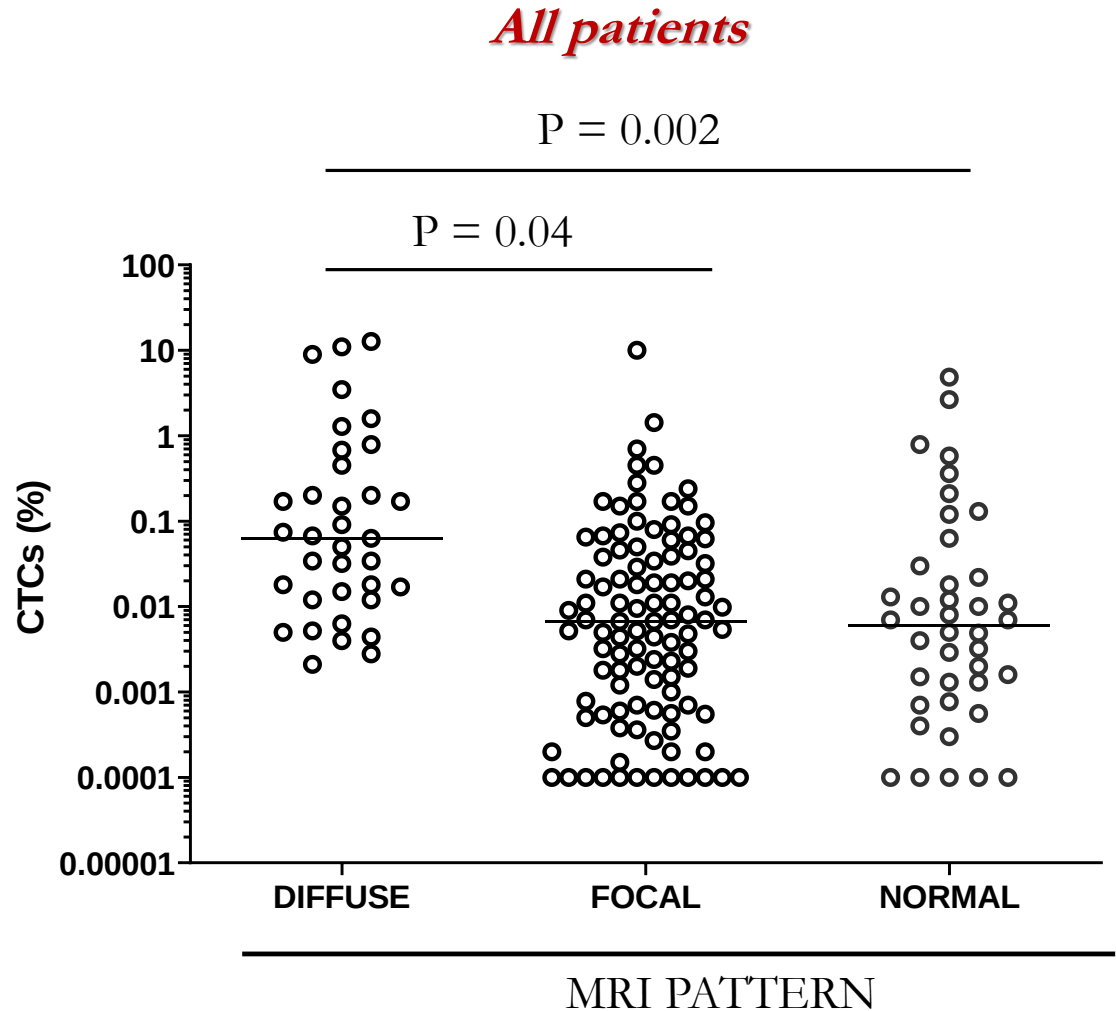
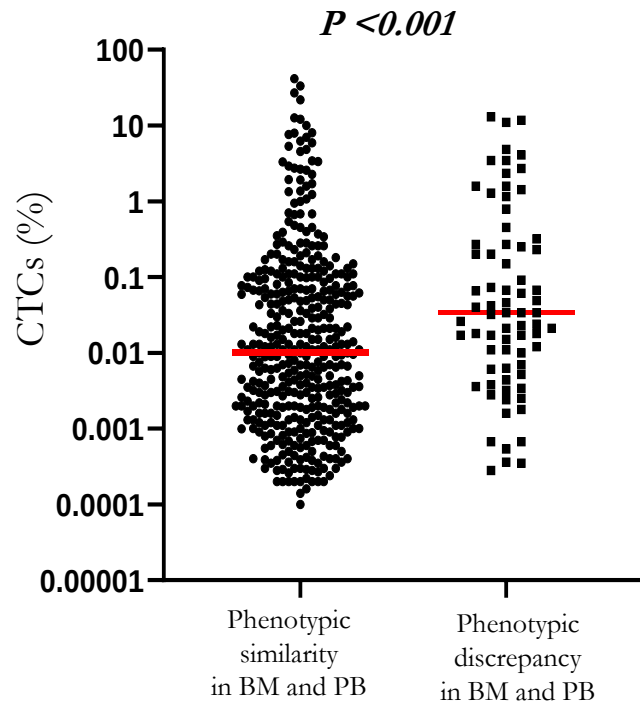
1. $CD38^{+}CD138^{++}CD19^{-}CD56^{-}CD45^{dim/+}CD27^{dim/+}\lambda$ (83%)

2. $CD38^{+}CD138^{++}CD19^{-}CD56^{++}CD45^{-/dim}CD27^{dim}\kappa$ (17%)

$CD38^{+}CD138^{++}CD19^{-}CD56^{++}CD45^{dim}/CD27^{dim}\kappa$

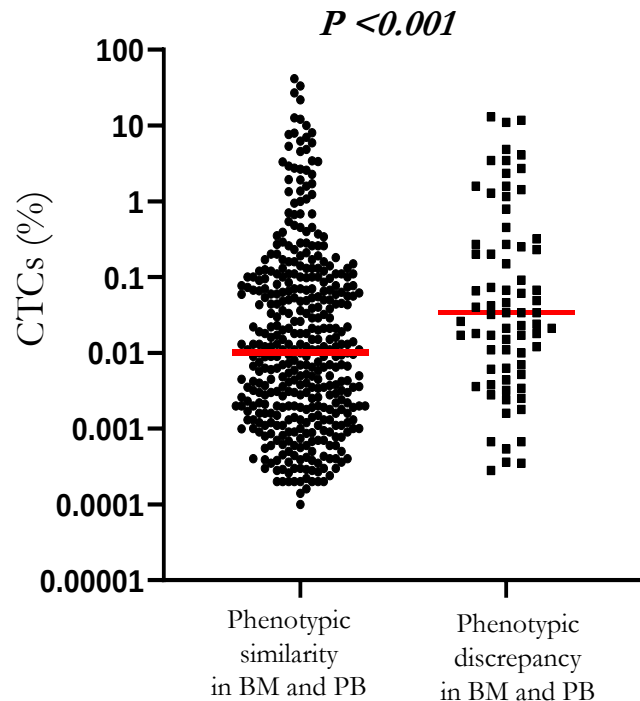
High CTCs as a marker of multiple infiltrated BM sites

*Phenotypic disagreement in
73/493 samples (14.8%)*

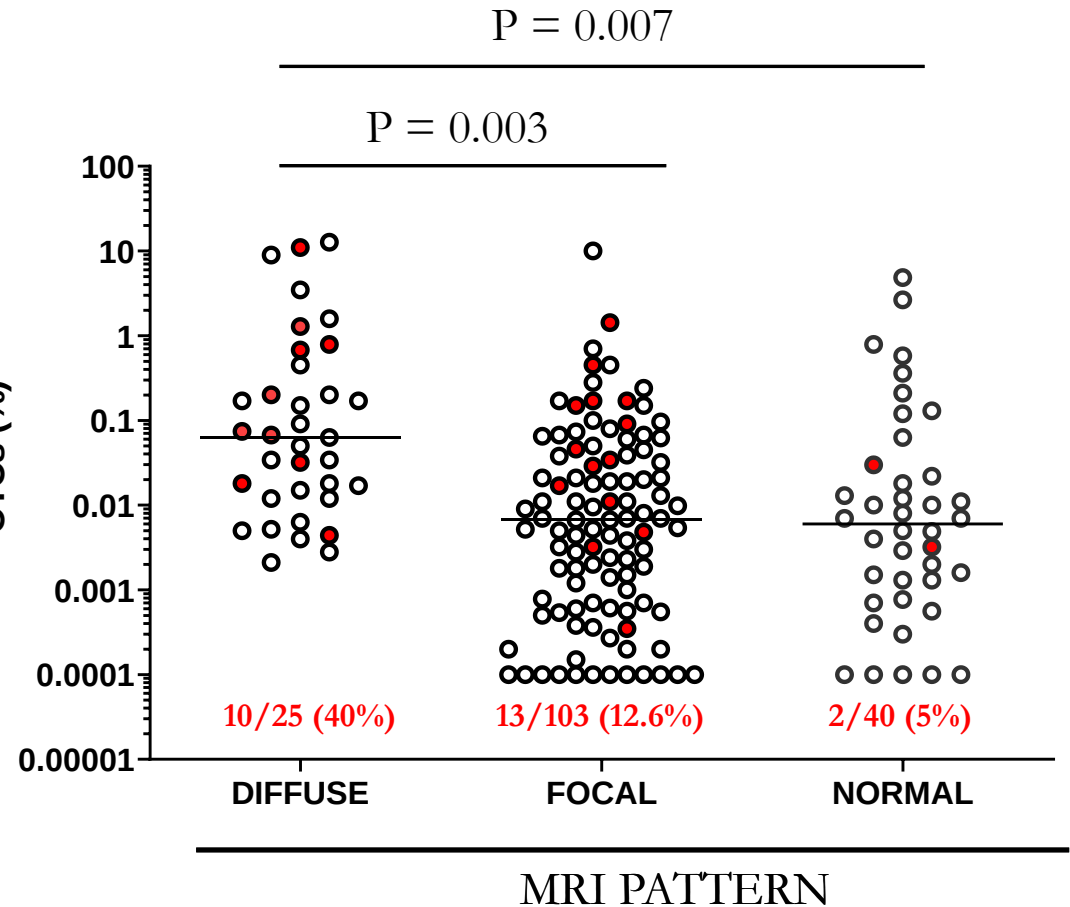


High CTCs as a marker of multiple infiltrated BM sites

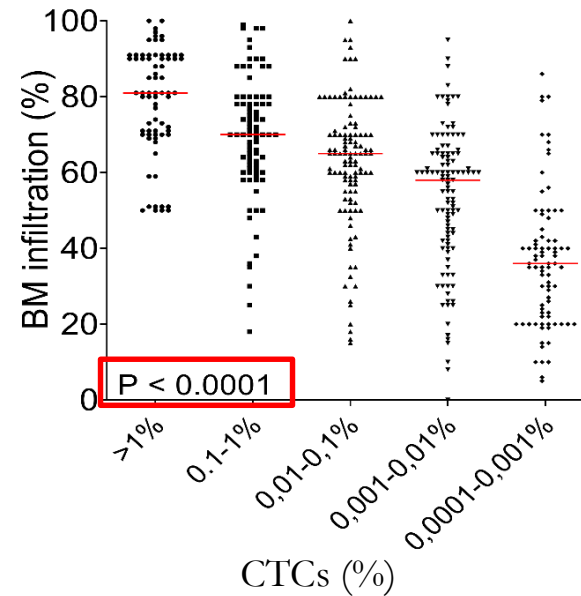
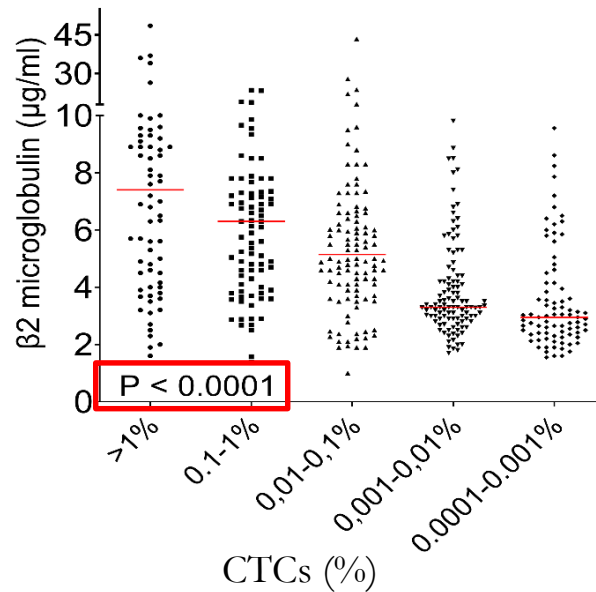
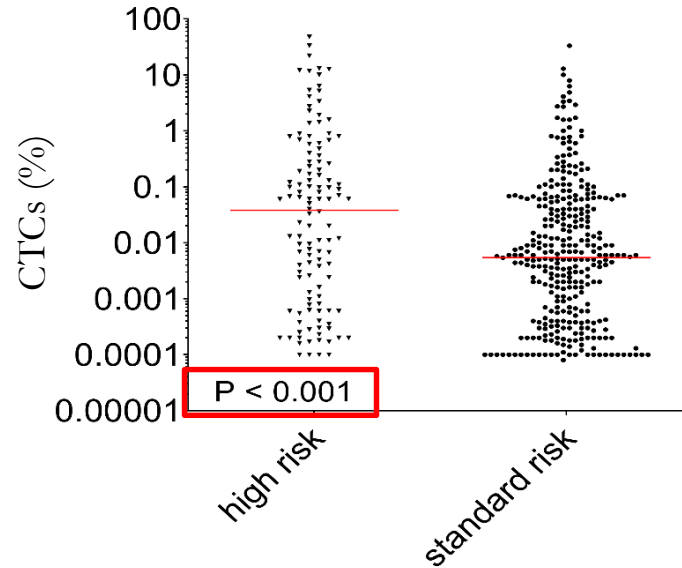
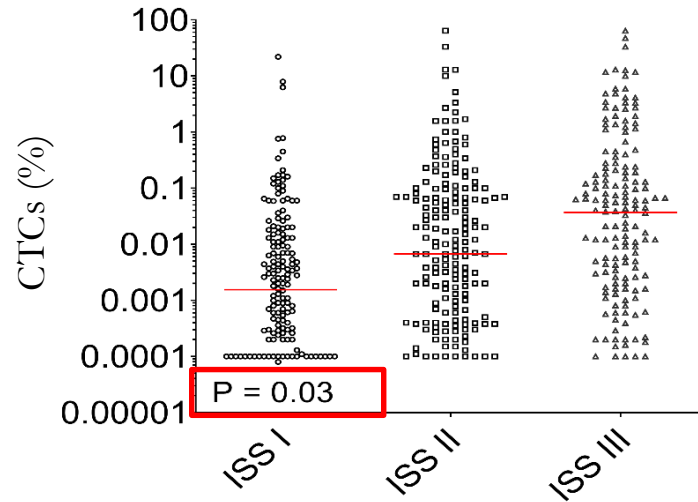
*Phenotypic disagreement in
73/493 samples (14.8%)*



Patients with phenotypic inconsistency



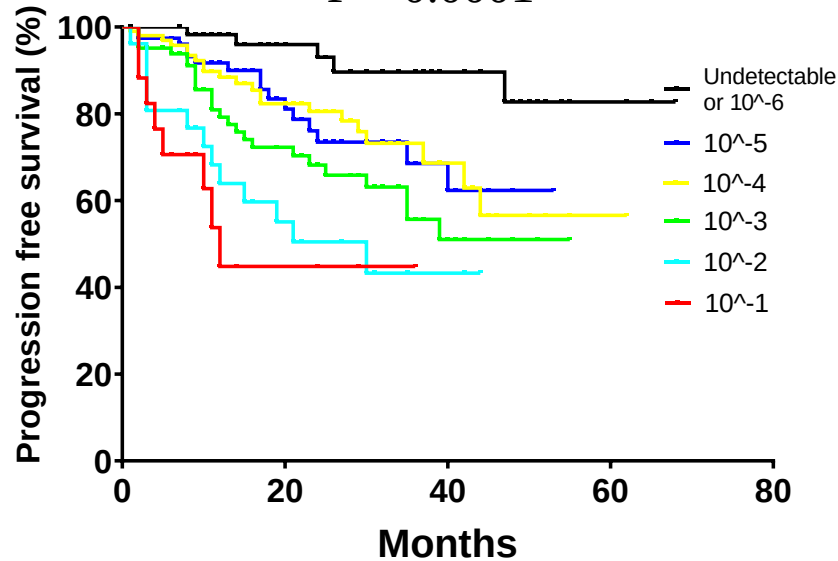
Higher levels of CTCs correlate with adverse clinical characteristics



Prognostic impact per CTCs log reduction

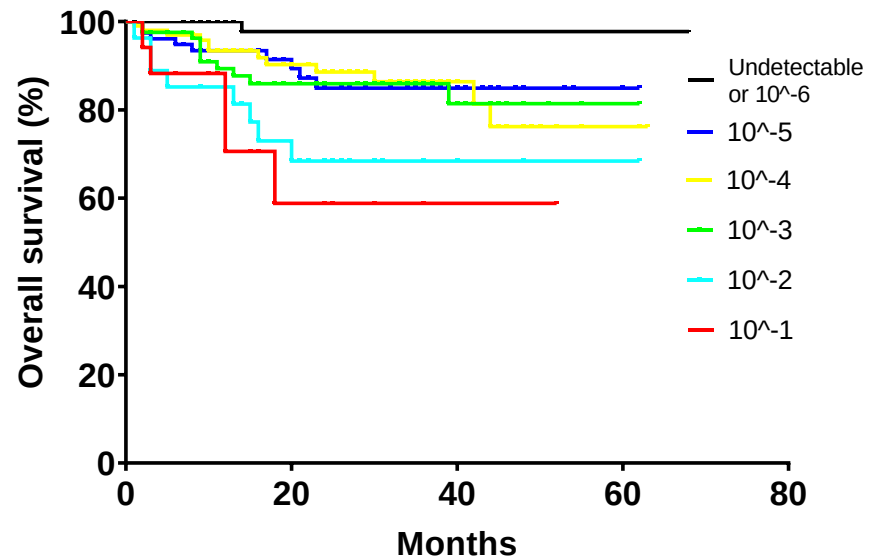
PFS

$P < 0.0001$

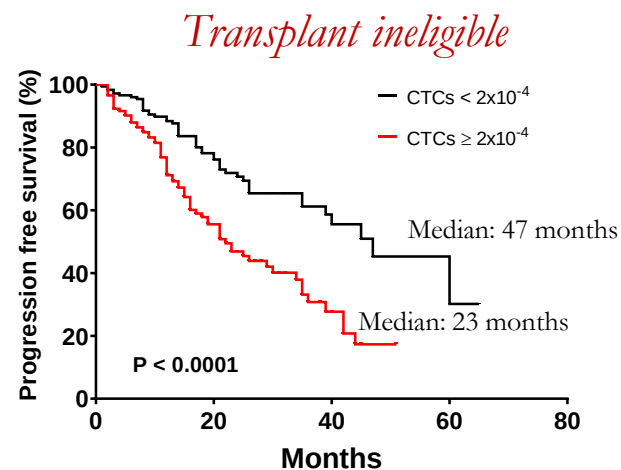
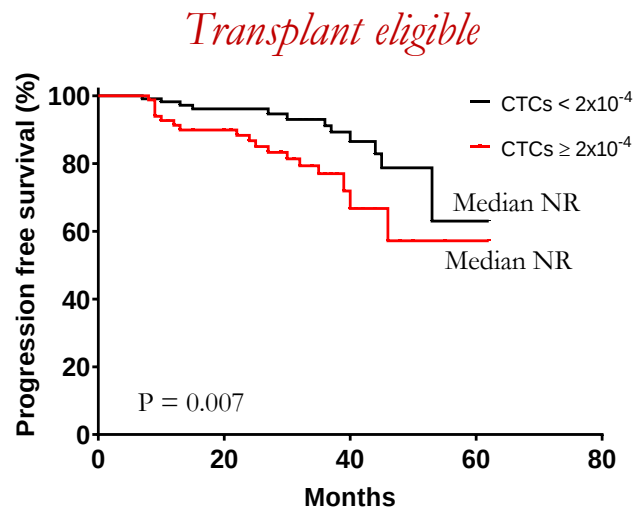
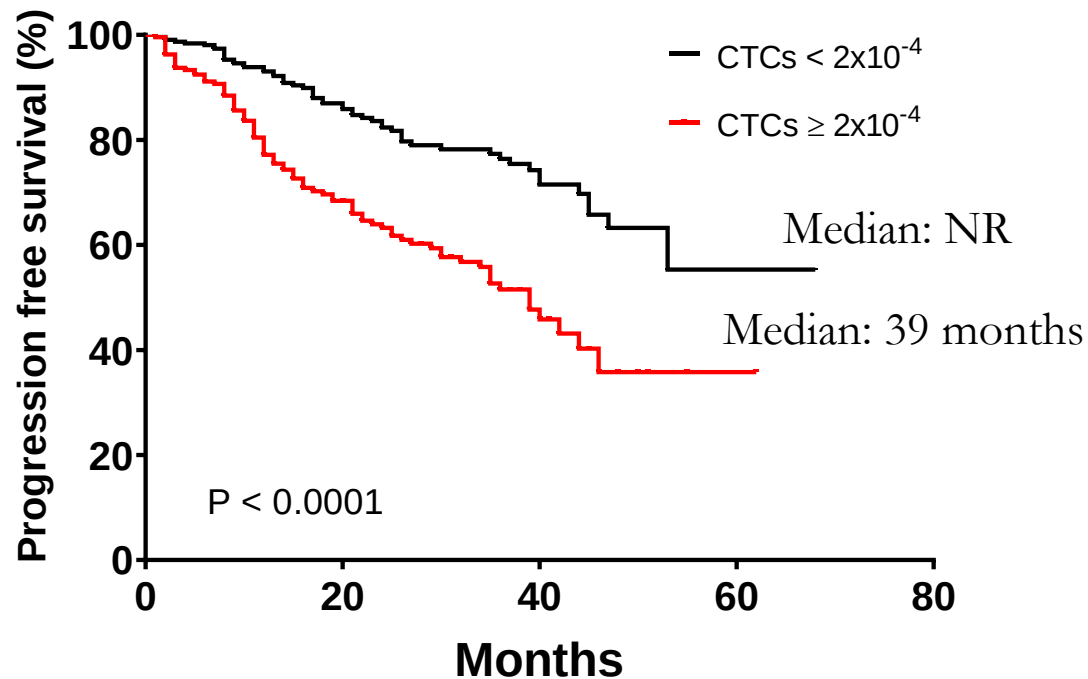


OS

$P = 0.001$



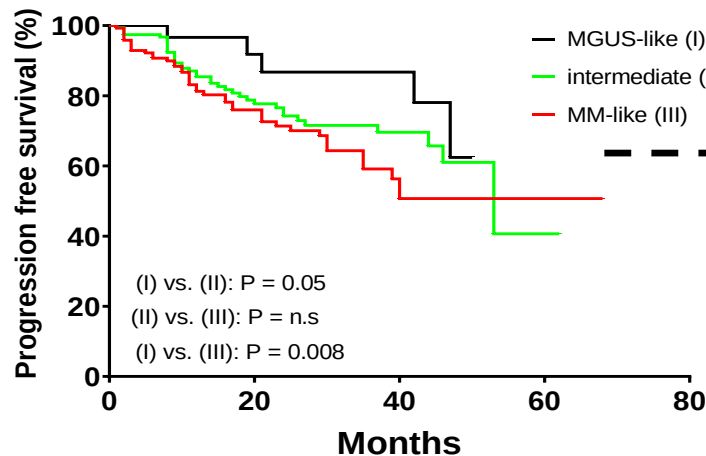
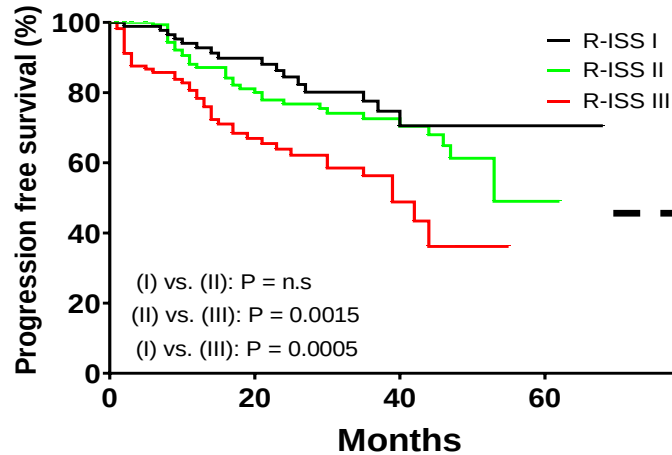
CTCs > 0.02% was the strongest INDEPENDENT predictor for reduced PFS (common in TE & TI)



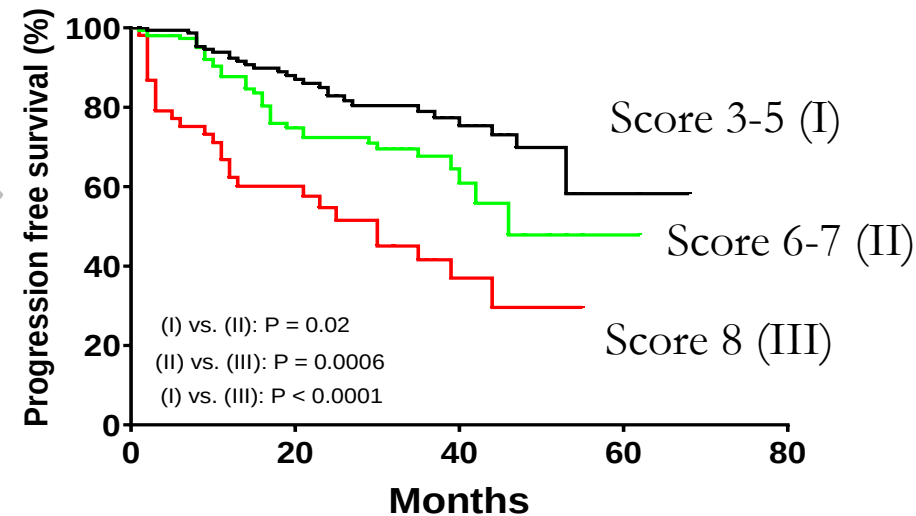
CTCs can improve MM patients' stratification system

Proposed model

R-ISS	Score	Phenotypic classification*	Score	CTCs	Score	Total Min
I	1	MGUS-like	1	< 0,02%	1	3
II	2	Intermediate	2	≥ 0,02%	2	Total Max
III	3	MM-like	3			8



+ CTCs



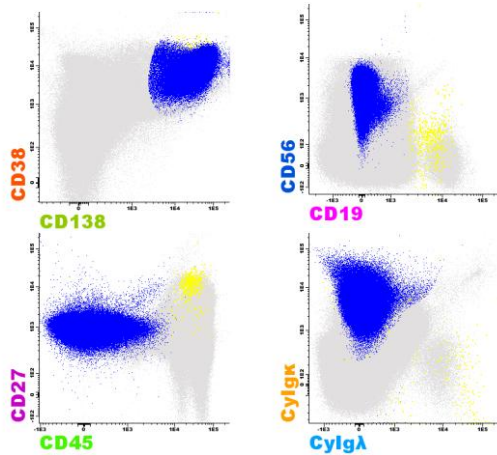
EUMELEIA clinical trial (patient 002-2)

Screening

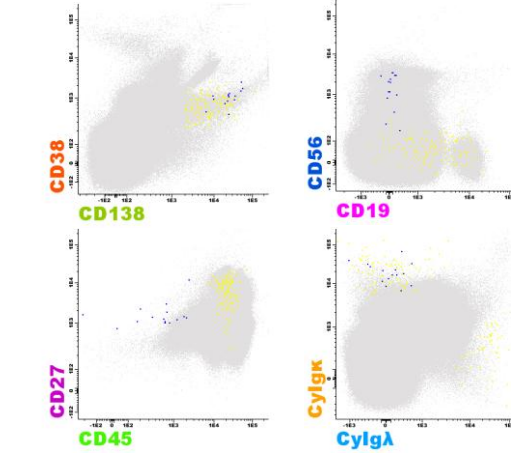
C2



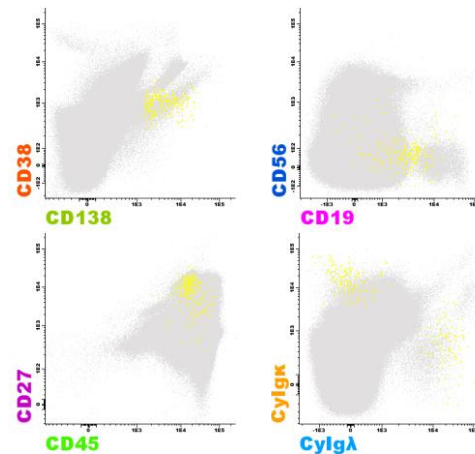
Level of CTCs
 10^{-2}



Level of CTCs
 10^{-6}



C3

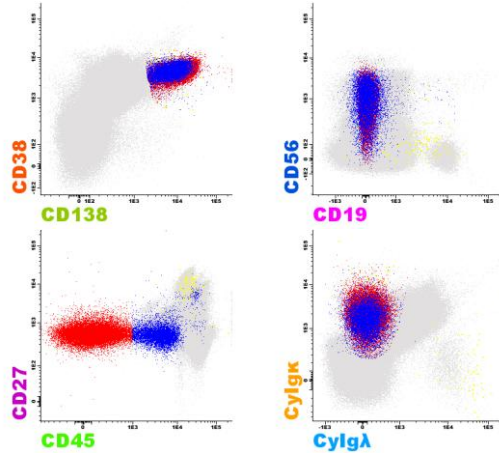


EUMELEIA clinical trial (patient 001-1)

C1

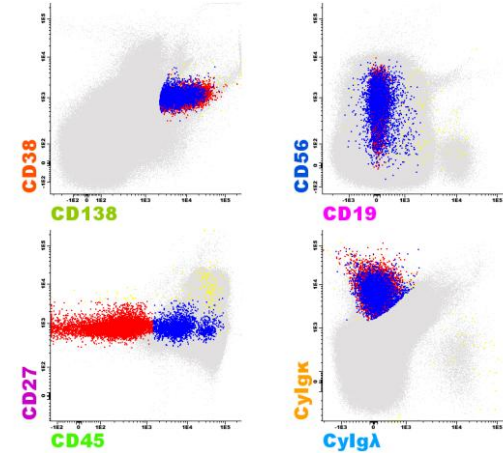
C2

Level of CTCs
 10^{-2}



- 1) $CD38^{dim} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{-} CD27^{dim/-}$ kappa (60%)
- 2) $CD38^{dim} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{dim/+} CD27^{dim/-}$ kappa (40%)

Level of CTCs
 10^{-3}



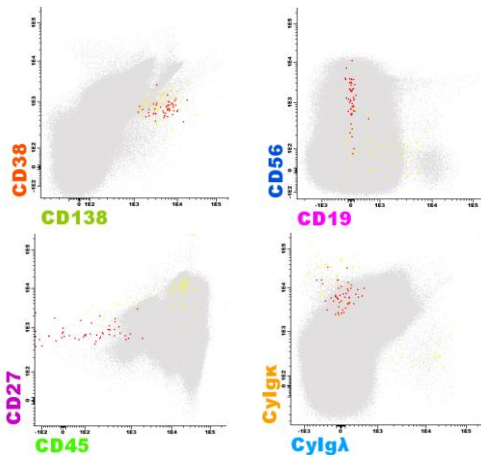
- 1) $CD38^{dim/-} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{-} CD27^{dim/-}$ kappa (74%)
- 2) $CD38^{dim/-} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{dim/+} CD27^{dim/-}$ kappa (26%)



C4

C3

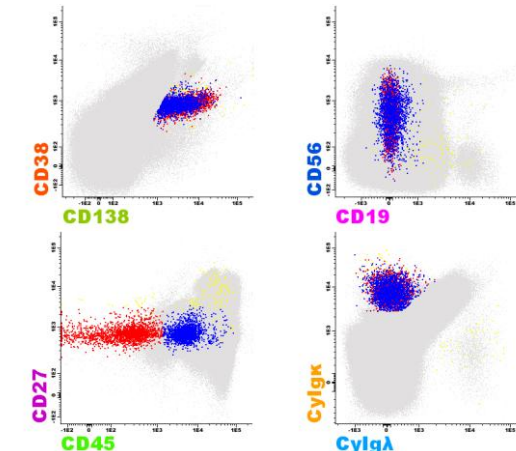
Level of CTCs
 10^{-6}



$CD38^{dim/-} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{-} CD27^{dim/-}$ kappa



Level of CTCs
 10^{-4}

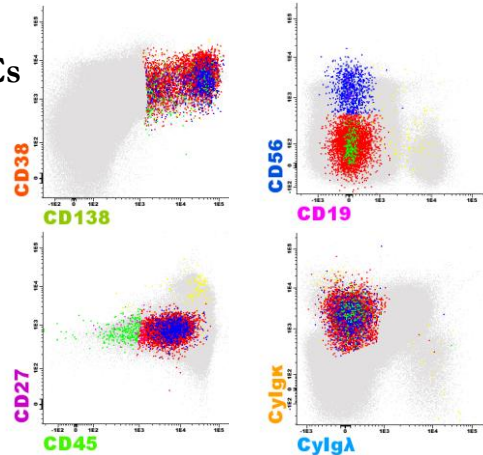


- 1) $CD38^{dim/-} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{-} CD27^{dim/-}$ kappa (56%)
- 2) $CD38^{dim/-} CD138^{++} CD19^{-} CD56^{dim/+} CD45^{dim/+} CD27^{dim/-}$ kappa (44%)

EUMELEIA clinical trial (patient 003-1)

C1

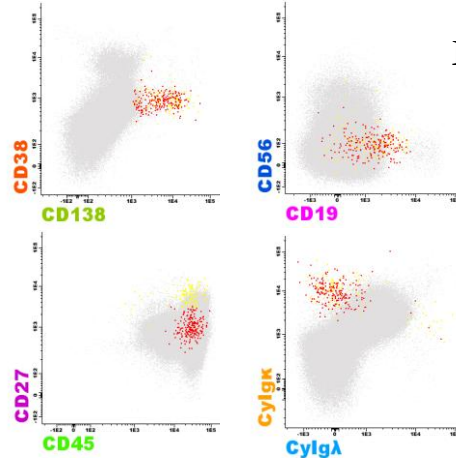
Level of CTCs
 10^{-3}



- 1) CD38⁺dim CD138⁺⁺ CD19⁻ CD56⁻ CD45⁺ CD27^{dim} kappa (94,0%)
- 2) CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁺ CD45⁺ CD27^{dim} kappa (5,8%)
- 3) CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁻ CD45⁻ CD27^{dim} kappa (0,2%)
- 4) CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁺ CD45⁻ CD27^{dim} kappa (0,01%)

C2

Level of CTCs
 10^{-4}



CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁻ CD45⁺ CD27^{dim} kappa



C3

C4

C5

C6

C7

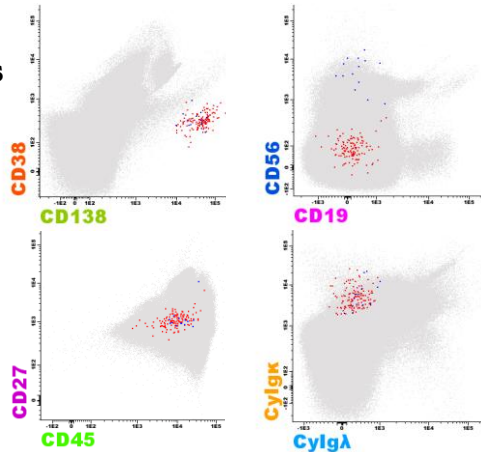
C8

C9

Undetectable
CTCs

C11

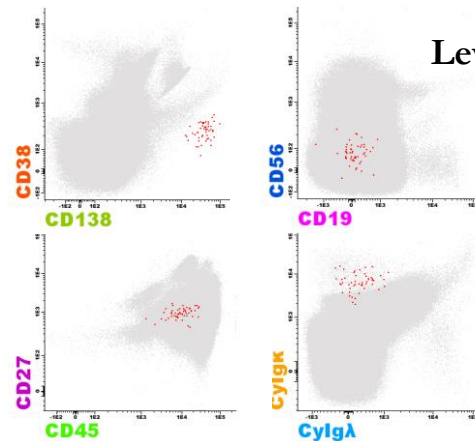
Level of CTCs
 10^{-5}



- 1) CD38⁻ CD138⁺⁺ CD19⁻ CD56⁻ CD45⁺ CD27^{dim} kappa (90,0%)
- 2) CD38⁻ CD138⁺⁺ CD19⁻ CD56⁺ CD45⁺ CD27^{dim} kappa (10,0%)

C10

Level of CTCs
 10^{-6}



CD38⁻ CD138⁺⁺ CD19⁻ CD56⁻ CD45⁺ CD27^{dim} kappa

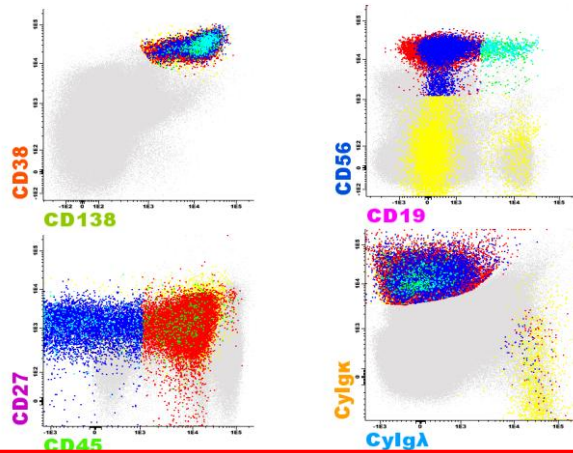


Unpublished data, pls do not copy

Clonal evolution (BM)



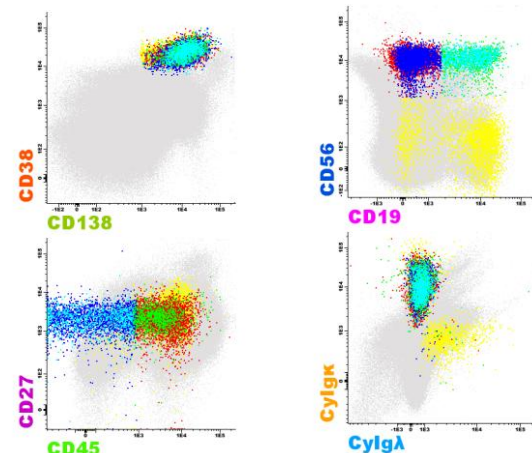
Diagnosis



- 1) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (71%)
- 2) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27⁺ kappa (25%)
- 3) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (2%)
- 4) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45⁻ CD27⁺ kappa (2%)



MRD

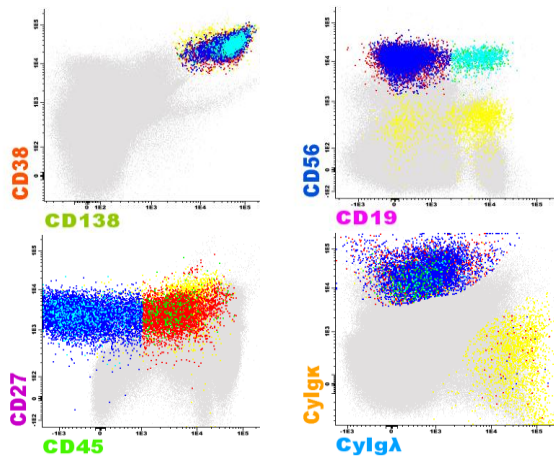


- 1) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (42%)
- 2) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27⁺ kappa (33%)
- 3) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (13%)
- 4) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45⁻ CD27⁺ kappa (12%)

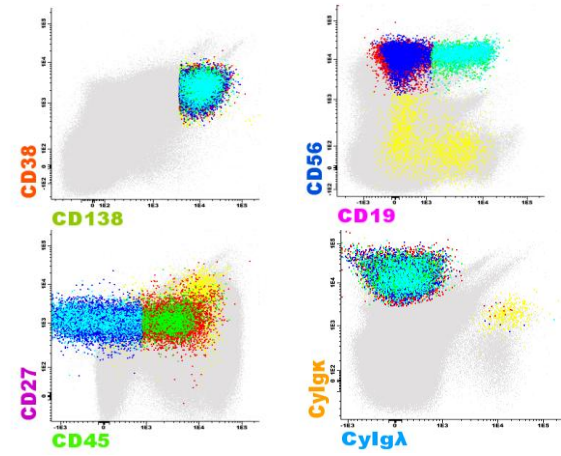
Daratumumab



Relapse



- 1) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27⁺ kappa (69%)
- 2) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (24%)
- 3) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45⁻ CD27⁺ kappa (5%)
- 4) CD38⁺ CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (2%)

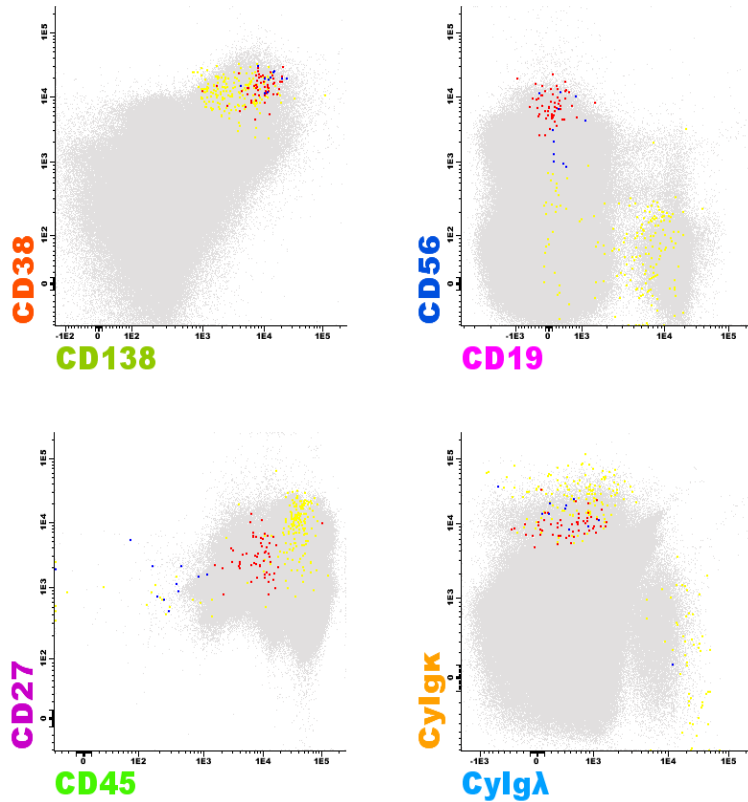


- 1) CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27^{dim} kappa (40%)
- 2) CD38^{dim} CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27^{dim} kappa (39%)
- 3) CD38^{dim} CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45⁻ CD27^{dim} kappa (12%)
- 4) CD38^{dim} CD138⁺⁺ CD19⁺ CD56⁺⁺ CD45^{+/dim} CD27^{dim} kappa (9%)

MRD

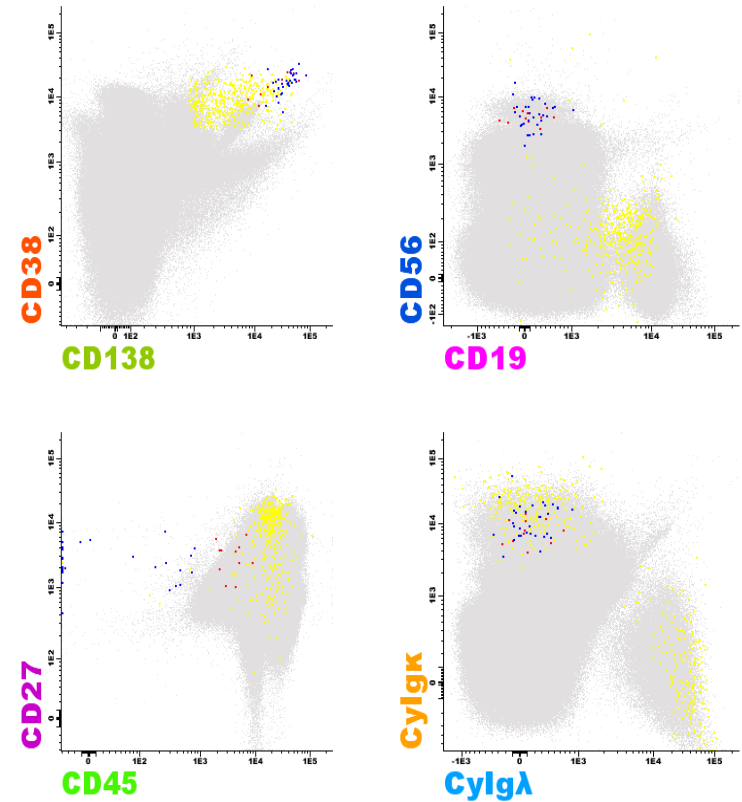
Clonal evolution in PB (CTCs)

At diagnosis



- 1) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (84%)
- 2) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27⁺ kappa (16%)

At relapse



- 1) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45^{+/dim} CD27⁺ kappa (26%)
- 2) CD38⁺ CD138⁺⁺ CD19⁻ CD56⁺⁺ CD45⁻ CD27⁺ kappa (74%)

To conclude:

- The identification of **novel biomarkers** with strong predictive/prognostic value is of utmost necessity in MM & plasma cell dyscrasias due to their apparent clinical heterogeneity features
- **MRD monitoring in MM** is clinically meaningful (level $\sim 10^6$)
 - ✓ new definition of CR including MRD negativity
 - ✓ **NGF complements** other cellular (eg. NGS) and imaging (eg. PET/CT) techniques
- **CTCs** are of clinical significance as a **biomarker**
 - ✓ CTC levels $>0.02\%$ correlated with adverse prognosis in NDMM patients
- **The BM & PB microenvironment** is likely associated with and/or may impact on response-to-treatment and prognosis of overall survival
- **FOCUS:** on the biology of clonal cells & the role of immune surveillance after therapy (including pts in $<CR$)

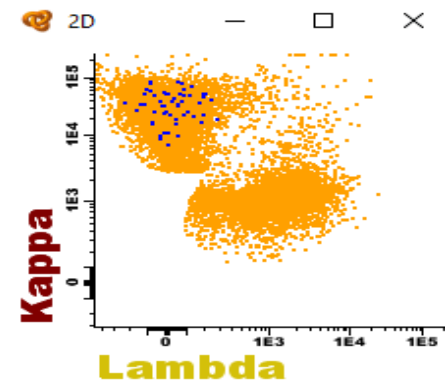
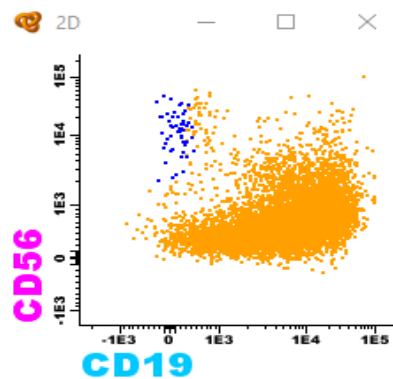


Σας ευχαριστώ

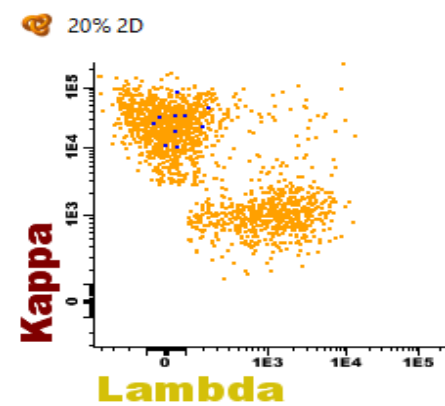
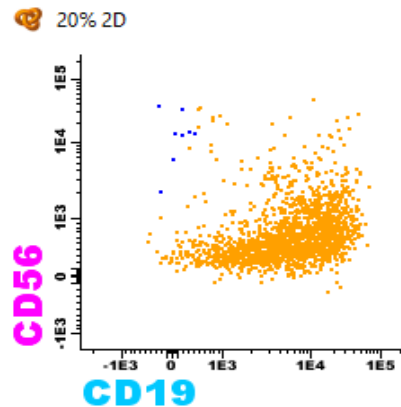
NGF: Cell Acquisition = 5×10^6 / tube

Aberrant plasma cells; Normal plasma cells

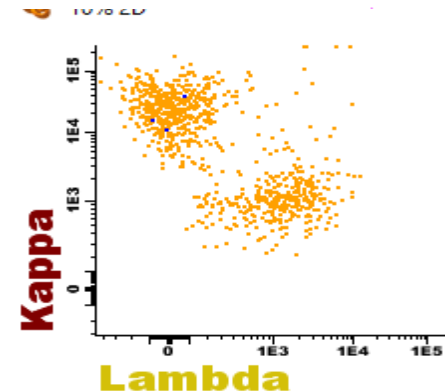
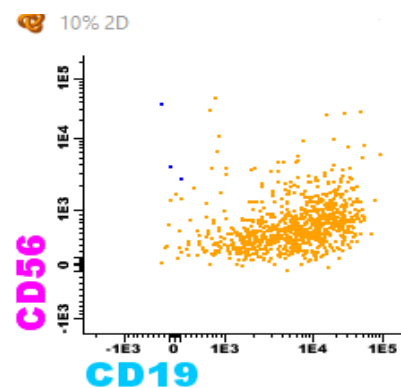
50 clonal cells detected
5,000,000 cells recorded



8 clonal cells detected
1,000,000 cells recorded



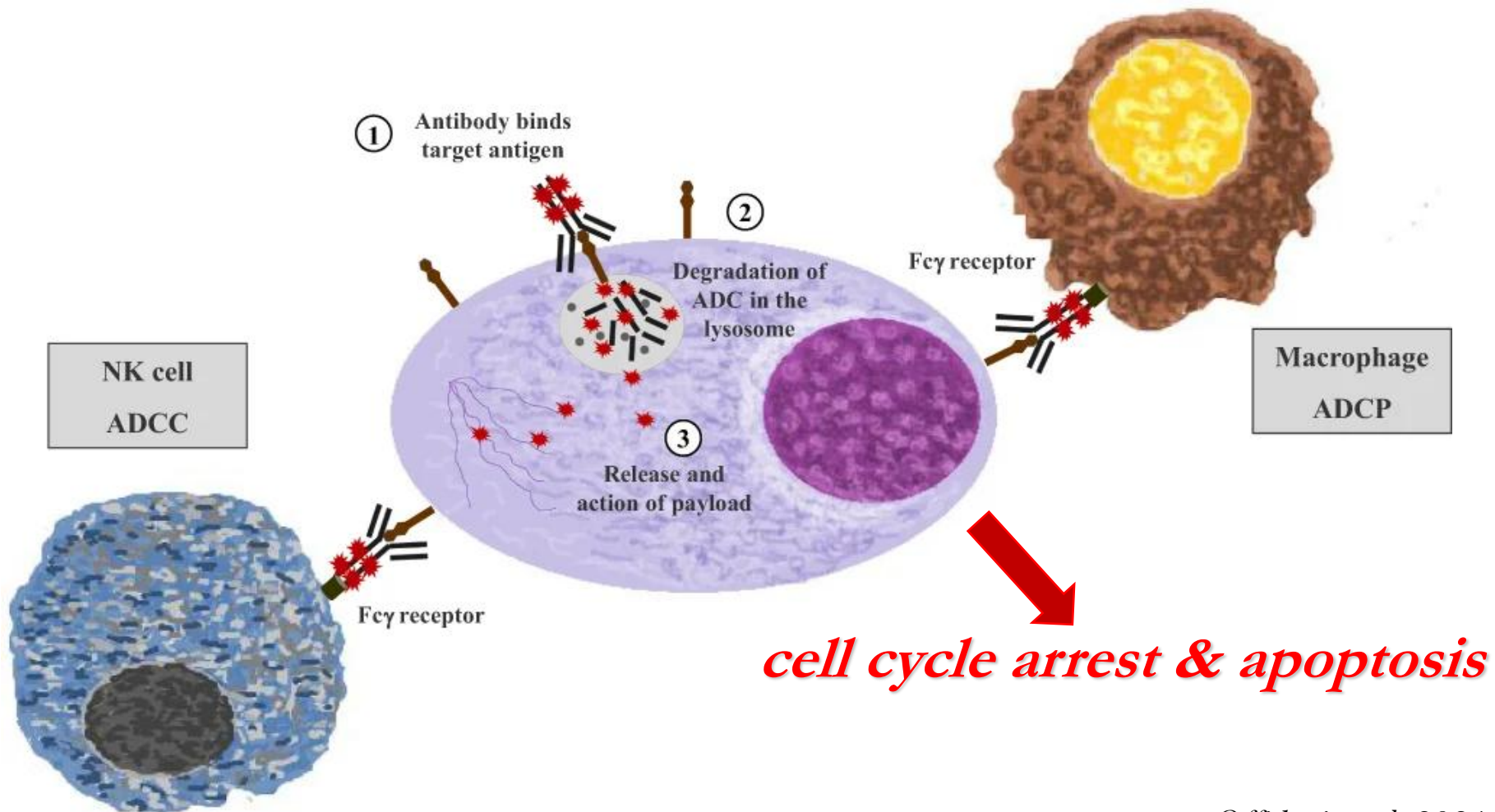
3 clonal cells detected
500,000 cells recorded



ADC Belamaf in multiple myeloma

Belantamab mafodotin

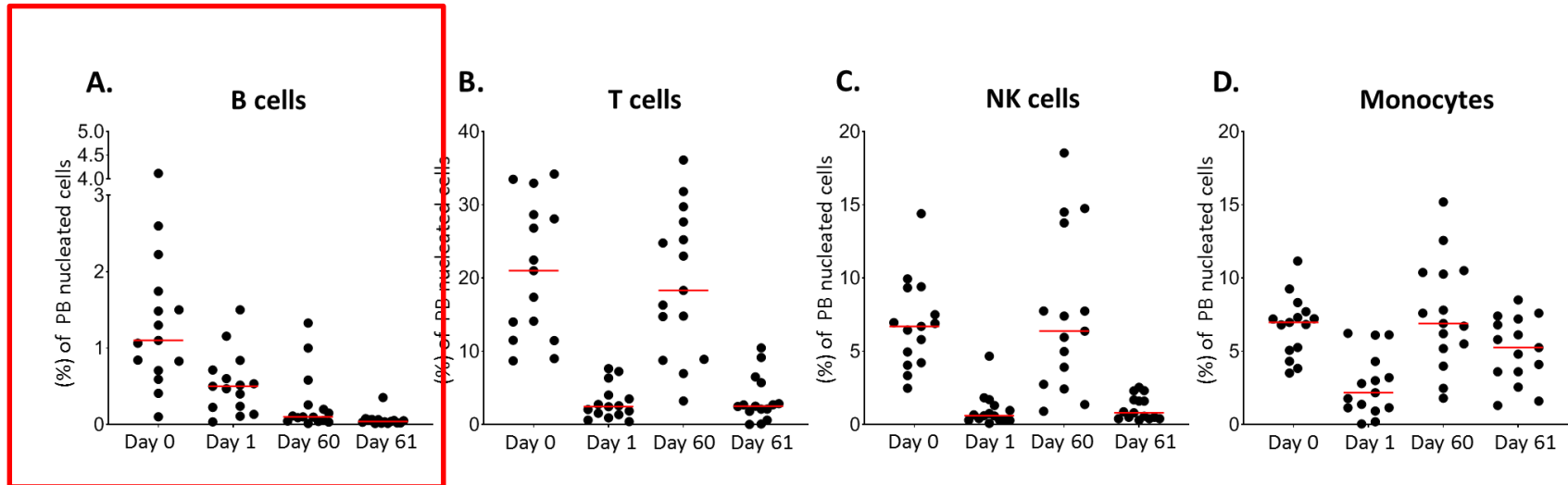
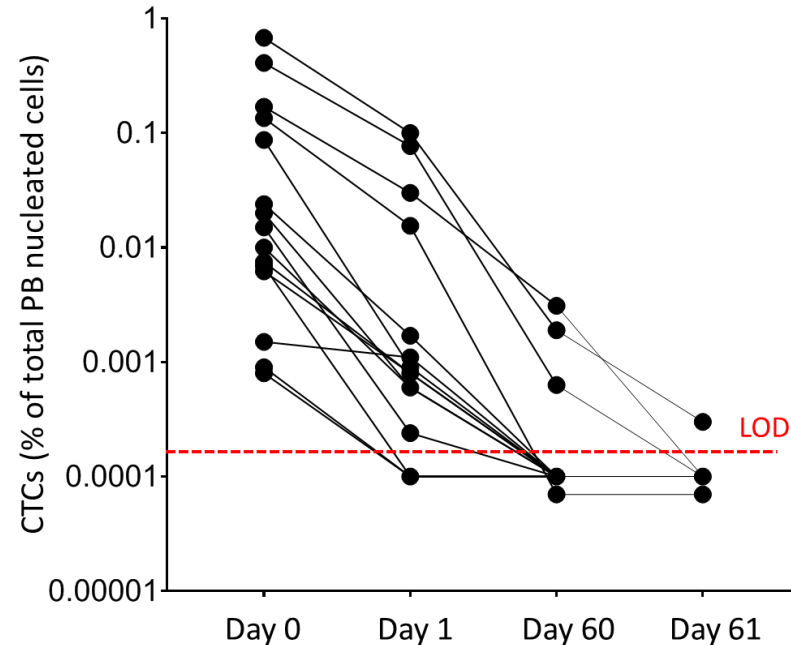
- IgG1; target BCMA; payload monomethyl auristatin F (microtubule inhibitor)
- direct killing of MM cells & ADCC & ADCP & **ICD**



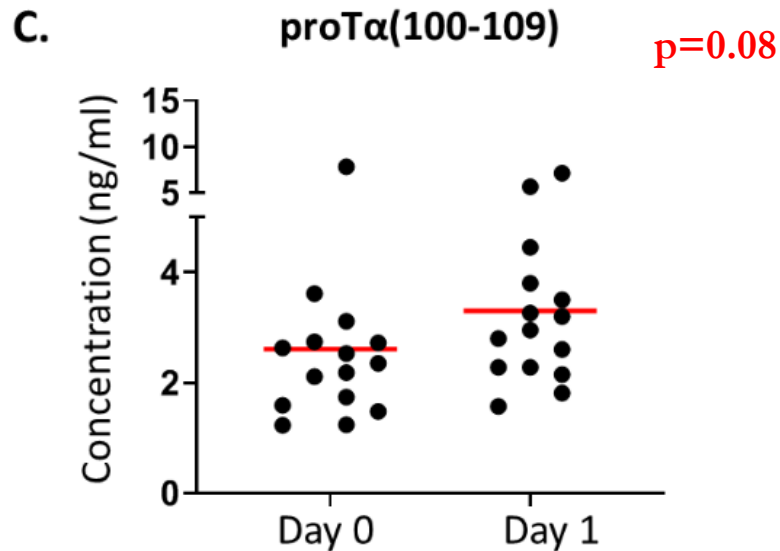
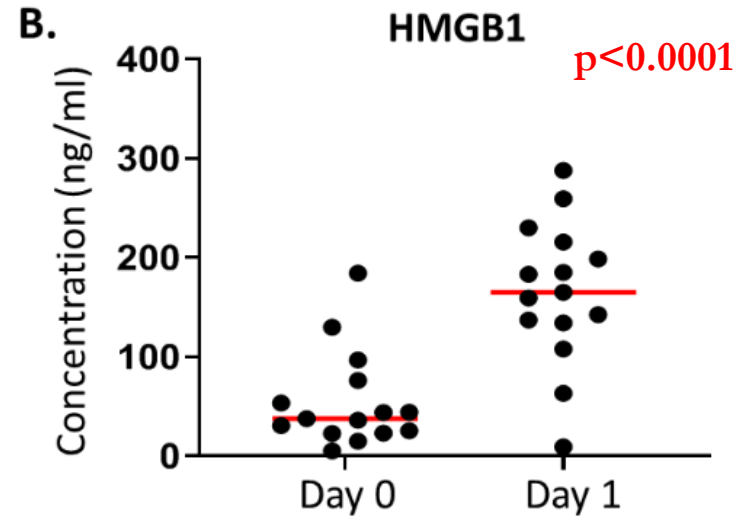
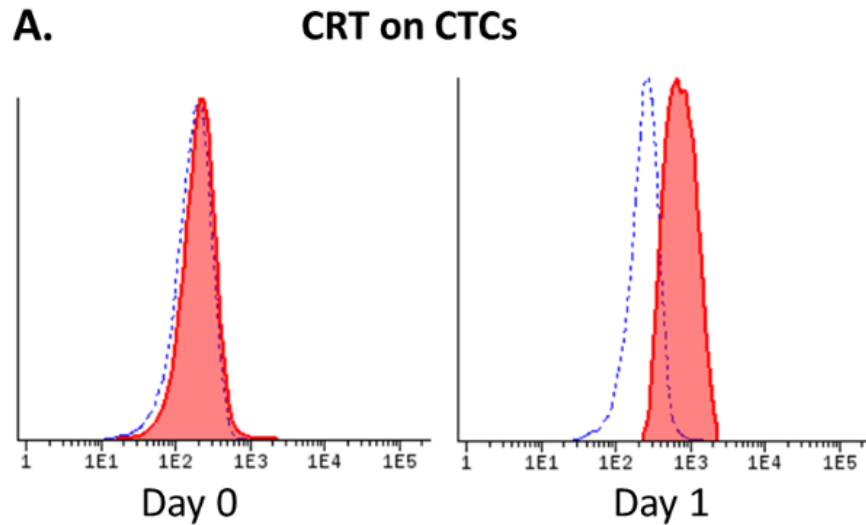
Belamaf efficiently eliminates MM cells (I)



- 15 newly diagnosed MM patients
- 2 cycles; 3-month response

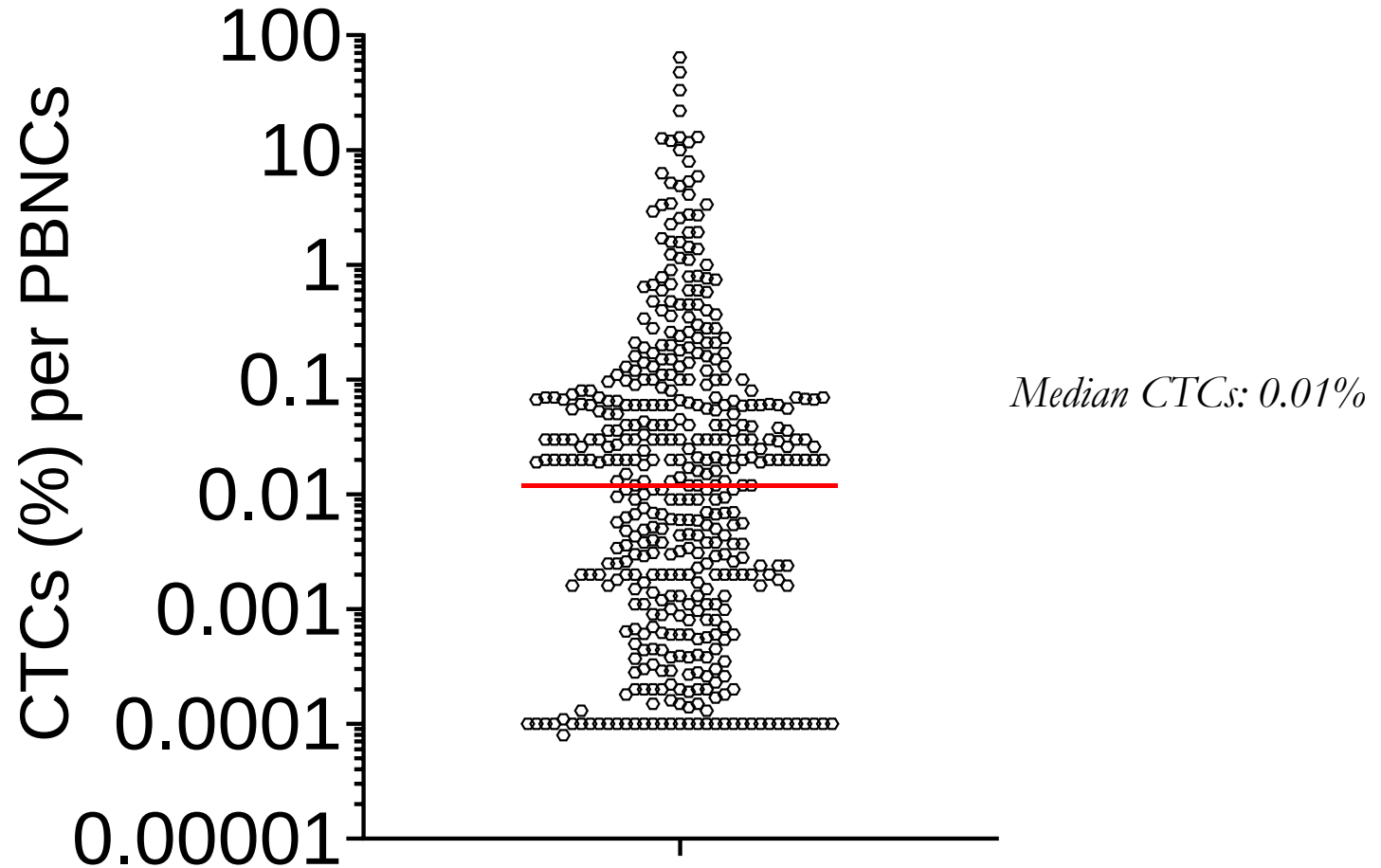


Belamaf induces ICD (II)



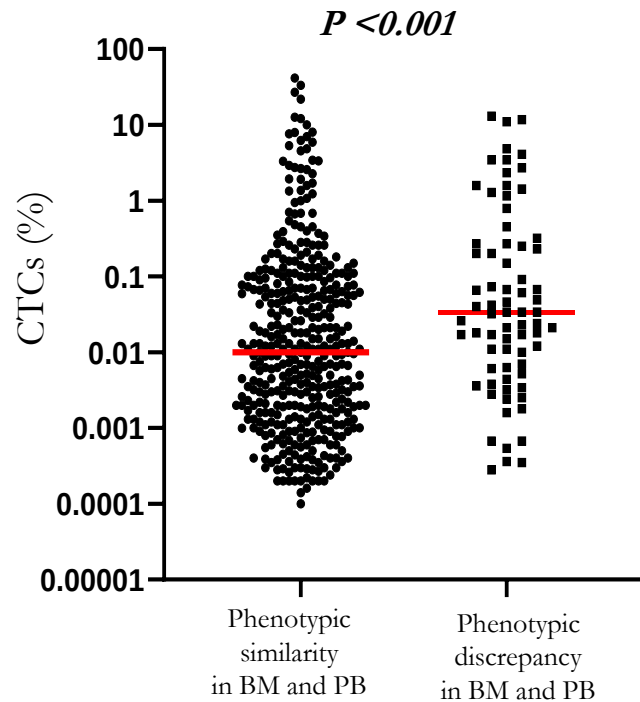
Frequency of CTCs in NDMM patients

Detection of CTCs in 493/550 samples (89.1%)



Phenotypic comparison of BM MM cells & CTCs

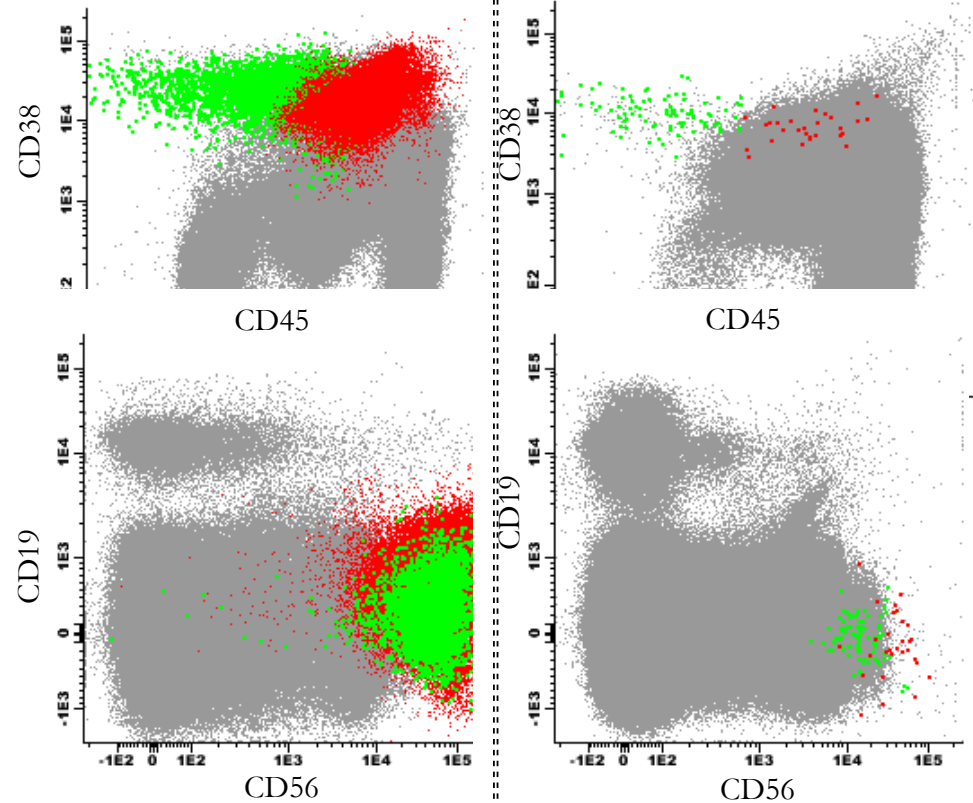
Phenotypic disagreement in
73/493 samples (14.8%)



Patient # 67

Bone marrow

Peripheral blood



APCs: 3,1% of nucleated cells

Clone 1 : 86% (CD45+) of APCs

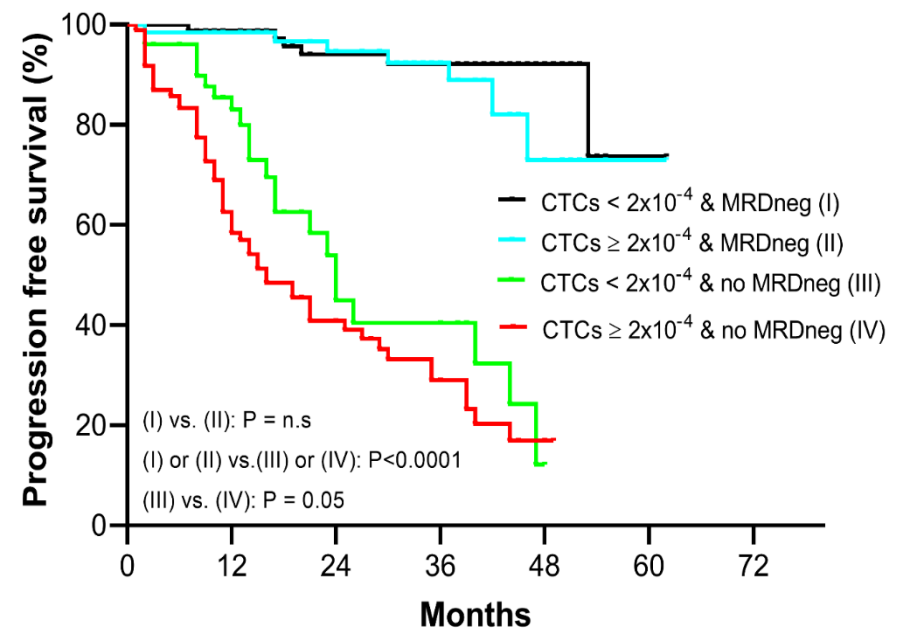
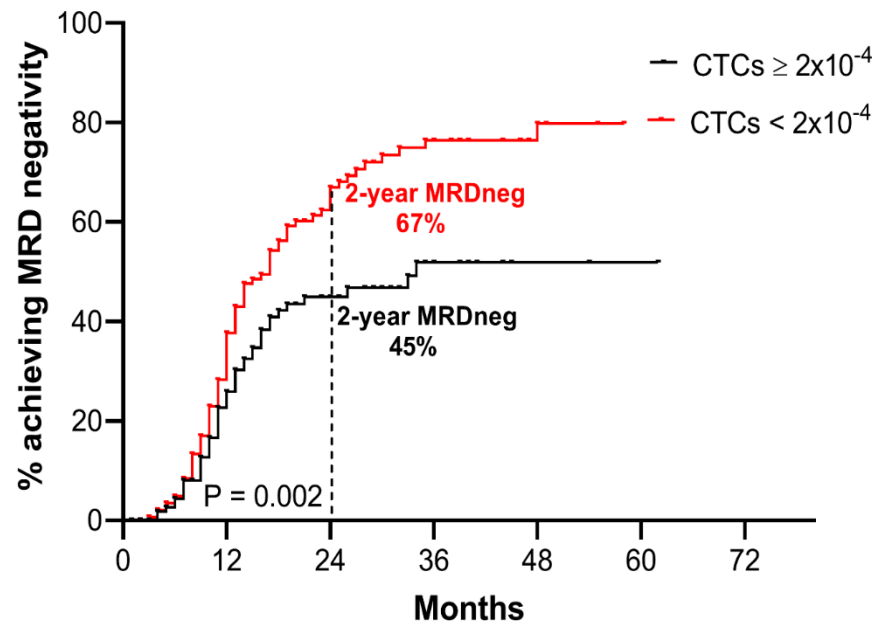
Clone 2 : 14% (CD45-) of APCs

CTCs: 0,005% of nucleated cells

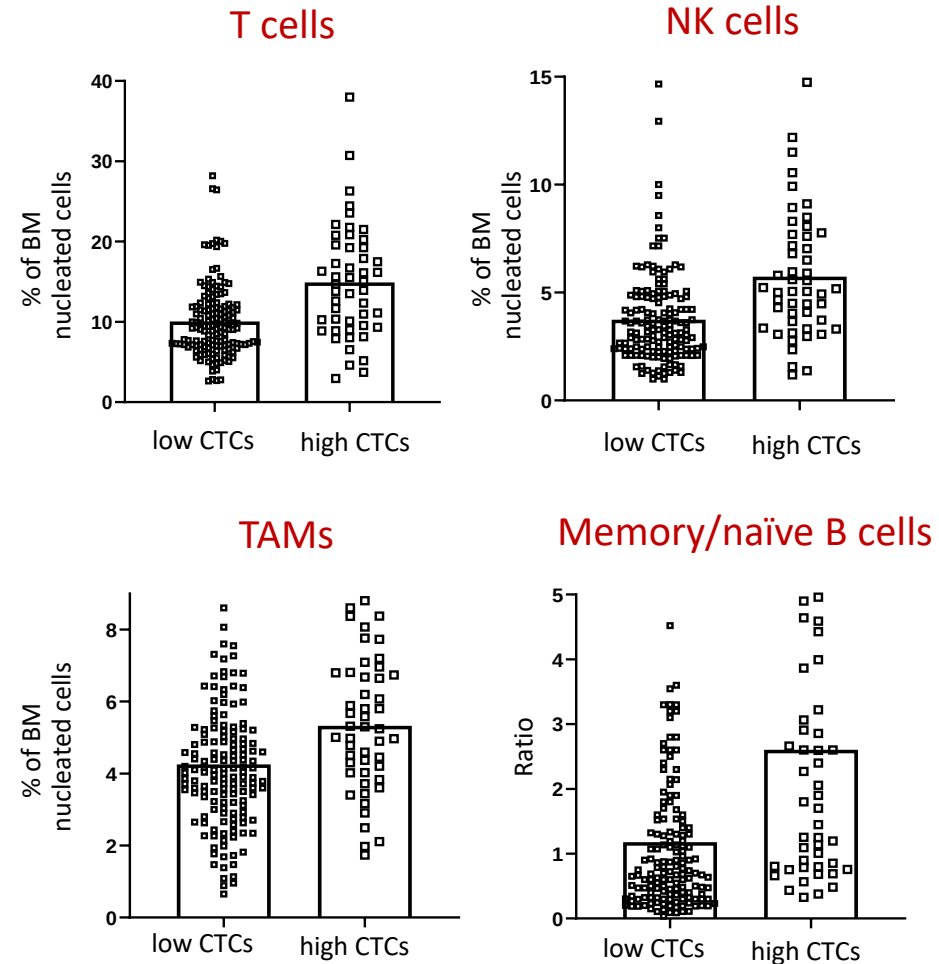
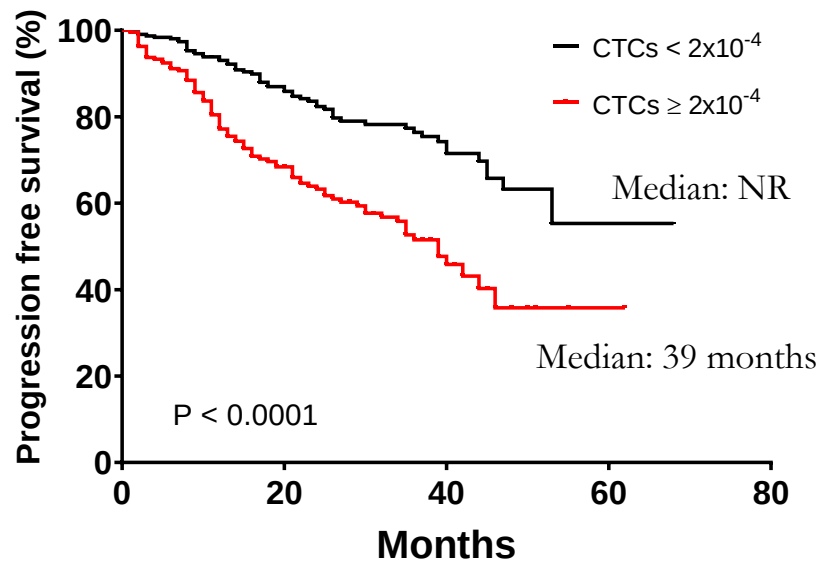
Clone 1 : 18% (CD45+) of CTCs

Clone 2 : 82% (CD45-) of CTCs

MRD negativity is less frequent in patients with high CTCs but can overcome unfavorable baseline prognostication



CTCs as prognostic determinants in NDMM patients



In all cases $P < 0.001$

Signatures in BM & PB of MM pts

Open Access

Article

Deep Phenotyping Reveals Distinct Immune Signatures Correlating with Prognostication, Treatment Responses, and MRD Status in Multiple Myeloma

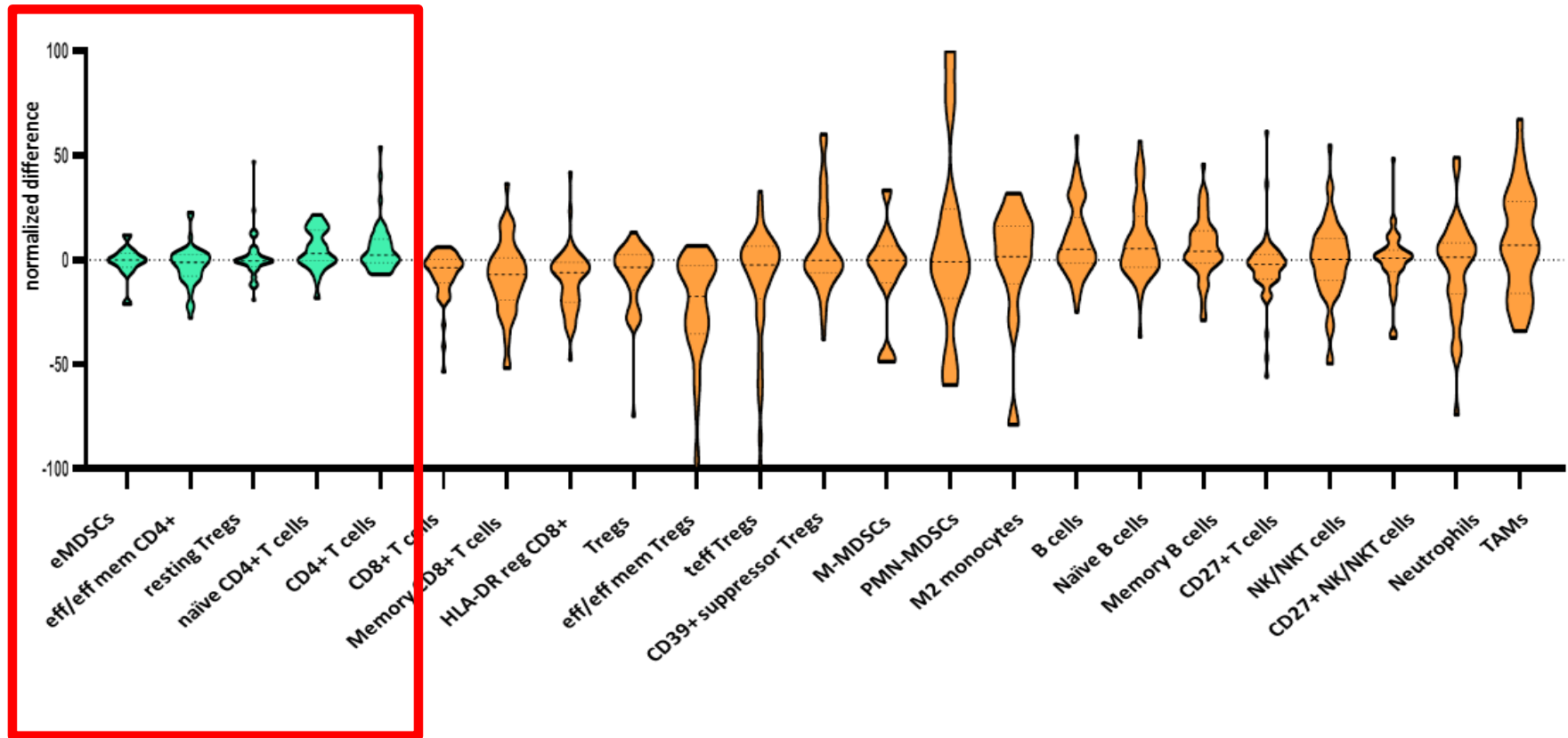


	FITC	PE	PerCP Cy5.5	BV421	BV510	PE-Cy7	APC	APC-Cy7
T cell panel 1	CD3	FoxP3	CD8	CD39	Ki67	CD45RA	CD25	CD4
T cell panel 2	CD3	FoxP3	CD45RO	CTLA-4	CD127	HLA-DR	CD25	CD4
MDSCs panel	CD14	HLA-DR	7-AAD	CD124	CD33	CD11b	CD15	CD3/19/56
MRD NGF panel	CD38	CD56	CD45	CD138	CD27	CD19	CD117	CD81

*32 distinct immune populations
in matched BM & PB samples*

Immune Subset	Expression of Markers
NGF MRD panel	
Plasma cells	CD38 ^{br} CD138+
B cells	CD19+CD45+
Naïve B cells	CD19+CD27-CD38 ⁻ /dimCD45+SSC ^{low}
B cell precursors	CD19+CD27-CD38 ^{br} CD45 ^{dim} SSC ^{low}
Memory B cells	CD19+CD27+CD38 ⁻ /dimCD45+SSC ^{low}
T cells	CD19-CD45+CD56-SSC ^{low}
CD27+ T cells	CD19-CD45+CD56-CD27+SSC ^{low}
NK/NKT cells	CD19-CD45+ CD56-SSC ^{low}
CD27+ NK/NKT cells	CD19-CD45+CD56-CD27+SSC ^{low}
Neutrophils	CD45 ^{dim} SSC ^{high}
Myeloid progenitors	CD38+CD45 ^{dim} CD117+SSC ^{high}
Monocytes—TAMs	CD38+CD45+CD81+SSC ^{int}
Mast cells	CD45 ^{dim} CD117 ^{br}
Erythroblasts	CD38-CD45-SSC ^{low}
Erythroid progenitors	CD38 ⁻ /dimCD45 ⁻ /dimCD117+SSC ^{low}
T cells panel	
T regulatory cells (Tregs)	CD3+CD4+CD25+CD127 ^{low} FoxP3+
Effector/effector memory Tregs	CD3+CD4+CD25+CD127 ^{low} FoxP3+CD45RA-CD45RO+HLA-DR-CTLA4+
Terminal effector Tregs (teff Tregs)	CD3+CD4+CD25+CD127 ^{low} FoxP3+CD45RA-CD45RO+HLA-DR+ CTLA4+
Resting Tregs	CD3+CD4+CD25+CD127 ^{low} FoxP3+CD45RA-CD45RO+HLA-DR-CTLA4-
CD39+ suppressor Tregs	CD3+CD4+CD25+CD127 ^{low} FoxP3+CD45RA-CD45RO+CD39+
CD4+ T cells	CD3+CD4+
Naïve CD4+ T cells	CD3+CD4+CD45RA+CD45RO-
Effector/Effector memory CD4+T cells	CD3+CD4+CD45RA-CD45RO+
CD8+ T cells	CD3+CD8+
CD8+ Tregs	CD3+CD8+CD25+FoxP3+
Memory CD8+ T cells	CD3+CD8+CD45RO+
HLA-DR regulatory CD8+ T cells	CD3+CD8+HLA-DR+
MDSCs panel	
Polymorphonuclear MDSCs (PMN-MDSCs)	CD14-CD11b+CD15+SSC ^{high}
Early MDSCs (eMDSCs)	Lin(CD3/CD14/CD15/CD19/CD56)-HLA-DR-CD33+
Monocytic MDSCs (M-MDSCs)	CD11b-CD14+HLA-DR ^{low/-} CD15-
M1 monocytes	Lin(CD3/CD14/CD15/CD19/CD56)-CD14+CD124-
M2 monocytes	Lin(CD3/CD14/CD15/CD19/CD56)-CD14+CD124+

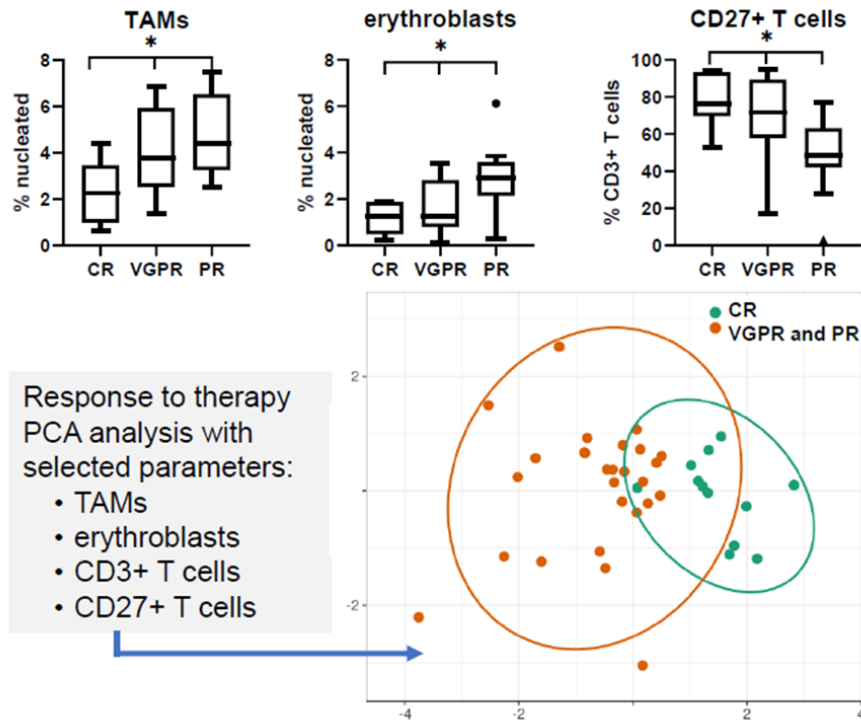
Pairwise differences in PB vs BM immune cells



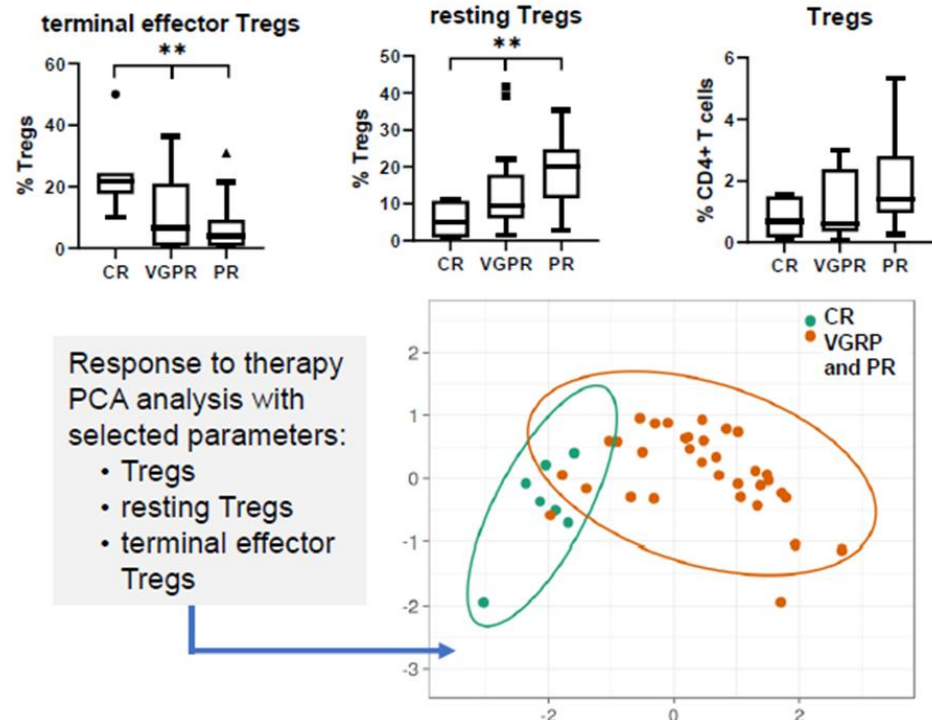
- High heterogeneity among patients
- Dissimilarities between the 2 sites (BM & PB)
- PB does not fully reflect the BM

Distinct BM and PB immune signatures correlate with different responses to the same induction therapy

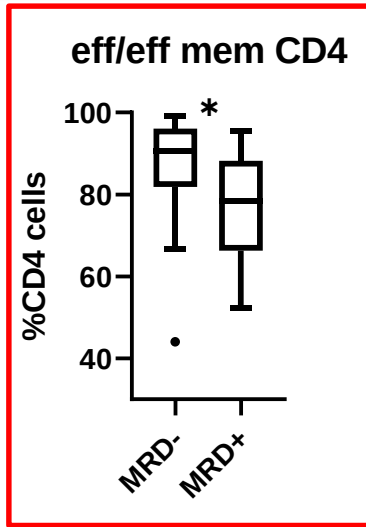
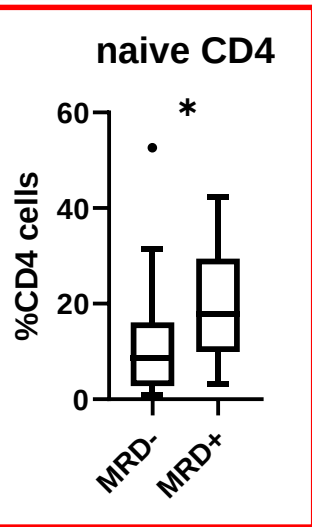
BONE MARROW



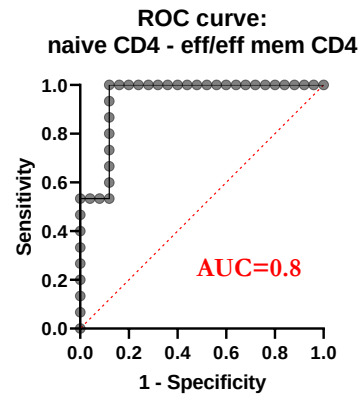
PERIPHERAL BLOOD



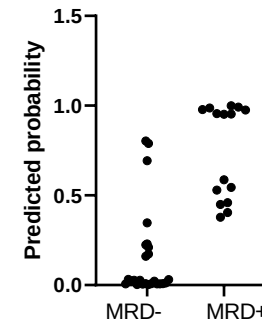
MRD scoring system based on naïve CD4+ & effector/effector memory CD4+ T cells



Selection of naïve and eff/eff mem CD4+ T cells



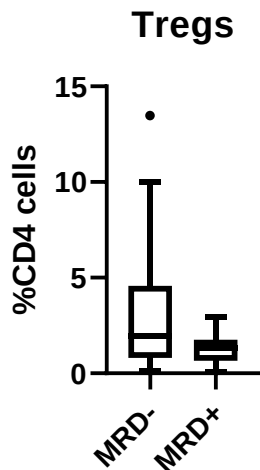
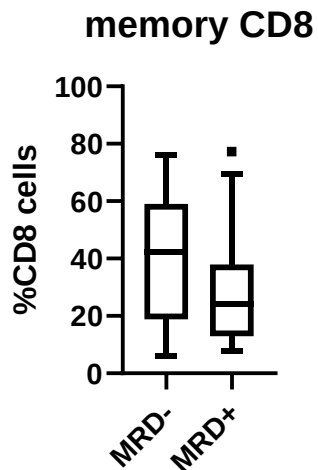
Predicted vs Observed



MRD scoring system:

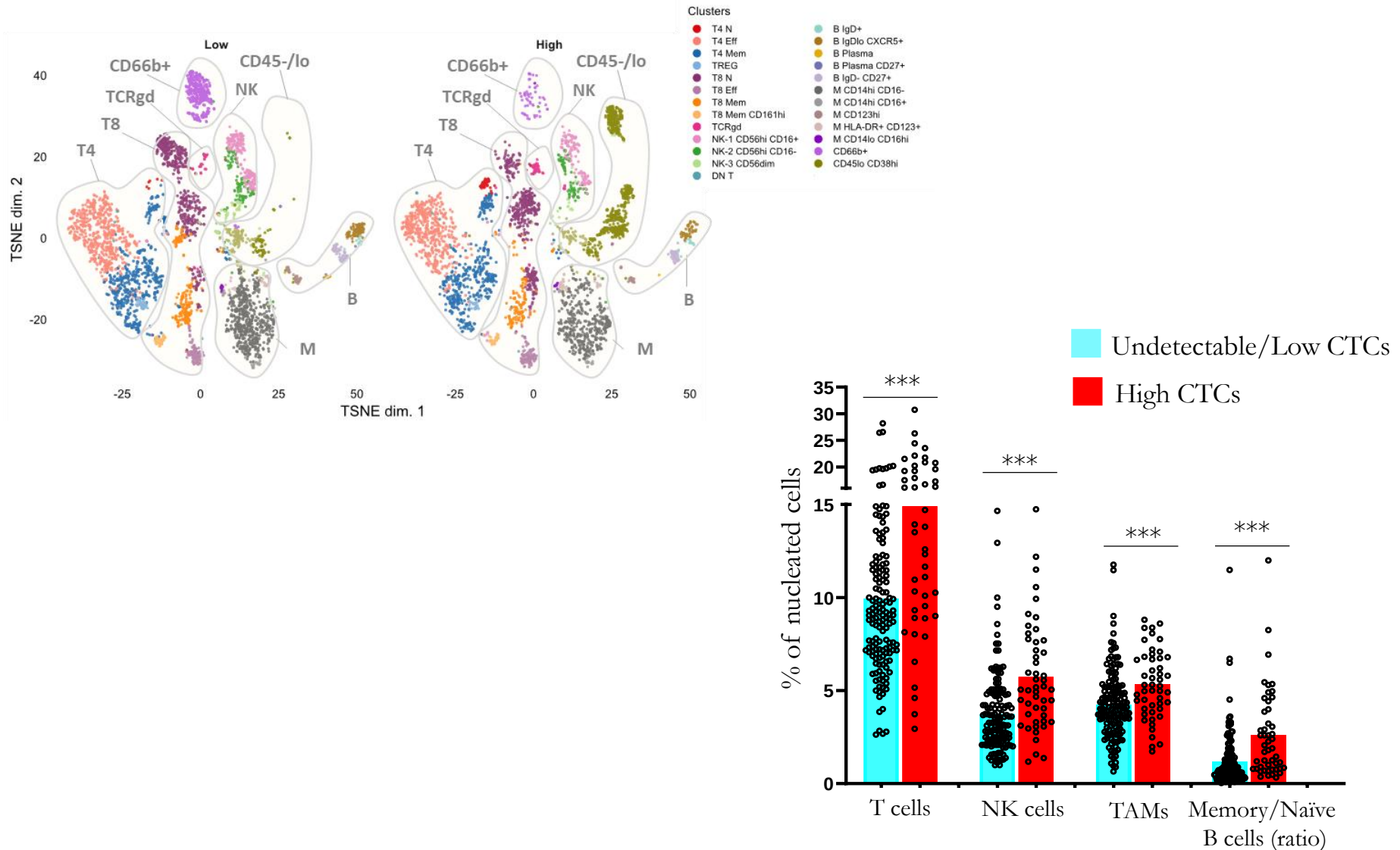
- MRD positive: naïve CD4+ T >8% and eff/eff mem CD4+ T <90%
- Otherwise MRD negative

Test performance by applying scoring system on a separate cohort of 20 MRD patients



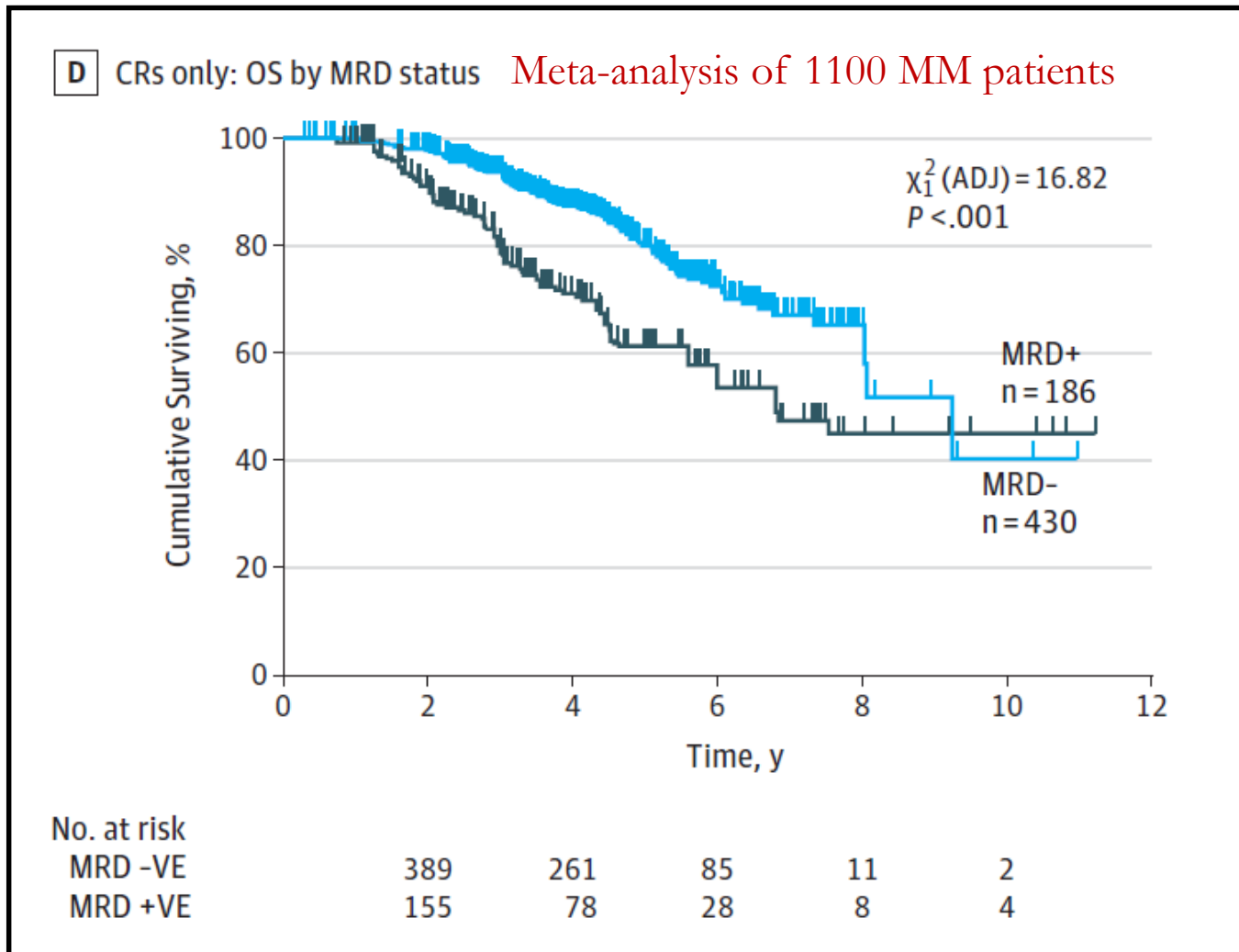
n=20	MRD+	MRD-	
Positive 8	True Positives 6	False Positives 2	PPV=TP/(TP+FP) 0.75
Negative 12	False Negatives 1	True Negatives 11	NPV=TN/(TN+FN) 0.92
Sensitivity=TP/(TP+FN) 0.86		Specificity=TN/(TN+FP) 0.85	

Different levels of CTCs correlate with distinct microenvironmental BM profiles



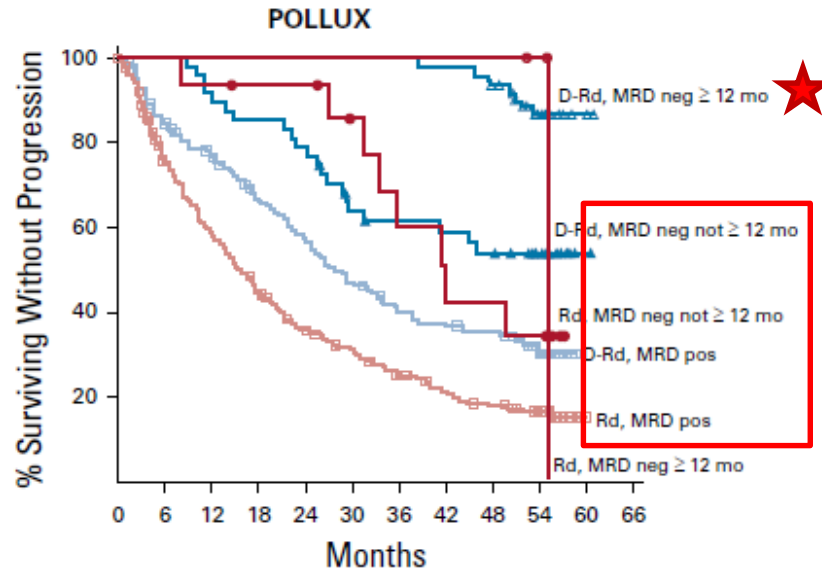
MRD is an excellent prognostic marker...

BUT: MRD-positivity not always fatal & MRD-negativity is not cure



Sustained MRD-negativity impacts on PFS

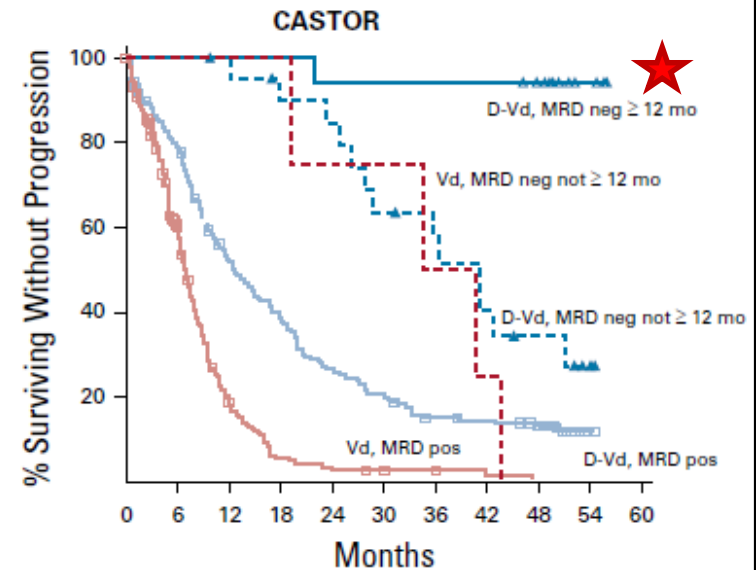
A



No. at risk

Rd, MRD neg not ≥ 12 mo	15	15	14	13	13	10	7	5	5	4	0	0
D-Rd, MRD neg not ≥ 12 mo	47	47	43	40	37	28	26	25	23	16	1	0
Rd, MRD neg ≥ 12 mo	4	4	4	4	4	4	4	4	4	3	0	0
D-Rd, MRD neg ≥ 12 mo	46	46	46	46	46	46	46	45	42	26	2	0
Rd, MRD pos	264	187	142	110	85	69	55	44	36	21	1	0
D-Rd, MRD pos	193	156	140	118	101	82	71	64	60	34	0	0

B

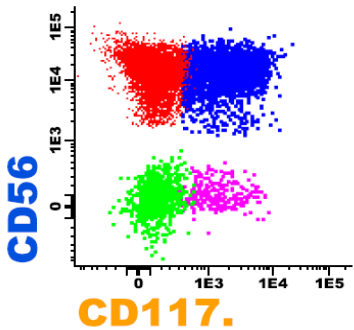
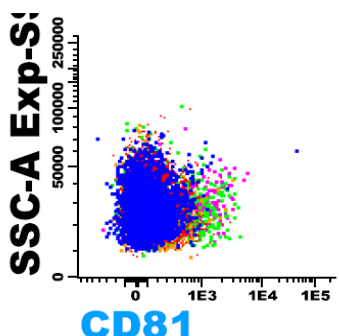
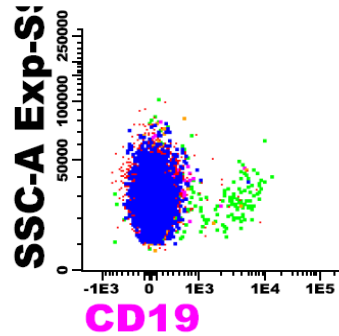
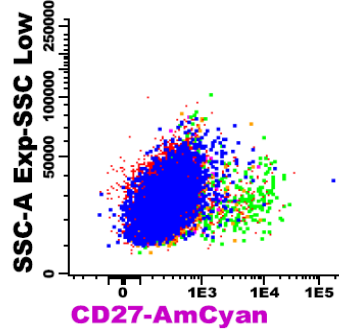
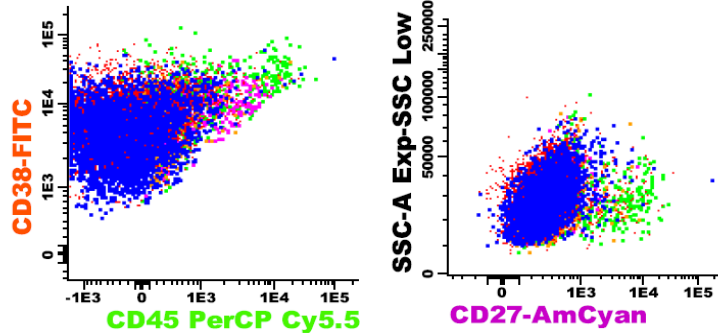


No. at risk

Vd, MRD neg not ≥ 12 mo	4	4	4	4	3	3	2	1	0	0	0
D-Vd, MRD neg not ≥ 12 mo	21	21	20	17	16	12	10	7	5	2	0
D-Vd, MRD neg ≥ 12	17	17	17	17	16	16	16	16	13	4	0
Vd, MRD pos	243	125	35	11	6	4	3	1	0	0	0
D-Vd, MRD pos	213	160	101	75	53	40	28	25	20	1	0

PFS similar between initially MRD+ pts & pts converting from MRD- to MRD+

Follow changes from diagnosis to MRD...+++



APCs1: 65,11%
APCs2: 23,82%
APCs3: 8,77%
APCs4: 2,18%

- the majority of BM diagnostic samples have >2 phenotypically distinct clonal subsets so...
- is the MRD clone identical with the main clone at diagnosis? **YES/NO**
- are there changes in the relative frequencies among clones? **YES/NO**
- are there newly appearing clones/subclones?
- why some clonal cells persist but remain under the control of the immune system?
- what changes in immune reconstitution may favor or not clonal expansion?

**UNDERSTAND *CLONAL EVOLUTION*
IN THE CONTEXT OF IMMUNE
PROFILING**